

Vascular Aging

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ABSTRACT: Vascular aging is a central determinant of healthy life span, not only influencing the susceptibility to cardiovascular diseases but also shaping the risk of systemic decline across multiple organs. It is driven by a variety of age-related factors, including cellular senescence, chronic inflammation, loss of proteostasis, mitochondrial dysfunction, genomic instability, epigenetic remodeling, and stem cell exhaustion. These processes interact with the unique mechanical and metabolic environment of the vasculature to create a distinctive pathological trajectory, manifested in part as arterial stiffening, impaired barrier integrity, and dysregulated vasomotor control. Recent advances in single-cell omics and cross-organ molecular clocks have revealed the heterogeneity and organ specificity of aging, underscoring the need for integrative frameworks that connect vascular biology with overall health. Meanwhile, the development of diverse therapeutic strategies—ranging from senolytic and immune-mediated clearance to metabolic and mitochondrial interventions—highlights the translational potential of targeting the aging vasculature. Looking ahead, multimodal biomarkers and precision medicine may transform vascular aging from an inevitable process into a modifiable determinant of health span.

Key Words: aging ■ DNA damage ■ inflammation ■ mitochondria ■ stem cells

You are as old as your arteries.

—Thomas Sydenham (1624-1689)

The links between vascular aging and overall life span and health have been known since the time of the great English physician Thomas Sydenham. However, the nature of this relationship, namely whether it indicates a correlative versus causative link, has significantly evolved over the past several decades. In large part because of our deeper understanding of how and why we age, there is a growing framework to place the age-dependent susceptibility for myriad diseases within the context of the biology of aging. This shift in understanding can be traced back to observations in basic science laboratories that noted that in simple organisms such as *Caenorhabditis elegans*, manipulating a single gene could extend the overall life span of the organism.¹⁻³ Remarkably, many of the genes and pathways identified in lower organisms are well conserved. For instance, the first well-described pathway in worms outlined a pathway encompassing an insulin-like receptor, regulating the activity of a downstream kinase (eg, PI3K), and ultimately modulating the activity of the FOXO fam-

ily of transcription factors.² The facts that genetic variants in FOXO family members appear to correlate with human longevity⁴ and, perhaps more germane to this review, these longevity effects appear particularly relevant to those individuals with cardiovascular risk factors⁵ suggest that aging is not random but rather, like all other biological processes, is under genetic and biochemical regulation. Dissecting these regulatory mechanisms thereby provides hope that a blueprint can emerge that will allow us to finally understand how to tackle the single greatest risk factor that underlies almost all major human diseases: age itself.

The characteristic phenotypes of vascular aging include deterioration of mechanical properties, disruption of barrier integrity, and dysregulation of vasomotor control (Figure 1). Vascular compliance and elasticity depend on the coordinated function of medial vascular smooth muscle cells (VSMCs) and elastic and collagen fibers. With advancing age, elastic fiber fragmentation and increased collagen deposition lead to arterial stiffening and reduced compliance. In large arteries, such alterations in mechanical properties result in a marked increase in pulse wave velocity and pulse pressure, both of which have been established as independent predictors of adverse cardiovascular outcomes.⁶

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Nonstandard Abbreviations and Acronyms

AGE	advanced glycation end-product
BBB	blood-brain barrier
CHIP	clonal hematopoiesis of indeterminate potential
COLCOT	Colchicine Cardiovascular Outcomes Trial
DAMP	damage-associated molecular pattern
EC	endothelial cell
ECM	extracellular matrix
IL	interleukin
LoDoCo2	Low-Dose Colchicine 2
NF-κB	nuclear factor- κ B
ROS	reactive oxygen species
SASP	senescence-associated secretory phenotype
VSMC	vascular smooth muscle cell

The vascular endothelium is central to barrier function. In aged endothelial cells (ECs), reduced expression of intercellular junction proteins (eg, claudins and occludins) and cytoskeletal remodeling contributes to increased permeability.⁷ Concurrently, basement membrane thickening and impaired endothelium-dependent vasodilation coexist, leading to reduced efficiency of molecular exchange along with increased nonspecific leakage. This bidirectional imbalance is particularly pronounced in high-barrier organs such as the blood–brain barrier (BBB), renal glomeruli, and pulmonary capillaries, thereby accelerating the progression of neurodegenerative disorders, chronic kidney disease, and pulmonary vascular pathologies.^{8,9}

Precise regulation of vascular relaxation and contraction is essential for maintaining vascular tone and

facilitating efficient exchange of substances. In aged ECs, reduced nitric oxide (NO) production and bioavailability impair endothelium-dependent vasodilation.¹⁰ Meanwhile, in aged VSMCs, abnormal calcium channel signaling and actin cytoskeleton remodeling weaken contractile capacity.¹¹

Given the rapid acceleration of global population aging, it is not surprising that vascular aging has emerged as a central determinant of healthy life span. It is now widely recognized not only as the fundamental pathological substrate of cardiovascular disorders such as hypertension and atherosclerosis but also as a shared contributor to organ-specific diseases, frailty, and multisystem aging. Indeed, at least in model systems, rates of vascular aging may in fact be rate limiting for the entire organism.¹²

DRIVERS OF VASCULAR AGING

As with other tissues and organs, vascular aging is driven by multiple interacting factors. However, owing to its unique structure and function, the vasculature exhibits distinctive responses to these influences. For instance, the vascular wall is continuously subjected to pulsatile pressure and shear stress, rendering it particularly sensitive to factors that affect the cytoskeleton and extracellular matrix (ECM).¹³ Throughout the life span, blood vessels remain in direct contact with a multitude of circulating factors, thereby being persistently exposed to lipoproteins, inflammatory mediators, and sources of oxidative stress.¹⁴

In the following sections, we examine how the mechanistic pathways governing organismal aging from the simple worm to complex mammals also drive vascular aging and thereby the susceptibility to cardiovascular disease (Figure 2). We include among the pathways to be discussed cellular senescence, chronic inflammation,

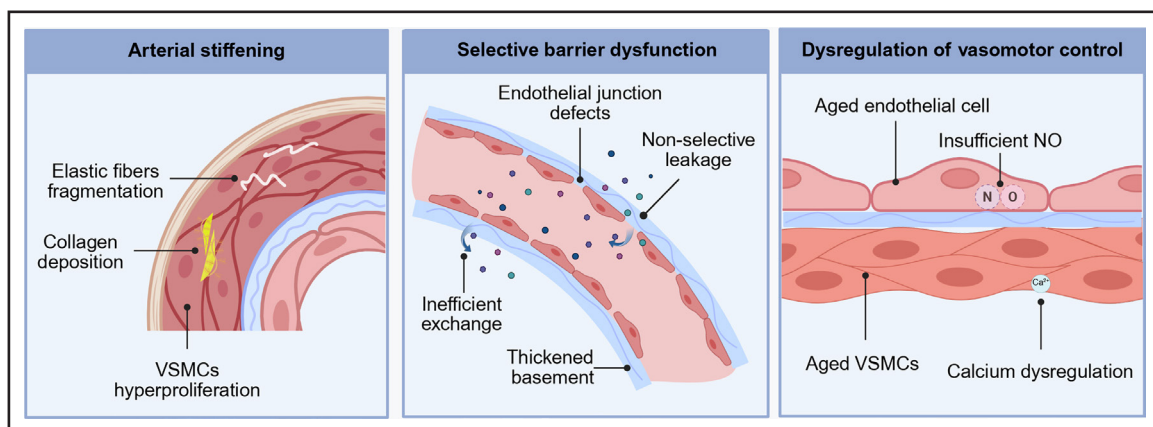


Figure 1. Characteristic phenotypes of vascular aging.

Arterial stiffening is characterized by fragmentation of elastic fibers, excessive collagen deposition, and vascular smooth muscle cell (VSMC) hyperproliferation. Selective barrier dysfunction with aging endothelial cells exhibiting impaired intercellular junctions and exchange and thickened basement membrane. Dysregulation of vasomotor control results in part from old endothelial cells exhibiting insufficient nitric oxide (NO) production, whereas aged VSMCs show calcium signaling dysregulation, together leading to impaired vascular tone regulation.

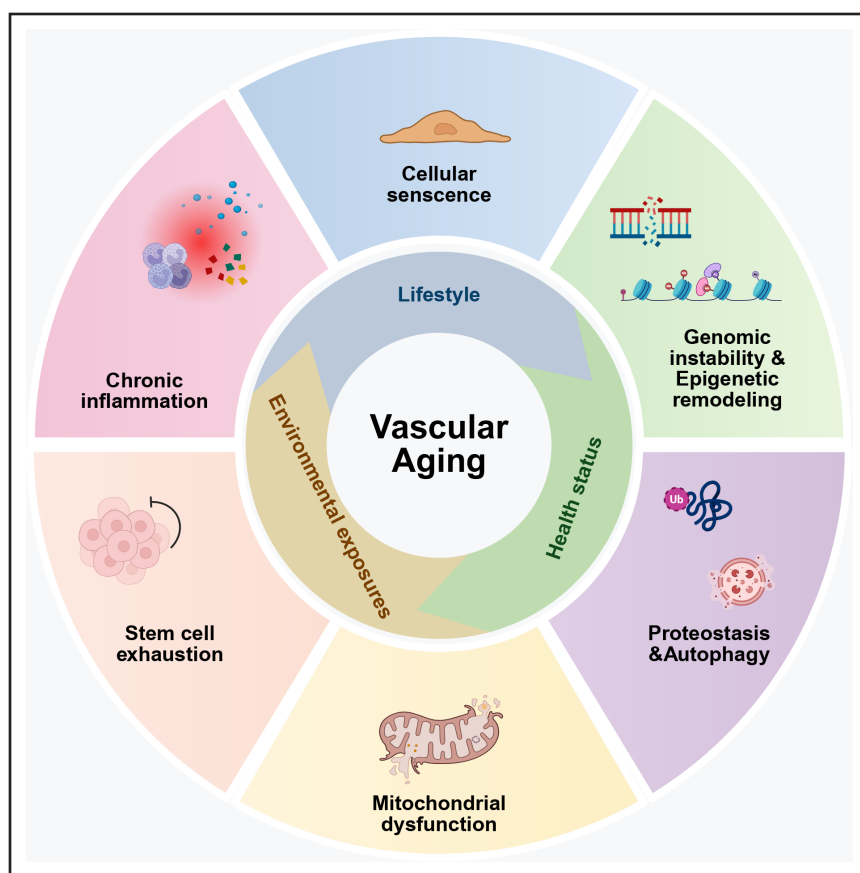


Figure 2. Risk factors and molecular drivers of vascular aging.

Identified cardiovascular risk factors modulate key molecular drivers to propel vascular aging. Individual health status (eg, insulin resistance, hypertension, and menopause), lifestyle (eg, diet, physical activity, smoking, and alcohol consumption), and environmental exposures (eg, geographic living conditions and pollution) all contribute to this process. These factors intersect with key molecular drivers, including cellular senescence, genomic instability and epigenetic remodeling, impaired proteostasis and autophagy, mitochondrial dysfunction, stem cell exhaustion, and chronic inflammation. Together, they orchestrate the complex progression of vascular aging. See text for additional details.

loss of proteostasis, mitochondrial dysfunction, genomic instability, epigenetic remodeling, and stem cell exhaustion.¹⁵ Together with the unique mechanical and metabolic milieu of the vasculature, these processes shape the distinctive pathological landscape of vascular aging.

