

Contents lists available at ScienceDirect

Diabetes & Metabolic Syndrome: Clinical Research & Reviews



journal homepage: www.elsevier.com/locate/dsx

Joint association of diabetes mellitus and inflammation status with biological ageing acceleration and premature mortality

Check for updates

Fan Tang ^{a,b,c,d,1}, Shuang Yang ^{a,b,c,1}, Hongbin Qiu^d, Yan Liu^d, Shaohong Fang ^{a,b,c}, Yiying Zhang ^{d,**}, Shanjie Wang ^{a,b,c,*}

^a Department of Cardiology, Second Affiliated Hospital of Harbin Medical University, Harbin, China

^b The Key Laboratory of Myocardial Ischemia, Chinese Ministry of Education, China

^c State Key Laboratory of Frigid Zone Cardiovascular Diseases (SKLFZCD), Harbin, China

^d Department of Epidemiology and Biostatistics, School of Public Health, Jiamusi University, Jiamusi, China

ARTICLE INFO

Keywords: CRP KDMAccel PhenoAgeAccel Diabetes Biological ageing Inflammation

ABSTRACT

Background: We aimed to investigate the associations of diabetes mellitus (DM) and C-reactive protein (CRP) with biological ageing acceleration and mortality risk.

Methods: We analyzed data from 41,634 adults with CRP and DM at baseline. Subjects were categorized into high CRP (>3 mg/L) and low CRP (\leq 3 mg/L) groups. The cross-sectional endpoints of the study were biological ageing indicators Klemera-Doubal method BioAge acceleration (KDMAccel) and Phenotypic age acceleration (PhenoAgeAccel), and the follow-up endpoints were all-cause mortality and cardiovascular mortality.

Results: In adults with high CRP, compared with those without DM, PhenoAgeAccel increased by 1.66 years (95 % CI: 1.38–1.93), and 8.74 years (95 % CI: 8.25–9.22) in adults with prediabetes and DM, respectively (p for interaction <0.001). Using the CRP_{low}/non-DM group as a reference, adults in the CRP_{high}/non-DM, CRP_{low}/DM, and CRP_{high}/DM groups had significantly advanced biological ageing. Compared to adults without DM, low CRP, and no ageing acceleration, the multivariable-adjusted HRs (95%CIs) of all-cause and cardiovascular mortality in those with DM, CRP, and ageing acceleration were 3.22 (2.79–3.72), and 3.57 (2.81–4.54), respectively. *Conclusions*: These findings suggest that the joint presence of low-grade inflammation and DM might be asso-

conclusions: These findings suggest that the joint presence of low-grade inflammation and DM might be asso ciated with higher odds of biological ageing acceleration and premature mortality.

1. Introduction

Ageing stands out as a primary contributor to chronic diseases, emerging as a significant public health burden [1]. However, individuals of the same age exhibit significant heterogeneity in terms of age-related diseases and mortality risk, indicating variations in underlying biological ageing processes [2,3]. An increasing body of literature suggests that compared with chronological age, biological age, and ageing acceleration are more effective at predicting morbidity and mortality risks [4]. Many indicators of biological ageing have been identified, including DNA methylation age [5], and algorithms that integrate information from epigenetic, proteomic, and metabolomic profiling, such as the Klemera Biological Age [6] and Phenotypic Age [7].

Type 2 Diabetes (T2D) impacts more than 30 million adults in the

United States, with an additional 88 million individuals estimated to have prediabetes [8]. Elevated C-reactive protein (CRP) serves as a sensitive biomarker indicating systemic inflammation. Low-grade inflammation is commonly observed in people with diabetes mellitus (DM) and is partly mediated by adipose tissue dysfunction and insulin resistance [9,10]. High CRP levels pose a higher risk of poor prognosis in individuals with both diabetes and non-diabetes than in individuals with normal CRP levels [11,12]. Chronic metabolic control/diabetic chronic vascular complications are associated with CRP levels in the people with DM [13]. However, population evidence linking inflammation and diabetes to biological ageing acceleration remains unclear. In mice induced to become obese through diet or genetic factors, treatment strategies aimed at decreasing the burden of senescent cells, either through genetic targeting or the administration of senolytic drugs,

Received 30 December 2023; Received in revised form 27 May 2024; Accepted 30 May 2024 Available online 31 May 2024

1871-4021/© 2024 Research Trust of DiabetesIndia (DiabetesIndia) and National Diabetes Obesity and Cholesterol Foundation (N-DOC). Published by Elsevier Ltd. All rights are reserved, including those for text and data mining, AI training, and similar technologies.

^{*} Corresponding author. 246 Xuefu Road, Nangang District, 150086, Harbin, China.