CELLULAR SENESCENCE

Cellular senescence is a stable and irreversible state of cell-cycle arrest induced by diverse set of endogenous and exogenous stresses (eg, DNA damage, telomere shortening, oxidative stress, mitochondrial dysfunction). Although senescent cells no longer divide, they remain metabolically active and display characteristic phenotypes, including upregulation of various cell cycle inhibitors (eg, p16INK4a and p21Cip1), heterochromatin reorganization (formation of senescence-associated heterochromatin foci), and acquisition of a senescence-associated secretory phenotype (SASP), that entail abundant secretion of inflammatory cytokines, chemokines, and matrix-degrading enzymes.¹⁶

Cellular senescence is believed to be a major driver of tissue homeostatic imbalance and the decline in organ function. With age, senescent cells accumulate across multiple tissues, leading not only to cell-intrinsic functional deterioration but also, through their elaborated SASP, to a chronic inflammatory milieu (inflammaging). Through paracrine signaling, SASP factors further induce

senescence in neighboring cells, forming a potential self-reinforcing feedback loop.¹⁷ The basis for this increase in senescent cell burden with age is incompletely understood and could reflect a combination of increased generation of senescent cells and impaired immune-mediated clearance. By whatever mechanism, numerous animal studies have shown that eliminating senescent cells (eg, through senolytics)^{18–20} or suppressing their SASP (eg, through senomorphic agents) can delay aging in multiple organs, improve metabolic/functional indices, and extend overall health span.¹⁷

In the vascular system, senescent cells are prevalent in ECs and VSMCs and constitute core drivers of vascular aging and related diseases such as atherosclerosis.²¹ Recent advances in single-cell and spatial transcriptomics have further delineated the heterogeneity and spatial organization of senescent vascular cells. In murine models of atherosclerosis, senescent ECs and VSMCs have been mapped during arterial remodeling, providing unprecedented resolution into how senescence potentially shapes vascular pathology.²² These studies also attempted to define vascular-specific transcriptional signature of senescence, what is increasingly viewed as a “senotype.” This reflects the growing realization that there is no universal senescent cell and that the signature of senescence depends on a variety of factors, including the cell type of origin and how senescence was induced.

In ECs, cellular senescence and SASP factors (eg, interleukin [IL]-1 β , tumor necrosis factor- α , C-reactive protein) can suppress endothelial NO synthase expression through nuclear factor- κ B (NF- κ B) signaling while elevating intracellular reactive oxygen species (ROS), thereby reducing NO signaling and impairing endothelium-dependent vasodilation.²³ Concurrently, these inflammatory mediators upregulate adhesion molecules such as vascular cell adhesion molecule-1 and intracellular cell adhesion molecule-1, promoting inflammatory cell accumulation in the vessel wall and further compromising barrier integrity.²⁴ In VSMCs, senescence drives phenotypic switching, characterized by downregulation of contractile markers (α -smooth muscle actin, SM22 α) and upregulation of osteogenic markers (Runx2, BMP2 [bone morphogenetic protein 2], osteopontin), leading to medial calcification, loss of elasticity, and luminal stiffening.²⁵

SASP-derived matrix-degrading enzymes and inflammatory cytokines also compromise ECM integrity, increase endothelial barrier permeability, and activate immune-cell adhesion and infiltration.²⁶ Infiltrating immune cells—including macrophages, T cells, and neutrophils—further drive ECM degradation, elastic fiber fragmentation, and wall thinning by releasing matrix metalloproteinases, elastases, and ROS.^{14,27–30} Furthermore, senescent intimal foam cells have been shown to exert deleterious effects throughout all stages of atherosclerosis, underscoring the pathogenic role of senescence within immune-derived vascular cell populations.³¹

CHRONIC INFLAMMATION

In older individuals, a chronic inflammatory state, called inflammaging, often exists. This low-intensity, sterile induction of the innate immune system is closely linked to many age-related diseases.¹⁵ The basis for inflammaging is not fully understood. Clearly, the accumulation of senescent cells with age and their accompanying SASP may be a contributor. Other contributors to inflammaging include immune senescence whereby both innate immunity and adaptive immunity show quantitative and qualitative decline (eg, restricted T/B-cell repertoire, proinflammatory skewing), often resulting in a reduced capacity to maximally respond to a challenge while maintaining a high basal inflammatory state.³² In parallel, aging weakens gut barrier integrity and promotes dysbiosis, facilitating mucosal translocation of microbe-derived molecules/metabolites that chronically stimulate immunity and result in a systemic inflammatory burden.³³ Moreover, in ostensibly “sterile” contexts, tissue damage and metabolic derangements that increase with age result in the augmented release of damage-associated molecular patterns (DAMPs) that engage multiple pattern recognition receptors to sustain inflammatory signaling.³⁴

Chronic inflammation is believed to be a core driver of age-related diseases and physiological decline.

Persistent low-grade inflammation disrupts stem cell self-renewal, accelerates cellular senescence, and impairs regenerative niches while amplifying metabolic dysregulation and oxidative injury.¹⁵ Cytokines such as IL-6, tumor necrosis factor- α , and IL-1 β not only induce paracrine senescence but also modulate hematopoietic,³⁵ nervous,³⁶ and muscular systems,³⁷ potentially contributing to immunosuppression, cognitive decline, and sarcopenia.

Because the vessel wall is in direct contact with a host of circulating factors, chronic inflammation has particularly profound vascular effects. Inflammatory mediators impair vascular function through NF- κ B, JAK-STAT, and related signaling pathways—mechanistically similar to senescence/SASP effects—and are not reiterated here. Advanced glycation end-products (AGEs) activate receptor for AGEs, activating inflammatory pathways, driving AGEs deposition, and accelerating ECM degradation.³⁸ DAMPs directly engage ECs, repeatedly triggering innate immune sensors such as the NLRP3 inflammasome and the cGAS-STING pathway. This leads to persistent upregulation of the IL-1 β /IL-6 axis that is associated with reduced endothelial NO synthase activity and glycocalyx damage, thereby weakening barrier function and altering hemorheology.³⁹ It is important to emphasize that the cGAS-STING innate immune pathway, triggered by the presence of double-stranded DNA in the cytosol, plays a broad and central role in propagating cellular senescence and inflammation.⁴⁰ Increasingly, this pathway is being linked to aging⁴¹ and to chronic inflammatory age-related diseases, including atherosclerosis.⁴² Selective, small-molecule STING inhibitors are advancing to the clinic, suggesting the translational potential of this pathway to modulate a host of inflammatory-mediated diseases,⁴³ including a range of cardiovascular and metabolic conditions.⁴⁴ Besides cGAS-STING, immune activation of the vessel wall is also modulated by activated complement that can form the membrane attack complex, which in ECs induces the NLRP3 inflammasome and IL-1 β signaling, resulting in upregulation of adhesion/proinflammatory molecules, further amplifying inflammation and matrix degradation.⁴⁵ Furthermore, neutrophil extracellular traps capture platelets and coagulation factors to promote thrombosis⁴⁶; neutrophil extracellular trap-borne elastase, myeloperoxidase, and matrix metalloproteinases weaken fibrous-cap collagen and accelerate ECM degradation, and microthrombus deposition plus local stress concentration increases plaque vulnerability and the risk of rupture/erosion.³⁰ Last, aging is often accompanied by impaired production/signaling of specialized proresolving mediators, hindering timely termination of the inflammation-thrombosis cascade, prolonging immune-cell retention and delaying repair, and thus perpetuating a vicious cycle of inflammation-injury-reinflammation.⁴⁷

LOSS OF PROTEOSTASIS AND DECLINE OF AUTOPHAGIC FUNCTION

Proteostasis refers to the cellular capacity—through molecular chaperones, folding surveillance, the ubiquitin-proteasome system, and the autophagy-lysosome axis—to maintain proper protein configurations, localization, and turnover. During aging, there is a generalized decline in the overall capacity of chaperone networks, the ubiquitin-proteasome system, and autophagy, leading to accumulation of misfolded/aggregated proteins and damaged organelles across tissues. This is seen throughout the body; notable nonvascular examples include phosphorylated τ and α -synuclein aggregates in the nervous system; desmin-related aggregates in myocardium; damaged mitochondria in skeletal muscle/liver due to impaired PINK1-Parkin-dependent mitophagy; lipofuscin and undegraded autophagosomes in retinal pigment epithelium/neurons; and misfolded protein and mitochondrial aggregates in renal tubular epithelium.⁴⁸

In the vasculature, ECs and VSMCs inhabit a microenvironment of sustained mechanical stress, high metabolic demand, and intensive signaling, making them particularly vulnerable to proteostasis imbalance with age. Chronic hemodynamic forces (shear and pulsatile pressure) require continual renewal/repair of membrane and cytoskeletal proteins; vascular cells also participate broadly in immune responses, synthesizing and secreting abundant membrane and soluble proteins for immune-cell adhesion and positioning, together imposing a substantial load on the protein synthesis and degradation machinery.²¹

With aging, molecular chaperones such as BiP/GRP78 decline in function, predisposing to misfolded-protein accumulation.²¹ During atherogenesis, chaperone-mediated autophagy activity in macrophages drops markedly, principally as a result of reduced lysosomal LAMP-2A expression, compromising proteostasis, promoting misfolded/damaged proteins and proteotoxicity, activating inflammation, and accelerating vascular pathology.⁴⁹ In addition, reduced ubiquitin-proteasome system activity allows oxidized/glycated proteins to persist and aggregate; such aggregates, together with AGE deposits, significantly alter arterial biomechanics, increasing stiffness and accelerating vascular aging.⁵⁰

Aging vessels also exhibit lysosomal dysfunction that further lowers clearance efficiency. Age-related defects in lysosomal acidification resulting from impaired v-ATPase activity and dysregulated proton/metabolite transport shift organelle pH and inhibit hydrolase maturation/activity.⁵¹ Lipid microenvironment disturbances (eg, bis[monoacylglycerol]phosphate imbalance) in aged cells destabilize lysosomal membranes and further impair enzyme function.⁵² Loss of proteostasis can aggravate lysosomal defects by making damaged lysosomes refractory to various quality control processes, including

ESCRT-mediated repair and ubiquitin-dependent lysophagy.⁵³ In this context, directly increasing lysosomal number or activity in the vessel wall appears to prevent the development of age-related diseases such as atherosclerosis.⁵⁴

In addition, certain misfolded proteins accumulate specifically in aged vessels. Medin amyloid (derived from the C-terminal fragment of MFG-E8) deposits in the vasculature of nearly all individuals >50 years of age and is the most common vascular-associated amyloid. Medin binds directly to elastin and collagen, promoting ECM cross-linking and stiffening while inducing VSMC phenotypic switching and inflammatory signaling, thereby accelerating structural and functional vascular aging.^{55,56} Another example of vascular proteostasis failure is the accumulation of progerin, a truncated lamin A isoform originally linked to the accelerated aging syndrome Hutchinson-Gilford progeria.⁵⁷ As a misprocessed protein, progerin is inefficiently cleared by the ubiquitin-proteasome system and the autophagy-lysosome pathway, thereby imposing chronic proteotoxic stress on vascular cells. Progerin also appears in normal aging cells and tissues, linking nuclear envelope dysfunction to physiological aging.⁵⁸ Last, somatic lamin A mutations were shown to generate mosaic progerin-positive VSMC clones in human arteries, directly implicating defective proteostasis in early vascular aging.⁵⁹

MITOCHONDRIAL DYSFUNCTION

Mitochondrial dysfunction denotes a constellation of changes that lead to a persistent imbalance in energy metabolism, redox homeostasis, calcium dynamics, or quality control. In older individuals, impaired mitochondrial functions result from cumulative mtDNA and nuclear DNA damage, imbalances in fusion/fission dynamics, impaired mitophagy, reduced NAD⁺ levels, and damage induced by chronic inflammation. These changes collectively reduce respiratory chain activity and membrane potential while elevating ROS, thereby disrupting intracellular signaling and driving metabolic reprogramming.⁶⁰

Mitochondrial dysfunction is a key driver of organismal aging, promoting DNA damage, inflammatory responses, and cellular senescence through energy insufficiency and excess ROS. Damaged mitochondria can release mtDNA fragments and other mtDAMPs that activate innate immune pathways such as cGAS-STING, inducing chronic inflammation and SASP-like phenotypes.⁶¹ In multiple animal models, enhancing mitochondrial quality control, boosting NAD⁺, or promoting mitochondrial biogenesis delays organ functional decline, whereas inhibiting mitophagy or disturbing dynamics accelerates tissue aging and dysfunction.⁶²

The vascular system, especially ECs, is particularly sensitive to mitochondrial dysfunction. Beyond general effects, elevated mitochondrial ROS reduces NO

bioavailability and impairs vasodilation. With aging, increased electron leak from complexes I/III elevates mtROS; excessive ROS reacts with NO to form peroxynitrite, oxidizing proteins, lipids, and mtDNA, thereby diminishing endothelium-dependent relaxation and amplifying inflammation.⁶³ Concomitantly, reduced oxidative phosphorylation efficiency and ATP supply aggravate vasomotor dysfunction. With an energetic deficit, ECs/VSMCs struggle to maintain membrane potential and calcium homeostasis; when mitochondrial membrane potential and calcium uptake decline, the imbalance between mitochondrial calcium uniporter-mediated influx and Na⁺/Ca²⁺ exchanger-mediated efflux produces abnormal cytosolic Ca²⁺ transients, impairing endothelial relaxation and driving excessive VSMC contraction.^{64–66} Mitochondrial calcium overload further activates calcium-dependent proteases and phospholipases, promoting injury and apoptosis and accelerating degenerative changes in the vascular wall.⁶⁷

Physiologically, moderate mtROS levels participate in regulating vasodilation, gene expression, and antioxidant responses. Aging-related mtROS excess likely upends this balance, subverting ROS physiological signaling and instead acting as a damaging agent that activates proinflammatory pathways and drives sustained SASP release.⁶⁸ As mentioned, mtDNA fragments and other mtDAMPs further activate cGAS-STING; similarly, leakage of mitochondrial dsRNA into the cytosol can activate NF-κB and SASP through other specific intracellular sensors, further promoting systemic inflammaging.^{40,61}

As a key mechanism maintaining mitochondrial homeostasis, mitophagy often declines with age.⁶⁹ The basis for this decline may relate to alterations in lysosomal function, as discussed previously, and to alterations in mitochondrial dynamics, a process closely linked to mitophagic clearance.⁷⁰ Impaired mitophagy, in turn, acts as a contributor to a host of pathological processes, including hypertension, atherosclerosis, and pulmonary hypertension.^{71–73}

GENOMIC INSTABILITY

Genomic instability represents the accumulation of errors and imbalance in maintaining DNA structure, sequence, and chromosomal integrity, encompassing base damage, single-/double-strand breaks, chromosomal rearrangements, replication stress, and telomere dysfunction. In aging, chronic exposure to oxidative stress, inflammation, metabolic abnormalities, and environmental toxins increases DNA damage burden, and the efficiency of various repair pathways (nucleotide excision repair, base excision repair, homologous recombination, nonhomologous end joining) declines. Replication stress plus telomere dysfunction renders chromosome ends prone to misrecognition as double-strand breaks, initiating a DNA damage response.⁷⁴

Persistent genomic instability drives aging through multiple mechanisms. First, DNA damage activates the DNA damage response, triggering the p53-p21 and p16-Rb axes to potentially induce irreversible senescence and SASP.¹⁷ Second, mutational burden and chromatin architectural drift disrupt transcriptional homeostasis and stem cell self-renewal, undermining regenerative potential.⁷⁵ Furthermore, nucleic acids released by DNA damage can be sensed by cGAS-STING and other innate immune pathways, triggering chronic inflammation and immune reprogramming and amplifying systemic inflammaging through inflammatory-metabolic positive feedback.^{40,76}

Within the vasculature, ECs must continually proliferate/regenerate to withstand shear, pulsatile pressure, and metabolic stress, rendering them especially vulnerable to DNA damage and telomere dysfunction. In aged vessel walls, ECs at disturbed-flow regions show elevated markers of DNA double-strand breaks (γH2AX, 53BP1, etc) and increased telomere-associated damage foci, which collectively impair endothelial NO synthase-NO signaling and endothelium-dependent dilation, increase barrier permeability, and promote inflammatory cell infiltration.^{23,74} While in VSMCs, DNA damage can activate p53 and NF-κB signaling, promoting a procalcific phenotype.^{25,77}

EPIGENETIC REMODELING

Epigenetic remodeling encompasses sustained alterations in DNA methylation, histone modifications, chromatin structure, and noncoding RNA regulation without changes to the underlying DNA sequence. In aging, cumulative replication errors, oxidative/inflammatory stress, metabolic shifts (eg, NAD⁺ decline, imbalance between methyl donors and demethylase activity), and changes in cellular composition collectively drive DNA methylation drift, histone-modification imbalance, and chromatin reorganization.⁷⁸

Age-related epigenetic changes correlate strongly with frailty phenotypes, chronic disease risk, and mortality across systems; in population and longitudinal cohorts, accelerated epigenetic aging was robustly associated with cardiovascular outcomes and metabolic/inflammatory phenotypes, suggesting that epigenetic remodeling not only may be a readout but also may actively contribute to functional decline.⁷⁹ Emerging evidence suggests that manipulating certain epigenetic proteins (eg, SIRT6/HDACs) or DNA methylation regulators (eg, DNMT/TET) or using the Yamanaka factors to induce epigenetic reprogramming can ameliorate aging phenotypes in animal models. However, the reversibility and safety of these approaches in humans will require additional and more rigorous validation.⁸⁰

In the vasculature, epigenetic remodeling alters chromatin accessibility and thus the transcriptome, directly

driving aging phenotypes of ECs and VSMCs such as increased calcification propensity in VSMCs,⁸¹ reduced sensitivity of the NO pathway leading to vasodilatory dysfunction, and induction of endothelial-mesenchymal transition.⁸² Epigenetic changes can also establish epigenetic preconditioning; for example, H3K27ac-mediated enhancer landscape remodeling sensitizes cells to exaggerated responses to various secreted cytokines, thereby subsequently predisposing them to endothelial-mesenchymal transition and inflammatory activation.⁸³ Hemodynamic shear also modulates the epigenome in an age-dependent fashion. For instance, under physiological laminar flow, the transcription factor KLF4 recruits the SWI/SNF chromatin remodeling complex to beneficially reshape the enhancer landscape, increasing the accessibility of protective enhancers, whereas with aging plus disturbed flow, ECs more readily assume proinflammatory/adhesive phenotypes and lose vasodilatory capacity.⁸⁴

STEM CELL EXHAUSTION

Stem cell exhaustion reflects the progressive loss of number and function of adult tissue stem/progenitor cells, including diminished self-renewal, reduced differentiation potential, and delayed responsiveness. This undermines tissue repair and homeostasis, accelerating aging. Stem cell exhaustion markedly compromises regenerative and reparative capacity and is a key mechanism driving functional deterioration across organ systems. Declining stem cell function not only impedes tissue repair but also alters the microenvironment, aggravating inflammation and metabolic abnormalities. Concurrent epigenetic changes and amplified heterogeneity within stem cell pools lower regenerative quality and may cause aberrant differentiation or fibrosis, thereby accelerating overall aging and chronic disease.¹⁵

The most characterized vascular-related stem cells are CD133⁺ endothelial progenitor cells, originating from bone marrow and peripheral blood, which differentiate into ECs, home to injury sites, and promote vascular regeneration.⁸⁵ With aging, endothelial progenitor cell numbers and function decline overall and are considered biological indicators of reduced vascular repair capacity. In addition, the CD34⁺/KDR⁺ endothelial progenitor cell subpopulation is widely used to assess vascular regenerative capacity in populations; reduced levels associate closely with vascular aging and adverse hemodynamic phenotypes, including reduced endothelial function, increased arterial stiffness, and abnormal blood pressure risk.^{86–88}

Another key cell type is the mesenchymal stem cell, located in perivascular regions (eg, adventitia and glandular basal layers), providing structural support, modulating immunity, and contributing to vascular stability. Aging reduces mesenchymal stem cell chemotaxis and immunomodulation and biases differentiation toward fibrotic/

osteogenic fates.⁸⁹ Moreover, pericytes and vascular wall-resident stem cells decline with aging and critically affect capillary regeneration.⁹⁰

Aging induces a myeloid-biased shift in hematopoietic stem/progenitor cells, characterized by enhanced myelopoiesis accompanied by relatively reduced lymphopoiesis.⁹¹ This altered hematopoietic pattern skews the immune microenvironment toward a proinflammatory state, manifested by exacerbated vascular inflammation, accelerated endothelial injury, and increased macrophage infiltration and foam-cell formation.⁹² Collectively, these changes act as amplifiers in the progression of atherosclerosis, arterial stiffening, and hypertension, particularly by elevating SASP factors and oxidative stress within the vascular wall.

Clonal hematopoiesis of indeterminate potential (CHIP) represents another aging-associated hematopoietic abnormality. Its central mechanism involves the acquisition of somatic driver mutations in hematopoietic stem cells (eg, *DNMT3A*, *TET2*, *ASXL1*), leading to the expansion of mutant clones.⁹³ Unlike simple myeloid bias, CHIP not only alters lineage determination but also enhances the proinflammatory responsiveness of myeloid cells, for instance, through upregulation of IL-1 β and IL-6, thereby contributing to systemic inflammation and vascular pathology.⁹⁴ Clinical and population-based studies have demonstrated that CHIP is significantly associated with increased risk of cardiovascular events, particularly atherosclerosis and ischemic heart disease.⁹⁵ Notably, this association is unidirectional: CHIP promotes the development of atherosclerosis, whereas atherosclerosis does not increase the incidence of clonal hematopoiesis.⁹⁶

THERAPEUTIC INTERVENTION STRATEGIES

Given the pivotal role of vascular aging in age-related diseases, interventions targeting the drivers of vascular aging have become an increased focus in the treatment of age-associated vascular disorders (Figure 3). However, substantial interindividual variability influences the onset, progression, and therapeutic responsiveness of vascular diseases, highlighting the need to incorporate more precise and tailored interventions.^{97,98} Accordingly, this section summarizes current clinical advances from 3 perspectives: (1) interventions targeting the drivers of aging, (2) repair of damaged vascular structure and function, and (3) personalized diagnosis and treatment.

CLINICAL INTERVENTIONS TARGETING AGING DRIVERS

Eliminating or reprogramming senescent cells is a potential direction for clinical intervention. Early studies

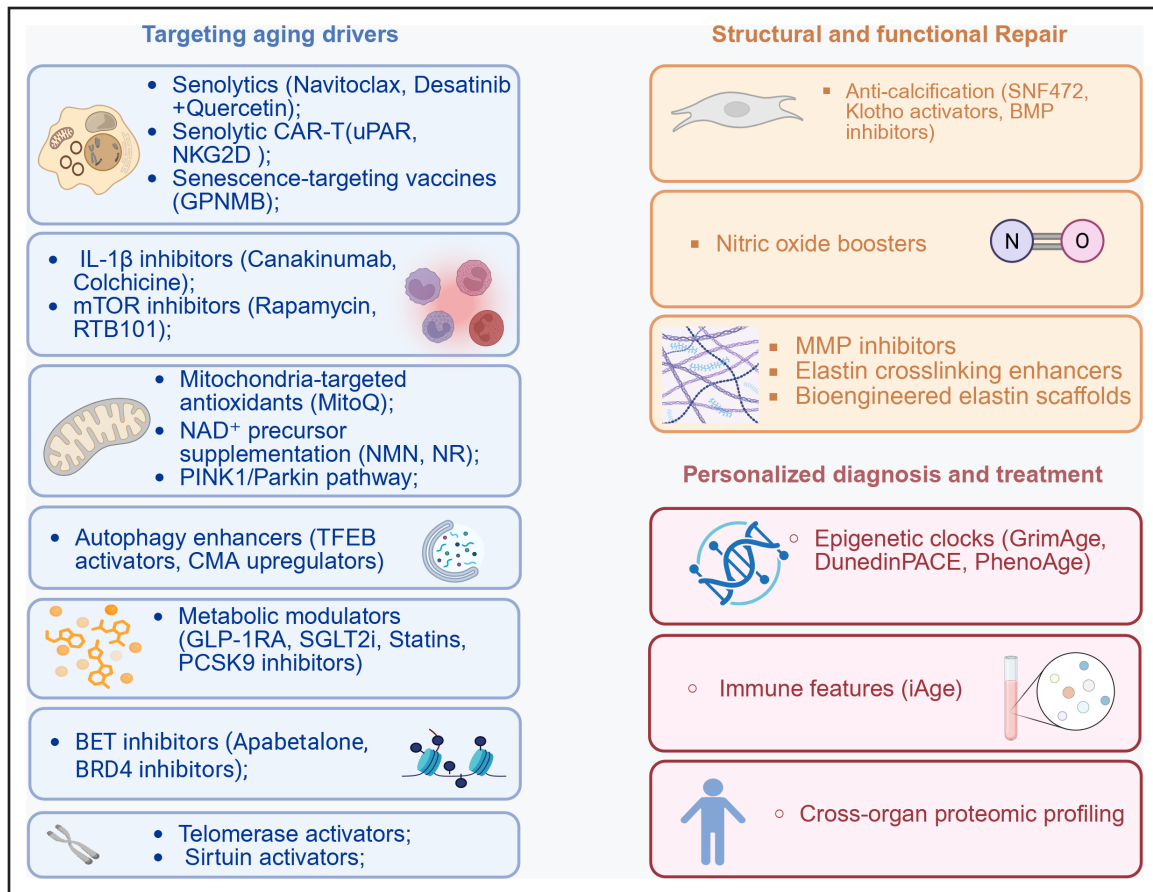


Figure 3. Therapeutic strategies and precision approaches for vascular aging.