^{**} Corresponding author. 258 Xuefu Road, Xiangyang District, 154007, Jiamusi, China.

E-mail addresses: zhangyiying@jmsu.edu.cn (Y. Zhang), shanjie_wang@hrbmu.edu.cn (S. Wang).

¹ FT and SY contributed equally to this article.

https://doi.org/10.1016/j.dsx.2024.103050

ameliorate diabetic phenotypes, resulting in improvements in glucose tolerance and insulin sensitivity [14]. The anti-ageing drug *ABT263* reverses hyperglycemia and restores normal β -*cell* gene expression profiles in mice with insulin resistance induced by the insulin receptor antagonist *S961* [15]. Recently, it has been demonstrated that inflammation contributes to chronic diseases associated with ageing [16]. In mouse models of progeria, interventions to reduce inflammation delay the onset of age-related characteristics and extend lifespan [17]. A low-level elevated inflammatory phenotype in *NF*- κ B gain-of-function mice is associated with a shorter lifespan and signs of biological ageing acceleration, supporting a causal relationship between chronic inflammation and age-related tissue dysfunction [18].

However, little research has been conducted on the joint association between cumulative exposure to biological ageing and future mortality risk. Therefore, this study aimed to investigate the relationships between different diabetic states (no diabetes, prediabetes, and diabetes), serum CRP levels, biological ageing acceleration, and mortality risk using the National Health and Nutrition Examination Survey (NHANES) data.

2. Methods

2.1. Study Population

The NHANES is a comprehensive cross-sectional survey conducted across the nation [19]. The survey stands out for combining interviews with comprehensive medical examinations. The overall sample of the survey was designed using a multistage stratified probability approach, ensuring its representativeness of the entire U.S. population. In this study, datasets from 11 two-year cycles (from 1999 to 2000 to 2017–2020) of the NHANES were selected. Additionally, the data from the 11 cycles underwent standardization and merging, employing interview weights in line with the National Center for Health Statistics recommendations.

Between 1999 and 2020, the NHANES program surveyed 116,876 participants. The exclusion criteria were as follows (Fig. 1): (1) participants aged less than 20 years (n = 52,563); (2) participants without available diabetes diagnostic data (n = 5572); (3) participants without available C-reactive protein data (n = 16,820); and (4) participants who

lacked biological ageing data (n = 287). Finally, 41,634 participants were recruited for analysis of biological age acceleration (n = 41,634) and phenotypic age acceleration (n = 41,408). The study received approval and written informed consent was obtained from all participants. For detailed information on the ethical review, please refer to the NHANES website.

2.2. Diabetes mellitus and serum C-reactive protein

The diabetic states included no diabetes, prediabetes, and diabetes. As described in our prior studies [20,21], diabetes was characterized by a self-reported diagnosis from a physician, the use of antihyperglycemic medications, plasma HbA1c levels $\geq 6.5 \%$ ($\geq 48 \text{ mmol/mol}$), or fasting plasma glucose (FPG) levels $\geq 7.0 \text{ mmol/L}$ (126 mg/dL); prediabetes was characterized by plasma HbA1c levels between 5.7 % and 6.4 % (39–46 mmol/mol) or FPG levels between 100 mg/dL and 125 mg/dL in adults without preexisting diabetes and antihyperglycemic agents. No diabetes was characterized by the absence of a prior diagnosis, no use of antihyperglycemic agents, HbA1c levels less than 5.7 %, or FPG levels less than 100 mg/dL. Blood samples were obtained during MEC visits using a standardized protocol [20].

CRP is considered a highly reliable indicator of the acute phase response to infectious diseases or other factors causing tissue injury and inflammation [22]. CRP levels were measured by latex-enhanced turbidimetry [22], and high-sensitivity c-reactive protein (hsCRP) concentrations were quantified by a Roche Cobas 6000 chemistry analyzer (Roche Diagnostics, Indianapolis, Indiana) using reagents and calibrators. The minimum reported level of CRP was approximately 0.2 mg/L (0.02 mg/dL), with slight variations depending on the calibrator lot. The lower limit of detection for hsCRP in the NHANES dataset was 0.15 mg/L. When the result is below the limit of detection, the variable's value is determined as the limit of detection divided by the square root of 2. For more detailed information on blood sample collection, storage, calibration, and quality control procedures for the determination of CRP, see the NHANES Laboratory Procedures Manual (CDC, 2020, 2021).