Multiple strategies include targeting the drivers of aging, directed therapies aimed at the vascular wall, and in-depth molecular phenotyping. See text for additional details. BMP indicates bone morphogenetic protein; CAR-T, chimeric antigen receptor T cell; CMA, chaperone-mediated autophagy; GLP-1RA, glucagon-like peptide-1 receptor agonist; IL-1 β , interleukin-1 β ; MMP, matrix metalloproteinase; mTOR, mechanistic target of rapamycin; PCSK9, proprotein convertase subtilisin/kexin type 9; SGLT2i, sodium-glucose cotransporter 2 inhibitor; and uPAR, urokinase-type plasminogen activator receptor.

demonstrated that BCL-2 family inhibitors such as navitoclax can reduce senescent cell burden in experimental models and improve vascular barrier integrity and function.⁹⁹ Ultimately, the clinical use of such drugs may be limited by side effects such as myelosuppression. Within this evolving landscape, increasing attention has shifted toward endothelial- and vascular-targeting senolytics that potentially modulate aging of the vascular wall with greater specificity. For instance, preclinical studies show that fisetin may selectively eliminate senescent ECs and VSMCs, suppresses SASP output, and ameliorates vascular inflammation. In mouse models of cardiac and vascular aging, fisetin restores endothelial NO bioavailability, improves vasomotor responses, and reduces arterial stiffening.¹⁰⁰ Independently, fisetin has also been shown to protect the aging vasculature by reducing NF- κ B-driven cytokine production and strengthening antioxidant defenses, thereby mitigating age-related vascular permeability and inflammatory remodeling.¹⁰¹ In addition to small molecule approaches, chimeric antigen receptor T cells targeting potential senescent cell

surface antigens such as the apparent increased expression of the urokinase-type plasminogen activator receptor on senescent cells have shown improvement when deployed in various cardiovascular and metabolic disease models.¹⁰² Similarly, vaccines targeting other “senescence antigens” such as the transmembrane protein GPNMB appear to effectively reduce plaque burden in atherosclerosis models.¹⁰³ Notably, beyond senolytics and immune-based approaches, commonly used metabolic drugs have recently been implicated in senescence regulation. Sodium-glucose cotransporter 2 inhibitors, initially developed as antidiabetic agents, have been reported in preclinical studies to modulate cellular senescence and to attenuate pathological aging phenotypes across multiple tissues.¹⁰⁴ It is important to note that not all senolytic interventions appear beneficial. Recent evidence suggests that in preclinical pulmonary hypertension models, senolytics may actually have a deleterious effect.⁹⁹ Similarly, in the liver, removal of senescent ECs can also be detrimental.¹⁰⁵ These observations highlight the need for careful evaluation of senescence-targeting

interventions within complex physiological environments, as well as the necessity for outcomes-level validation.

Inflammatory amplification loops are important drivers of vascular aging, with a particularly prominent role in atherosclerosis progression. The CANTOS trial (Canakinumab Anti-Inflammatory Thrombosis Outcomes Study; canakinumab is an IL-1 β inhibitor) provided perhaps the first clinical evidence that anti-inflammatory therapy can reduce cardiovascular event risk independently of lipid levels.¹⁰⁶ Subsequently, COLCOT (Colchicine Cardiovascular Outcomes Trial) and LoDoCo2 (Low-Dose Colchicine 2) confirmed that low-dose colchicine reduces cardiovascular risk in patients with myocardial infarction or chronic coronary disease.^{107,108} In contrast, the recent large, multinational CLEAR SYNERGY OASIS-9 trial (Colchicine and Spironolactone in Patients With Myocardial Infarction/SYNERGY Stent Registry–Organization to Assess Strategies of Ischemic Syndromes 9) reported no reduction in major cardiovascular events when colchicine was initiated within 72 hours of a percutaneous coronary intervention in the setting of an acute myocardial infarction.¹⁰⁹ These neutral findings contrast with the positive results of COLCOT and LoDoCo2 and suggest that the efficacy of colchicine may depend on clinical scenario, the underlying inflammatory state, or potentially the patient's genotype. In that context, gene-inflammation interaction studies have revealed that carriers of CHIP mutations (eg, TET2) may derive the most benefit from IL-1 β inhibition,¹¹⁰ suggesting that future anti-inflammatory therapies, be it colchicine or canakinumab, could be tailored to individual inflammatory-genetic profiles.

Preclinical studies have shown that mTOR (mechanistic target of rapamycin) inhibitors (eg, rapamycin) can improve vascular pathology and delay stiffening by suppressing inflammation and promoting the clearance of damaged cellular components.¹¹¹ Similarly, strategies that activate chaperone-mediated autophagy or enhance autophagy by stimulation of the master transcription factor TFEB have demonstrated benefits in atherosclerosis and remodeling models. As previously noted, in the endothelium, TFEB activation restrains inflammatory signaling and reduces atherosclerosis⁵⁴; in mouse macrophages, TFEB-driven autophagy–lysosome biogenesis, achieved by genetic TFEB augmentation or pharmacologically by the addition of trehalose, enhances cargo clearance, lowers IL-1 β , and attenuates plaque progression.¹¹² In a similar vein, chaperone-mediated autophagy enhancement reduces atherosclerotic burden and improves lesion phenotypes affecting both VSMCs and macrophages.¹¹³ Mitochondria-targeted antioxidants such as MitoQ have been shown to improve endothelial function in aged animals.¹¹⁴ Recent studies suggest that NAD⁺ precursor supplementation may confer vascular or systemic benefits, although to date, most evidence is derived predominantly from small clinical trials and preclinical models.

For example, in a small randomized controlled study, the NAD⁺ precursor NMN appeared to improve endothelial function and reduce arterial stiffness, but larger outcome-driven trials are needed.¹¹⁵ Recent studies have also emphasized the role of regulating mitochondrial dynamics and mitophagy (PINK1/Parkin pathway) in cardiovascular aging interventions.¹¹⁶

Telomerase activation has demonstrated vascular benefits in animal and other preclinical models, in which it delays arteriosclerosis and improves endothelial function; likewise, epigenetic modulation through sirtuin activation has been shown to enhance endothelial function and to reduce arterial stiffness in experimental systems.^{117,118} However, there is no clinical evidence for either approach; therefore, their translational relevance requires further validation. Furthermore, epigenetic targeting can suppress VSMC phenotypic switching. For example, in mice, BET inhibitors, which block epigenetic reader proteins that recognize acetylated histones, can suppress smooth muscle phenotype conversion, representing a potential antiremodeling agent for vascular disease.¹¹⁹

STRUCTURAL AND FUNCTIONAL REPAIR

During aging, blood vessels progressively accumulate structural and functional damage. Once established, such damage can amplify upstream pathological processes if left unaddressed. Therefore, reparative therapies targeting preexisting vascular structural and functional decline constitute an essential component of strategies to slow vascular aging.

Endothelial dysfunction is one of the earliest hallmarks of vascular aging. Previous evidence indicates that improving NO bioavailability and suppressing ROS production can partially reverse endothelial impairment.^{21,23} High-dose statins and proprotein convertase subtilisin/kexin type 9 inhibitors have been well documented to improve endothelial function, in addition to their lipid-lowering effects.¹²⁰ Several randomized trials have demonstrated that glucagon-like peptide-1 receptor agonists and sodium-glucose cotransporter 2 inhibitors significantly improve flow-mediated dilation and reduce arterial stiffness in patients with diabetes or high cardiovascular risk.¹²¹ Of note, although these agents can enhance surrogate indices of vascular function—most notably flow-mediated dilation and arterial stiffness—such parameters serve as physiological proxies rather than definitive clinical outcomes. Consequently, the extent to which improvements in endothelial or hemodynamic measures translate into durable reductions in cardiovascular events remains to be established.

Vascular calcification is highly prevalent in the elderly and strongly associated with increased cardiovascular event risk. Early studies have shown that inhibiting

osteogenic phenotypic switching can delay progression of calcification. More recent findings indicate that activators of Klotho, an antiaging protein that also regulates mineral metabolism, and BMP signaling inhibitors can partially reverse calcium deposition in animal models of arterial calcification.¹²² In humans, a phase II trial of SNF472 (myo-inositol hexaphosphate), an intravenous inhibitor of hydroxyapatite formation, demonstrated significantly reduced progression of coronary artery calcification and aortic valve calcification in patients with end-stage kidney disease compared with placebo.¹²³ These findings suggest that targeted anticalcification therapies are approaching clinical relevance, although further trials are needed to assess effects on cardiovascular outcomes.

Elastin fragmentation and collagen imbalance are major contributors to reduced arterial compliance with age. Early *in vitro* studies demonstrated that reconstructing ECM architecture could improve vascular biomechanical properties. Animal studies using matrix metalloproteinase-specific inhibitors and elastin cross-linking enhancers have shown improved arterial compliance and delayed hypertension progression.¹²⁴ In addition, bioengineered elastin replacements and endoluminal regenerative scaffolds have entered the preclinical stage, offering potential for localized vascular repair in high-risk patients.¹²⁵

The central nervous system is particularly sensitive to changes in vascular perfusion capacity and barrier function.⁹ Large-scale proteomic analyses have revealed that vascular integrity loss associated with aging leads to BBB dysfunction, disruption of the cerebrospinal fluid–plasma protein balance, and subsequent cognitive impairment.¹²⁶ In recent years, notable progress has been made in strategies aimed at restoring BBB integrity. For example, activation of the Wnt signaling pathway has been shown to repair damaged BBB, whereas restoration of the endothelial glycocalyx in aged mice improved barrier integrity, reduced microhemorrhages, and enhanced cognitive function.^{127,128} In parallel, regression of cerebral microvessels has been demonstrated to directly reduce neuronal activity, providing mechanistic evidence that vascular rarefaction itself can impair brain function.¹²⁹ It is important to note that age-related decline in vascular endothelial growth factor signaling has been identified as a critical driver of vascular rarefaction, impaired BBB repair, and neurovascular dysfunction. Restoring vascular endothelial growth factor activity can counteract these age-associated changes, thereby promoting healthy aging and, remarkably, extending life span in experimental mouse models.¹²

PERSONALIZED DIAGNOSIS AND TREATMENT

Even among individuals of the same chronological age, there are striking differences in the extent of aging. This

has motivated the development of molecular clocks to more accurately capture an individual's true biological age. Moreover, aging is heterogeneous not only between individuals but also within a single individual in that different organs exhibit divergent rates of decline. In recent years, organ, pathway, and proteome level–based clocks have been devised.^{130,131} Molecular clock biomarkers can also serve as indicators of disease prognosis.¹³² Longitudinal studies have confirmed associations between organ-specific aging signatures and multisystem outcomes, underscoring the translational potential and public health value of these strategies.¹³³

In the cardiovascular system, plasma proteome-derived aging signatures demonstrate that accelerated heart aging is associated with a 250% increased risk of heart failure, whereas accelerated vascular aging is linked not only to cardiovascular disease but also to the progression of Alzheimer disease.¹³⁴ Likewise, large-scale population studies using the epigenetic GrimAge clock reveal that accelerated aging is associated with increased cardiovascular age and a greater subclinical atherosclerotic burden, highlighting a close relationship between blood DNA methylation clocks and vascular aging.¹³⁵

Other, noninvasive, nonmolecular techniques can also be applied to monitor vascular aging, including retinal vascular caliber,^{136,137} vascular ultrasound, pulse wave velocity, coronary artery calcium scoring, and artificial intelligence–enabled ECGs.¹³⁸ Looking forward, integrating molecular clocks with these noninvasive techniques and cross-organ biomarkers into multimodal, multiorgan predictive models may enable precise evaluation of cardiovascular biological age and provide a foundation for early intervention in high-risk populations.

SUMMARY, DISCUSSION, AND FUTURE PERSPECTIVES

As discussed, in recent years, multiple laboratories have identified key molecular mechanisms that individually drive vascular aging. Increasingly, there is evidence that these pathways intersect and potentially amplify each other. For example, mitochondrial dysfunction can cause mtDNA leakage, that which the cGAS-STING pathway, thereby promoting chronic inflammation⁶¹; epigenetic remodeling acting in concert with inflammation can regulate phenotypic switching of VSMCs.¹³⁹

In addition to these molecular drivers, dietary and lifestyle interventions have emerged as important modulators of vascular aging.¹⁴⁰ Caloric or dietary restriction improves endothelial function, reduces arterial stiffness, and activates longevity mediators such as AMPK and SIRT1, thereby lowering oxidative and inflammatory stress. Regular aerobic exercise similarly enhances NO bioavailability and attenuates systemic inflammation, whereas plant-based or Mediterranean dietary patterns

improve metabolic profile and reduce vascular oxidative burden. Together, these nonpharmacological strategies provide practical and evidence-supported approaches that complement emerging molecular interventions aimed at slowing vascular aging.¹⁴⁰

Beyond lifestyle and metabolic influences, additional exogenous and biological modifiers shape vascular aging trajectories. Environmental and circadian disruptions have been shown to accelerate vascular dysfunction, inflammation, and rates of vascular aging,^{141,142} whereas well-established sex-specific hormonal differences contribute to differential susceptibility to endothelial dysfunction and arterial stiffness between men and women.¹⁴³

In the future, beyond continued discovery of new drivers and mapping of complex interaction networks, mechanistic studies should also strive to “distill complexity into essentials” by pinpointing core nodes at which multiple drivers converge, thereby increasing the feasibility of clinical intervention. Indeed, unlike most areas in biology, aging lacks a single coherent theory, and the precise epistatic relationship between the drivers we have discussed remains unclear.

In the clinic, a range of emerging technologies are steadily translating into practice. Next-generation molecular biomarkers are being integrated with traditional modalities for early identification of vascular aging and risk stratification for cardiovascular events¹⁴⁴; multiomics approaches are enabling personalized intervention strategies,¹⁴⁵ and several aging drivers have already become drug targets. Looking ahead, additional cutting-edge strategies are poised to enter the clinic, including bioengineering technologies (such as milli-spinner thrombectomy),¹⁴⁶ tissue engineering, targeted nanodelivery, and mRNA-based therapies.

All these mechanistic insights and potential therapeutic advances combine to lead to a sense of justified enthusiasm. However, in truth, skeptics might provide a counterargument that perhaps all we will have achieved with these myriad efforts is to finally understand what Thomas Sydenham intuitively knew nearly 400 years ago.

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Dr Finkel is a cofounder and stockholder in Generian Pharmaceuticals and Coloma Therapeutics. Dr Chan has served as a consultant for Merck, Janssen, and United Therapeutics; is a director, officer, and shareholder in Synhale Therapeutics and Amlyson Therapeutics; holds research grants from United Therapeutics, Bayer, and WoodNext Foundation; and has filed patent applications on the targeting of metabolism and inflammation in pulmonary hypertension and the targeting of circadian rhythm. Dr Wang reports no conflicts.

REFERENCES

- Lin K, Dorman JB, Rodan A, Kenyon C. daf-16: An HNF-3/forkhead family member that can function to double the life-span of *Caenorhabditis elegans*. *Science*. 1997;278:1319–1322. doi: 10.1126/science.278.5341.1319
- Kenyon CJ. The genetics of ageing. *Nature*. 2010;464:504–512. doi: 10.1038/nature08980
- Partridge L, Fuentelba M, Kennedy BK. The quest to slow ageing through drug discovery. *Nat Rev Drug Discov*. 2020;19:513–532. doi: 10.1038/s41573-020-0067-7
- Morris BJ, Willcox DC, Donlon TA, Willcox BJ. FOXO3: a major gene for human longevity: a mini-review. *Gerontology*. 2015;61:515–525. doi: 10.1159/000375235
- Chen R, Morris BJ, Donlon TA, Masaki KH, Willcox DC, Davy PMC, Allsopp RC, Willcox BJ. FOXO3 longevity genotype mitigates the increased mortality risk in men with a cardiometabolic disease. *Aging (Albany NY)*. 2020;12:23509–23524. doi: 10.18632/aging.202175
- Chirinos JA, Segers P, Hughes T, Townsend R. Large-artery stiffness in health and disease: JACC state-of-the-art review. *J Am Coll Cardiol*. 2019;74:1237–1263. doi: 10.1016/j.jacc.2019.07.012
- Krouwer VJ, Hekking LH, Langelaar-Makkinje M, Regan-Klapisz E, Post JA. Endothelial cell senescence is associated with disrupted cell-cell junctions and increased monolayer permeability. *Vasc Cell*. 2012;4:12. doi: 10.1186/2045-824X-4-12
- Greene C, Connolly R, Brennan D, Laffan A, O’Keefe E, Zaporozhan L, O’Callaghan J, Thomson B, Connolly E, Argue R, et al. Blood-brain barrier disruption and sustained systemic inflammation in individuals with long COVID-associated cognitive impairment. *Nat Neurosci*. 2024;27:421–432. doi: 10.1038/s41593-024-01576-9
- Baaten CCFMJ, Vondenhoff S, Noels H. Endothelial cell dysfunction and increased cardiovascular risk in patients with chronic kidney disease. *Circ Res*. 2023;132:970–992. doi: 10.1161/circresaha.123.321752
- Förstermann U, Xia N, Li H. Roles of vascular oxidative stress and nitric oxide in the pathogenesis of atherosclerosis. *Circ Res*. 2017;120:713–735. doi: 10.1161/CIRCRESAHA.116.309326
- Touyz RM, Alves-Lopes R, Rios FJ, Camargo LL, Anagnostopoulou A, Arner A, Montezano AC. Vascular smooth muscle contraction in hypertension. *Cardiovasc Res*. 2018;114:529–539. doi: 10.1093/cvr/cvy023
- Grunewald M, Kumar S, Sharife H, Volinsky E, Gilleles-Hillel A, Licht T, Peryakova A, Hinden L, Azar S, Friedmann Y, et al. Counteracting age-related VEGF signaling insufficiency promotes healthy aging and extends life span. *Science*. 2021;373:eabc8479. doi: 10.1126/science.abc8479
- Lim XR, Harraz OF. Mechanosensing by vascular endothelium. *Annu Rev Physiol*. 2024;86:71–97. doi: 10.1146/annurev-physiol-042022-030946
- Björkegren JLM, Lusis AJ. Atherosclerosis: recent developments. *Cell*. 2022;185:1630–1645. doi: 10.1016/j.cell.2022.04.004
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. Hallmarks of aging: an expanding universe. *Cell*. 2023;186:243–278. doi: 10.1016/j.cell.2022.11.001
- Di Micco R, Krizhanovsky V, Baker D, d’Adda di Fagagna F. Cellular senescence in ageing: from mechanisms to therapeutic opportunities. *Nat Rev Mol Cell Biol*. 2021;22:75–95. doi: 10.1038/s41580-020-00314-w
- Wang B, Han J, Elisseeff JH, Demaria M. The senescence-associated secretory phenotype and its physiological and pathological implications. *Nat Rev Mol Cell Biol*. 2024;25:958–978. doi: 10.1038/s41580-024-00727-x
- Baker DJ, Childs BG, Durik M, Wijers ME, Sieben CJ, Zhong J, Saltness RA, Jeganathan KB, Verzosa GC, Pezeshki A, et al. Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. *Nature*. 2016;530:184–189. doi: 10.1038/nature16932
- Xu M, Pirtskhalava T, Farr JN, Weigand BM, Palmer AK, Weivoda MM, Inman CL, Ogrodnik MB, Hachfeld CM, Fraser DG, et al. Senolytics improve physical function and increase lifespan in old age. *Nat Med*. 2018;24:1246–1256. doi: 10.1038/s41591-018-0092-9

20. Amor C, Feucht J, Leibold J, Ho YJ, Zhu C, Alonso-Curbelo D, Mansilla-Soto J, Boyer JA, Li X, Giavridis T, et al. Senolytic CAR T cells reverse senescence-associated pathologies. *Nature*. 2020;583:127–132. doi: 10.1038/s41586-020-2403-9
21. Abdellatif M, Rainer PP, Sedej S, Kroemer G. Hallmarks of cardiovascular ageing. *Nat Rev Cardiol*. 2023;20:754–777. doi: 10.1038/s41569-023-00881-3
22. Mazan-Mamczarz K, Tsitsipatis D, Childs BG, Carr AE, Dos Santos CR, Anerillas C, Romero B, Gregg JM, Henry-Smith C, Okereke AN, et al. Single-cell and spatial transcriptomics map senescent vascular cells in arterial remodeling during atherosclerosis in mice. *Nat Aging*. 2025;5:1528–1547. doi: 10.1038/s43587-025-00889-z
23. Bloom SI, Islam MT, Lesniewski LA, Donato AJ. Mechanisms and consequences of endothelial cell senescence. *Nat Rev Cardiol*. 2023;20:38–51. doi: 10.1038/s41569-022-00739-0
24. Li S, He RC, Wu SG, Song Y, Zhang KL, Tang ML, Bei YR, Zhang T, Lu JB, Ma X, et al. LncRNA PSMB8-AS1 instigates vascular inflammation to aggravate atherosclerosis. *Circ Res*. 2024;134:60–80. doi: 10.1161/CIRCRESAHA.122.322360
25. Zhai X, Cao S, Wang J, Qiao B, Liu X, Hua R, Zhao M, Sun S, Han Y, Wu S, et al. Carbonylation of Runx2 at K176 by 4-hydroxynonenal accelerates vascular calcification. *Circulation*. 2024;149:1752–1769. doi: 10.1161/CIRCULATIONAHA.123.065830
26. Rolas L, Stein M, Barkaway A, Reglero-Real N, Sciacca E, Yaseen M, Wang H, Vazquez-Martinez L, Golding M, Blacksell IA, et al. Senescent endothelial cells promote pathogenic neutrophil trafficking in inflamed tissues. *EMBO Rep*. 2024;25:3842–3869. doi: 10.1038/s44319-024-00182-x
27. Engelen SE, Robinson AJB, Zurke YX, Monaco C. Therapeutic strategies targeting inflammation and immunity in atherosclerosis: how to proceed? *Nat Rev Cardiol*. 2022;19:522–542. doi: 10.1038/s41569-021-00668-4
28. Chen R, Zhang H, Tang B, Luo Y, Yang Y, Zhong X, Chen S, Xu X, Huang S, Liu C. Macrophages in cardiovascular diseases: molecular mechanisms and therapeutic targets. *Signal Transduct Target Ther*. 2024;9:130. doi: 10.1038/s41392-024-01840-1
29. Schumski A, Ortega-Gómez A, Wichapong K, Winter C, Lemnitzer P, Viola JR, Pinilla-Vera M, Folco E, Solis-Mezarino V, Völker-Albert M, et al. Endotoxemia accelerates atherosclerosis through electrostatic charge-mediated monocyte adhesion. *Circulation*. 2021;143:254–266. doi: 10.1161/CIRCULATIONAHA.120.046677
30. Cao J, Roth S, Zhang S, Kopczak A, Mami S, Asare Y, Georgakis MK, Messerer D, Horn A, Shemer R, et al; DEMDAS Study Group. DNA-sensing inflammasomes cause recurrent atherosclerotic stroke. *Nature*. 2024;633:433–441. doi: 10.1038/s41586-024-07803-4
31. Childs BG, Baker DJ, Wijshake T, Conover CA, Campisi J, van Deursen JM. Senescent intimal foam cells are deleterious at all stages of atherosclerosis. *Science*. 2016;354:472–477. doi: 10.1126/science.aaf6659
32. Nikolic-Zugich J. The twilight of immunity: emerging concepts in aging of the immune system. *Nat Immunol*. 2018;19:10–19. doi: 10.1038/s41590-017-0006-x
33. Ghosh TS, Shanahan F, O'Toole PW. The gut microbiome as a modulator of healthy ageing. *Nat Rev Gastroenterol Hepatol*. 2022;19:565–584. doi: 10.1038/s41575-022-00605-x
34. Gong T, Liu L, Jiang W, Zhou R. DAMP-sensing receptors in sterile inflammation and inflammatory diseases. *Nat Rev Immunol*. 2020;20:95–112. doi: 10.1038/s41577-019-0215-7
35. Kovtonyuk LV, Caiado F, Garcia-Martin S, Manz EM, Helbling P, Takizawa H, Boettcher S, Al-Shahrour F, Nombela-Arrieta C, Slack E, et al. IL-1 mediates microbiome-induced inflammaging of hematopoietic stem cells in mice. *Blood*. 2022;139:44–58. doi: 10.1182/blood.2021011570
36. Udeochu JC, Amin S, Huang Y, Fan L, Torres ERS, Carling GK, Liu B, McGurran H, Coronas-Samano G, Kauwe G, et al. Tau activation of microglial cGAS-IFN reduces MEF2C-mediated cognitive resilience. *Nat Neurosci*. 2023;26:737–750. doi: 10.1038/s41593-023-01315-6
37. Moiseeva V, Cisneros A, Sica V, Deryagin O, Lai Y, Jung S, Andrés E, An J, Segalés J, Ortet L, et al. Senescence atlas reveals an aged-like inflamed niche that blunts muscle regeneration. *Nature*. 2023;613:169–178. doi: 10.1038/s41586-022-05355-x
38. Yan SF, Ramasamy R, Naka Y, Schmidt AM. Glycation, inflammation, and RAGE: a scaffold for the macrovascular complications of diabetes and beyond. *Circ Res*. 2003;93:1159–1169. doi: 10.1161/01.RES.0000103862.26506.3D
39. Ma M, Jiang W, Zhou R. DAMPs and DAMP-sensing receptors in inflammation and diseases. *Immunity*. 2024;57:752–771. doi: 10.1016/j.immuni.2024.03.002
40. Gulen MF, Samson N, Keller A, Schwabenland M, Liu C, Gluck S, Thacker WV, Favre L, Mangeat B, Kroese LJ, et al. cGAS-STING drives ageing-related inflammation and neurodegeneration. *Nature*. 2023;620:374–380. doi: 10.1038/s41586-023-06373-1
41. Liu Y, Xu P. cGAS, an innate dsDNA sensor with multifaceted functions. *Cell Insight*. 2025;4:100249. doi: 10.1016/j.cellin.2025.100249
42. Wang SY, Chen YS, Jin BY, Bilal A. The cGAS-STING pathway in atherosclerosis. *Front Cardiovasc Med*. 2025;12:1550930. doi: 10.3389/fcvm.2025.1550930
43. Decout A, Katz JD, Venkatraman S, Ablasser A. The cGAS-STING pathway as a therapeutic target in inflammatory diseases. *Nat Rev Immunol*. 2021;21:548–569. doi: 10.1038/s41577-021-00524-z
44. Oduro PK, Zheng X, Wei J, Yang Y, Wang Y, Zhang H, Liu E, Gao X, Du M, Wang Q. The cGAS-STING signaling in cardiovascular and metabolic diseases: future novel target option for pharmacotherapy. *Acta Pharm Sin B*. 2022;12:50–75. doi: 10.1016/j.japsb.2021.05.011
45. Xie CB, Qin L, Li G, Fang C, Kirkiles-Smith NC, Tellides G, Pober JS, Jane-Wit D. Complement membrane attack complexes assemble NLRP3 inflammasomes triggering IL-1 activation of IFN- γ -primed human endothelium. *Circ Res*. 2019;124:1747–1759. doi: 10.1161/CIRCRESAHA.119.314845
46. Döring Y, Libby P, Soehnlein O. Neutrophil extracellular traps participate in cardiovascular diseases: recent experimental and clinical insights. *Circ Res*. 2020;126:1228–1241. doi: 10.1161/CIRCRESAHA.120.315931
47. Doran AC. Inflammation resolution: implications for atherosclerosis. *Circ Res*. 2022;130:130–148. doi: 10.1161/CIRCRESAHA.121.319822
48. Louros N, Schymkowitz J, Rousseau F. Mechanisms and pathology of protein misfolding and aggregation. *Nat Rev Mol Cell Biol*. 2023;24:912–933. doi: 10.1038/s41580-023-00647-2
49. Qiao L, Ma J, Zhang Z, Sui W, Zhai C, Xu D, Wang Z, Lu H, Zhang M, Zhang C, et al. Deficient chaperone-mediated autophagy promotes inflammation and atherosclerosis. *Circ Res*. 2021;129:1141–1157. doi: 10.1161/CIRCRESAHA.121.318908
50. Ungvari Z, Tarantini S, Donato AJ, Galvan V, Csiszar A. Mechanisms of vascular aging. *Circ Res*. 2018;123:849–867. doi: 10.1161/CIRCRESAHA.118.311378
51. Lee JH, Yang DS, Goulbourne CN, Im E, Stavrides P, Pensalfini A, Chan H, Bouchet-Marquis C, Bleivas C, Berg MJ, et al. Faulty autolysosome acidification in Alzheimer's disease mouse models induces autophagic build-up of A β in neurons, yielding senile plaques. *Nat Neurosci*. 2022;25:688–701. doi: 10.1038/s41593-022-01084-8
52. Nyame K, Xiong J, Alsohybe HN, de Jong APH, Peña IV, de Miguel R, Brummelkamp TR, Hartmann G, Nijman SMB, Raaben M, et al. PLA2G15 is a BMP hydrolase and its targeting ameliorates lysosomal disease. *Nature*. 2025;642:474–483. doi: 10.1038/s41586-025-08942-y
53. Gahlot P, Kravic B, Rota G, van den Boom J, Levantovsky S, Schulze N, Maspero E, Polo S, Behrends C, Meyer H. Lysosomal damage sensing and lysophagy initiation by SPG20-ITCH. *Mol Cell*. 2024;84:1556–1569.e10. doi: 10.1016/j.molcel.2024.02.029
54. Lu H, Fan Y, Qiao C, Liang W, Hu W, Zhu T, Zhang J, Chen YE. TFEB inhibits endothelial cell inflammation and reduces atherosclerosis. *Sci Signal*. 2017;10:ea4214. doi: 10.1126/scisignal.aah4214
55. Degenhardt K, Wagner J, Skodras A, Candlish M, Koppelman AJ, Wild K, Maxwell R, Rotermund C, von Zweydford F, Gloeckner CJ, et al. Medin aggregation causes cerebrovascular dysfunction in aging wild-type mice. *Proc Natl Acad Sci U S A*. 2020;117:23925–23931. doi: 10.1073/pnas.2011133117
56. Wagner J, Degenhardt K, Veit M, Louros N, Konstantoulea K, Skodras A, Wild K, Liu P, Obermüller U, Bansal V, et al. Medin co-aggregates with vascular amyloid- β in Alzheimer's disease. *Nature*. 2022;612:123–131. doi: 10.1038/s41586-022-05440-3
57. Eriksson M, Brown WT, Gordon LB, Glynn MW, Singer J, Scott L, Erdos MR, Robbins CM, Moses TY, Berglund P, et al. Recurrent de novo point mutations in lamin A cause Hutchinson-Gilford progeria syndrome. *Nature*. 2003;423:293–298. doi: 10.1038/nature01629
58. Scaffidi P, Misteli T. Lamin A-dependent nuclear defects in human aging. *Science*. 2006;312:1059–1063. doi: 10.1126/science.1127168
59. Revêchon G, Witasp A, Viceconte N, Helgadottir HT, Machtel P, Stefani F, Whisenant D, Sola-Carvajal A, McGuinness D, Abutaleb NO, et al. Recurrent somatic mutation and progerin expression in early vascular aging of chronic kidney disease. *Nat Aging*. 2025;5:1046–1062. doi: 10.1038/s43587-025-00882-6
60. Amorim JA, Coppotelli G, Rolo AP, Palmeira CM, Ross JM, Sinclair DA. Mitochondrial and metabolic dysfunction in ageing and

- age-related diseases. *Nat Rev Endocrinol.* 2022;18:243–258. doi: 10.1038/s41574-021-00626-7
61. Victorelli S, Salmonowicz H, Chapman J, Martini H, Vizioli MG, Riley JS, Cloix C, Hall-Younger E, Machado Espindola-Netto J, Jurk D, et al. Apoptotic stress causes mtDNA release during senescence and drives the SASP. *Nature.* 2023;622:627–636. doi: 10.1038/s41586-023-06621-4
 62. Picca A, Faltg J, Auwerx J, Ferrucci L, D'Amico D. Mitophagy in human health, ageing and disease. *Nat Metab.* 2023;5:2047–2061. doi: 10.1038/s42255-023-00930-8
 63. Ottolini M, Hong K, Cope EL, Daneva Z, DeLalio LJ, Sokolowski JD, Marziano C, Nguyen NY, Altschmied J, Haendeler J, et al. Local peroxynitrite impairs endothelial transient receptor potential vanilloid 4 channels and elevates blood pressure in obesity. *Circulation.* 2020;141:1318–1333. doi: 10.1161/CIRCULATIONAHA.119.043385
 64. Garbincius JF, Elrod JW. Mitochondrial calcium exchange in physiology and disease. *Physiol Rev.* 2022;102:893–992. doi: 10.1152/physrev.00041.2020
 65. Bertero E, Maack C. Calcium signaling and reactive oxygen species in mitochondria. *Circ Res.* 2018;122:1460–1478. doi: 10.1161/CIRCRESAHA.118.310082
 66. Metwally E, Sanchez Solano A, Lavanderos B, Yamasaki E, Thakore P, McClenaghan C, Rios N, Radi R, Feng Earley Y, Nichols CG, et al. Mitochondrial Ca²⁺-coupled generation of reactive oxygen species, peroxynitrite formation, and endothelial dysfunction in Cantú syndrome. *JCI Insight.* 2024;9:e176212. doi: 10.1172/jci.insight.176212
 67. Bertero E, O'Rourke B, Maack C. Mitochondria do not survive calcium overload during transplantation. *Circ Res.* 2020;126:784–786. doi: 10.1161/CIRCRESAHA.119.316291
 68. Miwa S, Kashyap S, Chini E, von Zglinicki T. Mitochondrial dysfunction in cell senescence and aging. *J Clin Invest.* 2022;132:e158447. doi: 10.1172/jci158447
 69. Sun N, Yun J, Liu J, Malide D, Liu C, Rovira II, Holmström KM, Fergusson MM, Yoo YH, Combs CA, et al. Measuring in vivo mitophagy. *Mol Cell.* 2015;60:685–696. doi: 10.1016/j.molcel.2015.10.009
 70. Burman JL, Pickles S, Wang C, Sekine S, Vargas JNS, Zhang Z, Youle AM, Nezhich KL, Wu X, Hammer JA, et al. Mitochondrial fission facilitates the selective mitophagy of protein aggregates. *J Cell Biol.* 2017;216:3231–3247. doi: 10.1083/jcb.201612106
 71. Pei Y, Ren D, Yin Y, Shi J, Ai Q, Hao W, Luo X, Zhang C, Zhao Y, Bai C, et al. Endothelial FUNDC1 deficiency drives pulmonary hypertension. *Circ Res.* 2025;136:e1–e19. doi: 10.1161/CIRCRESAHA.124.325156
 72. Zhang Y, Weng J, Huan L, Sheng S, Xu F. Mitophagy in atherosclerosis: from mechanism to therapy. *Front Immunol.* 2023;14:1165507. doi: 10.3389/fimmu.2023.1165507
 73. Schreckenberger ZJ, Wenceslau CF, Joe B, McCarthy CG. Mitophagy in hypertension-associated premature vascular aging. *Am J Hypertens.* 2020;33:804–812. doi: 10.1093/ajh/hpaa058
 74. Rossiello F, Jurk D, Passos JF, d'Adda di Fagnana F. Telomere dysfunction in ageing and age-related diseases. *Nat Cell Biol.* 2022;24:135–147. doi: 10.1038/s41556-022-00842-x
 75. Mitchell E, Spencer Chapman M, Williams N, Dawson KJ, Mende N, Calderbank EF, Jung H, Mitchell T, Coorens THH, Spencer DH, et al. Clonal dynamics of haematopoiesis across the human lifespan. *Nature.* 2022;606:343–350. doi: 10.1038/s41586-022-04786-y
 76. Nassour J, Aguiar LG, Correia A, Schmidt TT, Mainz L, Przetocka S, Haggblom C, Tadepalle N, Williams A, Shokhirev MN, et al. Telomere-to-mitochondria signalling by ZBP1 mediates replicative crisis. *Nature.* 2023;614:767–773. doi: 10.1038/s41586-023-05710-8
 77. Lanzer P, Hannan FM, Lanzer JD, Janzen J, Raggi P, Furniss D, Schuchardt M, Thakker R, Fok PW, Saez-Rodriguez J, et al. Medial arterial calcification: JACC state-of-the-art review. *J Am Coll Cardiol.* 2021;78:1145–1165. doi: 10.1016/j.jacc.2021.06.049
 78. Loyfer N, Magenheimer J, Peretz A, Cann G, Bredno J, Klochendler A, Fox-Fisher I, Shabi-Porat S, Hecht M, Pelet T, et al. A DNA methylation atlas of normal human cell types. *Nature.* 2023;613:355–364. doi: 10.1038/s41586-022-05580-6
 79. Joyce BT, Gao T, Zheng Y, Ma J, Hwang SJ, Liu L, Nannini D, Horvath S, Lu AT, Bai Allen N, et al. Epigenetic age acceleration reflects long-term cardiovascular health. *Circ Res.* 2021;129:770–781. doi: 10.1161/circresaha.121.318965
 80. Yang JH, Hayano M, Griffin PT, Amorim JA, Bonkowski MS, Apostolidis JK, Salfati EL, Blanchette M, Munding EM, Bhakta M, et al. Loss of epigenetic information as a cause of mammalian aging. *Cell.* 2023;186:305–326.e27. doi: 10.1016/j.cell.2022.12.027
 81. Mocci G, Sukhvasi K, Örd T, Bankier S, Singha P, Arasu UT, Agbabiaye OO, Mäkinen P, Ma L, Hodonsky CJ, et al. Single-cell gene-regulatory networks of advanced symptomatic atherosclerosis. *Circ Res.* 2024;134:1405–1423. doi: 10.1161/CIRCRESAHA.123.323184
 82. Augustin HG, Koh GY. A systems view of the vascular endothelium in health and disease. *Cell.* 2024;187:4833–4858. doi: 10.1016/j.cell.2024.07.012
 83. Jin YJ, Liang G, Li R, Wang S, Alnouri MW, Bentsen M, Kuenne C, Günther S, Yan Y, Li Y, et al. Phosphorylation of endothelial histone H3.3 serine 31 by PKN1 links flow-induced signaling to proatherogenic gene expression. *Nat Cardiovasc Res.* 2025;4:180–196. doi: 10.1038/s44161-024-00593-y
 84. Moonen JR, Chappell J, Shi M, Shinohara T, Li D, Mumbach MR, Zhang F, Nair RV, Nasser J, Mai DH, et al. KLF4 recruits SWI/SNF to increase chromatin accessibility and reprogram the endothelial enhancer landscape under laminar shear stress. *Nat Commun.* 2022;13:4941. doi: 10.1038/s41467-022-32566-9
 85. Sun S, Meng Y, Li M, Tang X, Hu W, Wu W, Li G, Pang Q, Wang W, Liu B. CD133+ endothelial-like stem cells restore neovascularization and promote longevity in progeroid and naturally aged mice. *Nat Aging.* 2023;3:1401–1414. doi: 10.1038/s43587-023-00512-z
 86. Jiang L, Chen T, Sun S, Wang R, Deng J, Lyu L, Wu H, Yang M, Pu X, Du L, et al. Nonbone marrow CD34+ cells are crucial for endothelial repair of injured artery. *Circ Res.* 2021;129:e146–e165. doi: 10.1161/CIRCRESAHA.121.319494
 87. Kadir RRA, Rakkar K, Othman OA, Sprigg N, Bath PM, Bayraktutan U. Analysis of endothelial progenitor cell subtypes as clinical biomarkers for elderly patients with ischaemic stroke. *Sci Rep.* 2023;13:21843. doi: 10.1038/s41598-023-48907-7
 88. Hill JM, Zalos G, Halcox JP, Schenke WH, Waclawiw MA, Quyyumi AA, Finkel T. Circulating endothelial progenitor cells, vascular function, and cardiovascular risk. *N Engl J Med.* 2003;348:593–600. doi: 10.1056/NEJMoa022287
 89. Pouikli A, Parekh S, Maleszewska M, Nikopoulou C, Baghdadi M, Tripodi I, Folz-Donahue K, Hinze Y, Mesaros A, Hoey D, et al. Chromatin remodeling due to degradation of citrate carrier impairs osteogenesis of aged mesenchymal stem cells. *Nat Aging.* 2021;1:810–825. doi: 10.1038/s43587-021-00105-8
 90. Tamiato A, Tombar LS, Fischer A, Muhly-Reinholz M, Vanicek LR, Toğru BN, Neitz J, Glaser SF, Merten M, Rodriguez Morales D, et al. Age-dependent RGS5 loss in pericytes induces cardiac dysfunction and fibrosis. *Circ Res.* 2024;134:1240–1255. doi: 10.1161/CIRCRESAHA.123.324183
 91. Ross JB, Myers LM, Noh JJ, Collins MM, Carmody AB, Messer RJ, Dhuey E, Hasenkrug KJ, Weissman IL. Depleting myeloid-biased haematopoietic stem cells rejuvenates aged immunity. *Nature.* 2024;628:162–170. doi: 10.1038/s41586-024-07238-x
 92. Patterson MT, Firulyova MM, Xu Y, Hillman H, Bishop C, Zhu A, Hickok GH, Schrank PR, Ronayne CE, Caillot Z, et al. Trem2 promotes foamy macrophage lipid uptake and survival in atherosclerosis. *Nat Cardiovasc Res.* 2023;2:1015–1031. doi: 10.1038/s44161-023-00354-3
 93. Jaiswal S, Fontanillas P, Flannick J, Manning A, Grauman PV, Mar BG, Lindsley RC, Mermel CH, Burt N, Chavez A, et al. Age-related clonal hematopoiesis associated with adverse outcomes. *N Engl J Med.* 2014;371:2488–2498. doi: 10.1056/NEJMoa1408617
 94. Fuster JJ, MacLauchlan S, Zuriaga MA, Polackal MN, Ostriker AC, Chakraborty R, Wu CL, Sano S, Muralidharan S, Rius C, et al. Clonal hematopoiesis associated with TET2 deficiency accelerates atherosclerosis development in mice. *Science.* 2017;355:842–847. doi: 10.1126/science.aag1381
 95. Jaiswal S, Natarajan P, Silver AJ, Gibson CJ, Bick AG, Shvartz E, McConkey M, Gupta N, Gabriel S, Ardissino D, et al. Clonal hematopoiesis and risk of atherosclerotic cardiovascular disease. *N Engl J Med.* 2017;377:1111–1121. doi: 10.1056/NEJMoa1701719
 96. Díez-Díez M, Ramos-Neble BL, de la Barrera J, Silla-Castro JC, Quintas A, Vázquez E, Rey-Martín MA, Izzi B, Sánchez-García L, García-Lunar I, et al. Unidirectional association of clonal hematopoiesis with atherosclerosis development. *Nat Med.* 2024;30:2857–2866. doi: 10.1038/s41591-024-03213-1
 97. Regitz-Zagrosek V, Gebhard C. Gender medicine: effects of sex and gender on cardiovascular disease manifestation and outcomes. *Nat Rev Cardiol.* 2023;20:236–247. doi: 10.1038/s41569-022-00797-4
 98. Marston NA, Pirruccello JP, Melloni GEM, Kamanu F, Bonaca MP, Giugliano RP, Scirica BM, Wiviott SD, Bhatt DL, Steg PG, et al. Clonal hematopoiesis, cardiovascular events and treatment benefit in 63,700 individuals from five TIMI randomized trials. *Nat Med.* 2024;30:2641–2647. doi: 10.1038/s41591-024-03188-z

99. Born E, Lipskaia L, Breau M, Houssaini A, Beaulieu D, Marcos E, Pierre R, Do Cruzeiro M, Lefevre M, Derumeaux G, et al. Eliminating senescent cells can promote pulmonary hypertension development and progression. *Circulation*. 2023;147:650–666. doi: 10.1161/CIRCULATIONAHA.122.058794
100. Rodriguez Morales D, Larcher V, Ruz Jurado M, Arifaj D, Tombor LS, Zanders L, Zeiher AM, Kuppe C, John D, Wagner JUG, et al. Vascular niches are the primary hotspots in cardiac aging. *Circ Res*. 2025;137:1353–1367. doi: 10.1161/CIRCRESAHA.125.327060
101. Zhu Y, Doornebal EJ, Pirtskhalava T, Giorgadze N, Wentworth M, Fuhrmann-Stroissnigg H, Niedernhofer LJ, Robbins PD, Tchkonja T, Kirkland JL. New agents that target senescent cells: the flavone, fisetin, and the BCL-XL inhibitors, A1331852 and A1155463. *Aging (Albany NY)*. 2017;9:955–963. doi: 10.18632/aging.101202
102. Di Micco R. Goodbye, senescent cells: CAR-T cells unleashed to fight ageing. *Nat Rev Mol Cell Biol*. 2024;25:955. doi: 10.1038/s41580-024-00792-2
103. Suda M, Shimizu I, Katsumi G, Yoshida Y, Hayashi Y, Ikegami R, Matsumoto N, Yoshida Y, Mikawa R, Katayama A, et al. Senolytic vaccination improves normal and pathological age-related phenotypes and increases lifespan in progeroid mice. *Nat Aging*. 2021;1:1117–1126. doi: 10.1038/s43587-021-00151-2
104. Katsumi G, Shimizu I, Suda M, Yoshida Y, Furihata T, Joki Y, Hsiao CL, Jiaqi L, Fujiki S, Abe M, et al. SGLT2 inhibition eliminates senescent cells and alleviates pathological aging. *Nat Aging*. 2024;4:926–938. doi: 10.1038/s43587-024-00642-y
105. Grosse L, Wagner N, Emelyanov A, Molina C, Lacas-Gervais S, Wagner KD, Bulavin DV. Defined p16^{high} senescent cell types are indispensable for mouse healthspan. *Cell Metab*. 2020;32:87–99.e6. doi: 10.1016/j.cmet.2020.05.002
106. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, Fonseca F, Nicolau J, Koenig W, Anker SD, et al; CANTOS Trial Group. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med*. 2017;377:1119–1131. doi: 10.1056/NEJMoa1707914
107. Tardif JC, Kouz S, Waters DD, Bertrand OF, Diaz R, Maggioni AP, Pinto FJ, Ibrahim R, Gamra H, Kiwan GS, et al. Efficacy and safety of low-dose colchicine after myocardial infarction. *N Engl J Med*. 2019;381:2497–2505. doi: 10.1056/NEJMoa1912388
108. Nidorf SM, Fiolet ATL, Mosterd A, Eikelboom JW, Schut A, Opstal TSJ, The SHK, Xu XF, Ireland MA, Lenderink T, et al; LoDoCo2 Trial Investigators. Colchicine in patients with chronic coronary disease. *N Engl J Med*. 2020;383:1838–1847. doi: 10.1056/NEJMoa2021372
109. Jolly SS, d'Entremont MA, Lee SF, Mian R, Tyrwhitt J, Kedev S, Montalescot G, Cornel JH, Stankovic G, Moreno R, et al; CLEAR Investigators. Colchicine in acute myocardial infarction. *N Engl J Med*. 2025;392:633–642. doi: 10.1056/NEJMoa2405922
110. Svensson EC, Madar A, Campbell CD, He Y, Sultan M, Healey ML, Xu H, D'Aco K, Fernandez A, Wache-Mainier C, et al. TET2-driven clonal hematopoiesis and response to canakinumab: an exploratory analysis of the CANTOS randomized clinical trial. *JAMA Cardiol*. 2022;7:521–528. doi: 10.1001/jamacardio.2022.0386
111. Boada C, Zinger A, Tsao C, Zhao P, Martinez JO, Hartman K, Naoi T, Sukhoveshin R, Sushnitha M, Molinaro R, et al. Rapamycin-loaded biomimetic nanoparticles reverse vascular inflammation. *Circ Res*. 2020;126:25–37. doi: 10.1161/CIRCRESAHA.119.315185
112. Sergin I, Evans TD, Zhang X, Bhattacharya S, Stokes CJ, Song E, Ali S, Dehestani B, Holloway KB, Micevych PS, et al. Exploiting macrophage autophagy-lysosomal biogenesis as a therapy for atherosclerosis. *Nat Commun*. 2017;8:15750. doi: 10.1038/ncomms15750
113. Madrigal-Matute J, de Bruijn J, van Kuijk K, Riascos-Bernal DF, Diaz A, Tasset I, Martín-Segura A, Gijbels MJJ, Sander B, Kaushik S, et al. Protective role of chaperone-mediated autophagy against atherosclerosis. *Proc Natl Acad Sci U S A*. 2022;119:e2121133119. doi: 10.1073/pnas.2121133119
114. Song R, Dasgupta C, Mulder C, Zhang L. MicroRNA-210 controls mitochondrial metabolism and protects heart function in myocardial infarction. *Circulation*. 2022;145:1140–1153. doi: 10.1161/CIRCULATIONAHA.121.056929
115. Qiu Y, Xu S, Chen X, Wu X, Zhou Z, Zhang J, Tu Q, Dong B, Liu Z, He J, et al. NAD⁺ exhaustion by CD38 upregulation contributes to blood pressure elevation and vascular damage in hypertension. *Signal Transduct Target Ther*. 2023;8:353. doi: 10.1038/s41392-023-01577-3
116. Ai L, de Freitas Germano J, Huang C, Aniaq M, Sawaged S, Sin J, Thakur R, Rai D, Rainville C, Sterner DE, et al. Enhanced Parkin-mediated mitophagy mitigates adverse left ventricular remodeling after myocardial infarction: role of PR-364. *Eur Heart J*. 2025;46:380–393. doi: 10.1093/eurheartj/ehae782
117. Mojiri A, Walther BK, Jiang C, Matrone G, Holgate R, Xu Q, Morales E, Wang G, Gu J, Wang R, et al. Telomerase therapy reverses vascular senescence and extends lifespan in progeria mice. *Eur Heart J*. 2021;42:4352–4369. doi: 10.1093/eurheartj/ehab547
118. Yang K, Velagapudi S, Akhmedov A, Kraler S, Lapikova-Bryhinska T, Schmiady MO, Wu X, Geng L, Camici GG, Xu A, et al. Chronic SIRT1 supplementation in diabetic mice improves endothelial function by suppressing oxidative stress. *Cardiovasc Res*. 2023;119:2190–2201. doi: 10.1093/cvr/cvad102
119. Dutzmann J, Haertlé M, Daniel JM, Kloss F, Musmann RJ, Kalies K, Knöpp K, Pilowski C, Sirisko M, Sieweke JT, et al. BET bromodomain-containing epigenetic reader proteins regulate vascular smooth muscle cell proliferation and neointima formation. *Cardiovasc Res*. 2021;117:850–862. doi: 10.1093/cvr/cvaa121
120. Oesterle A, Laufs U, Liao JK. Pleiotropic effects of statins on the cardiovascular system. *Circ Res*. 2017;120:229–243. doi: 10.1161/CIRCRESAHA.116.308537
121. Ussher JR, Drucker DJ. Glucagon-like peptide 1 receptor agonists: cardiovascular benefits and mechanisms of action. *Nat Rev Cardiol*. 2023;20:463–474. doi: 10.1038/s41569-023-00849-3
122. Lim K, Lu TS, Molostvov G, Lee C, Lam FT, Zehnder D, Hsiao LL. Vascular Klotho deficiency potentiates the development of human artery calcification and mediates resistance to fibroblast growth factor 23. *Circulation*. 2012;125:2243–2255. doi: 10.1161/CIRCULATIONAHA.111.053405
123. Raggi P, Bellasi A, Bushinsky D, Bover J, Rodriguez M, Ketteler M, Sinha S, Salcedo C, Gillotti K, Padgett C, et al. Slowing progression of cardiovascular calcification with SNF472 in patients on hemodialysis: results of a randomized phase 2b study. *Circulation*. 2020;141:728–739. doi: 10.1161/CIRCULATIONAHA.119.044195
124. Regnault V, Lacolley P, Laurent S. Arterial stiffness: from basic primers to integrative physiology. *Annu Rev Physiol*. 2024;86:99–121. doi: 10.1146/annurev-physiol-042022-031925
125. Naegeli KM, Kural MH, Li Y, Wang J, Hugentobler EA, Niklason LE. Bioengineering human tissues and the future of vascular replacement. *Circ Res*. 2022;131:109–126. doi: 10.1161/CIRCRESAHA.121.319984
126. Oh HS, Le Guen Y, Rappoport N, Urey DY, Farinas A, Rutledge J, Channappa D, Wagner AD, Mormino E, Brunet A, et al. Plasma proteomics links brain and immune system aging with healthspan and longevity. *Nat Med*. 2025;31:2703–2711. doi: 10.1038/s41591-025-03798-1
127. Martin M, Vermeiren S, Bostaille N, Eubelen M, Spitzer D, Vermeersch M, Profaci CP, Pozuelo E, Toussay X, Raman-Nair J, et al. Engineered Wnt ligands enable blood-brain barrier repair in neurological disorders. *Science*. 2022;375:eabm4459. doi: 10.1126/science.abm4459
128. Shi SM, Suh RJ, Shon DJ, Garcia FJ, Buff JK, Atkins M, Li L, Lu N, Sun B, Luo J, et al. Glycocalyx dysregulation impairs blood-brain barrier in ageing and disease. *Nature*. 2025;639:985–994. doi: 10.1038/s41586-025-08589-9
129. Gao X, Chen XJ, Ye M, Li JL, Lu N, Yao D, Ci B, Chen F, Zheng L, Yi Y, et al. Reduction of neuronal activity mediated by blood-vessel regression in the adult brain. *Nat Commun*. 2025;16:5840. doi: 10.1038/s41467-025-60308-0
130. Ding Y, Zuo Y, Zhang B, Fan Y, Xu G, Cheng Z, Ma S, Fang S, Tian A, Gao D, et al. Comprehensive human proteome profiles across a 50-year lifespan reveal aging trajectories and signatures. *Cell*. 2025;188:5763–5784.e26. doi: 10.1016/j.cell.2025.06.047
131. Goeminne LJE, Vladimirova A, Eames A, Tyshkovskiy A, Argentieri MA, Ying K, Moqri M, Gladyshev VN. Plasma protein-based organ-specific aging and mortality models unveil diseases as accelerated aging of organismal systems. *Cell Metab*. 2025;37:205–222.e6. doi: 10.1016/j.cmet.2024.10.005
132. Yang Y, Lu X, Liu N, Ma S, Zhang H, Zhang Z, Yang K, Jiang M, Zheng Z, Qiao Y, et al. Metformin decelerates aging clock in male monkeys. *Cell*. 2024;187:6358–6378.e29. doi: 10.1016/j.cell.2024.08.021
133. Kivimäki M, Frank P, Pentti J, Jokela M, Nyberg ST, Blake A, Lindbohm JV, Oh HS, Singh-Manoux A, Wyss-Coray T, et al. Proteomic organ-specific ageing signatures and 20-year risk of age-related diseases: the Whitehall II observational cohort study. *Lancet Digit Health*. 2025;7:e195–e204. doi: 10.1016/j.landig.2025.01.006
134. Oh HS, Rutledge J, Nachun D, Pálóvcis R, Abiose O, Moran-Losada P, Channappa D, Urey DY, Kim K, Sung YJ, et al. Organ aging signatures in the plasma proteome track health and disease. *Nature*. 2023;624:164–172. doi: 10.1038/s41586-023-06802-1
135. Lu AT, Binder AM, Zhang J, Yan Q, Reiner AP, Cox SR, Corley J, Harris SE, Kuo PL, Moore AZ, et al. DNA methylation GrimAge version 2. *Aging (Albany NY)*. 2022;14:9484–9549. doi: 10.18632/aging.204434

136. Yu Z, Chen R, Gui P, Wang W, Razzak I, Alinejad-Rokny H, Zeng X, Shang X, Zhang L, Yang X, et al. A cross population study of retinal aging biomarkers with longitudinal pre-training and label distribution learning. *NPJ Digit Med*. 2025;8:344. doi: 10.1038/s41746-025-01751-7
137. Ortín Vela S, Beyeler MJ, Trofimova O, Iuliani I, Vargas Quiros JD, de Vries VA, Meloni I, Elwakil A, Hoogewoud F, Liefers B, et al. Phenotypic and genetic characteristics of retinal vascular parameters and their association with diseases. *Nat Commun*. 2024;15:9593. doi: 10.1038/s41467-024-52334-1
138. Raisi-Estabragh Z, Szabo L, Schuermans A, Salih AM, Chin CWL, Vágó H, Altmann A, Ng FS, Garg P, Pavanello S, et al. Noninvasive techniques for tracking biological aging of the cardiovascular system: JACC family series. *JACC Cardiovasc Imaging*. 2024;17:533–551. doi: 10.1016/j.jcmg.2024.03.001
139. Chakraborty A, Li Y, Zhang C, Li Y, Rebello KR, Li S, Xu S, Vasquez HG, Zhang L, Luo W, et al. Epigenetic induction of smooth muscle cell phenotypic alterations in aortic aneurysms and dissections. *Circulation*. 2023;148:959–977. doi: 10.1161/CIRCULATIONAHA.123.063332
140. Kodithuwakku V, Evans JT, Breslin M, Gall S, Climie RE. Modifiable factors associated with early vascular ageing in youth: a systematic review. *J Hypertens*. 2025;43:1912–1922. doi: 10.1097/HJH.0000000000004130
141. Chen Z, Xiong ZF, Liu X. Research progress on the interaction between circadian clock and early vascular aging. *Exp Gerontol*. 2021;146:111241. doi: 10.1016/j.exger.2021.111241
142. Anea CB, Zhang M, Stepp DW, Simkins GB, Reed G, Fulton DJ, Rudic RD. Vascular disease in mice with a dysfunctional circadian clock. *Circulation*. 2009;119:1510–1517. doi: 10.1161/CIRCULATIONAHA.108.827477
143. Regitz-Zagrosek V, Kararigas G. Mechanistic pathways of sex differences in cardiovascular disease. *Physiol Rev*. 2017;97:1–37. doi: 10.1152/physrev.00021.2015
144. Si J, Ma Y, Yu C, Sun D, Pang Y, Pei P, Yang L, Millwood IY, Walters RG, Chen Y, et al; China Kadoorie Biobank Collaborative Group. DNA methylation age mediates effect of metabolic profile on cardiovascular and general aging. *Circ Res*. 2024;135:954–966. doi: 10.1161/CIRCRESAHA.124.325066
145. Barnett SN, Cujba AM, Yang L, Maceiras AR, Li S, Kedlian VR, Pett JP, Polanski K, Miranda AMA, Xu C, et al. An organotypic atlas of human vascular cells. *Nat Med*. 2024;30:3468–3481. doi: 10.1038/s41591-024-03376-x
146. Chang Y, Wu S, Li Q, Pulli B, Salmi D, Yock P, Heit JJ, Zhao RR. Milli-spinner thrombectomy. *Nature*. 2025;642:336–342. doi: 10.1038/s41586-025-09049-0