Elevated CRP is defined as >3 mg/L according to the recommendations [23]. All subjects were categorized into two groups: high CRP (>3



Fig. 1. Flow of study

AbbreviationsNHANES, the National Health and Nutrition Examination Survey; CRP, C-reactive protein; KDM, Klemera-Doubal method; KDM Accel, KDM Acceleration; PhenoageAccel, Phenotypic Age Acceleration.

mg/L) and low CRP (\leq 3 mg/L). Additionally, based on their CRP levels and diabetic states, participants were classified into four groups: CRP_{low}/non-DM, CRP_{high}/non-DM, CRP_{low}/DM, and CRP_{high}/DM.

2.3. Biological ageing indicators

Biological ageing was calculated using the R package BioAge package [24,24], including the Klemera-Doubal method BioAge (KDM Bio-Age) [6] and Phenotypic age (PhenoAge) [7]. We included biomarkers without CRP to calculate these 2 indicators of biological ageing. Biological ageing calculated using the biomarker algorithms without CRP correlates strongly with biological ageing measures estimated using the algorithms with CRP (r ranging from 0.9527 to 0.9996) [25]. To assess the biological acceleration of ageing, we computed residuals by regressing chronological age on biological age, called KDM BioAge acceleration (KDMAccel) and Phenotypic age acceleration (PhenoAgeAccel).

$$KDM BioAge = \frac{\sum_{i=1}^{n} (x_i - q_i) \frac{k_i}{s_i^2} + \frac{Chronological Age}{s_{BA}^2}}{\sum_{i=1}^{n} \left(\frac{k_i}{s_i}\right)^2 + \frac{1}{s_{BA}^2}}$$

KDMAccel = KDM BioAge – Chronological Age

Let x represent the value of the biomarker measured for the individual. For each biomarker i, the parameters k, q, and s are estimated through a regression analysis using the actual age of the biomarker in the reference sample. q, k, and s represent the regression intercept, slope, and root-mean-square error, respectively. The S_{BA} is a scaling factor that equals the square root of the variance of the parenchymal age explained by the biomarker, as established in the reference sample.

Phenotypic Age = 141.50 +
$$\frac{\ln \left[-0.00553 \times \ln \left(\exp \left(\frac{-1.51714 \times \exp(xb)}{0.0076927} \right) \right) \right]}{0.09165}$$

PhenoAgeAccel = *Phenotypic Age* - *Chronological Age*

where:

 $xb = -19.907 - 0.0336 \times Albumin + 0.0095 \times Creatinine + 0.1953$

imes Glucose + 0.0954 imes LnCRP - 0.0120 imes Lymphocyte Percent

+ 0.0268 \times Mean Cell Volume + 0.3306 \times Red Cell Distribution Width

+ 0.00188 \times Alkaline Phosphatase + 0.0554 \times White Blood Cell Count

+ 0.0804 imes Chronological Age

In our study, PhenoAge advancement was calculated as the disparity between the predicted biological age and the chronological age [26]. PhenoAge advancement was normalized to have a mean of 0 and a standard deviation of 1. A positive value signifies advanced states of biological ageing, whereas a negative value indicates delayed biological ageing. Additionally based on their CRP levels and diabetic states and PhenoAge advancement, the participants were classified into eight groups: CRP_{low}/non-DM/non-Ageing acceleration, CRP_{low}/non-DM/Ageing acceleration, CRP_{low}/DM/non-Ageing acceleration, CRP_{low}/D M/Ageing acceleration, CRP_{high}/non-DM/non-Ageing acceleration, CRP_{high}/non-DM/Ageing acceleration, CRP_{high}/DM/non-Ageing acceleration, and CRP_{high}/DM/Ageing acceleration.

2.4. All-cause and cardiovascular mortality

The follow-up endpoints of this study were all-cause and cardiovascular mortality. Mortality data for NHANES participants were obtained by linking to the National Death Index. According to the International Classification of Diseases, Tenth Revision (ICD-10) [27], we evaluated all-cause mortality and cardiovascular mortality. For each participant, the follow-up duration was calculated starting from their participation date until either death or December 31, 2019.

2.5. Other variables

We included sociodemographic variables, lifestyle factors, and health-related factors as covariates. The sociodemographic variables included age, sex (male/female), race/ethnicity (Mexican-American, other race, non-Hispanic white, non-Hispanic black), education level (less than high school, high school graduate, more than high school), marital status (married/cohabitating, divorced/widowed/separated, never married), and poverty income ratio (PIR, <1.3, 1.3-3.5, >3.5). Body mass index (BMI) was classified as normal (<25.0 kg/m²), overweight (25.0–29.9 kg/m²), or obese (>30.0 kg/m²). Lifestyle variables encompassed smoking status (never, former, current) heavy alcohol consumption (male >20 g/day, female >10 g/day), and physical activity (moderate-equivalent exercise: at least 150 min per week to meet the recommended target). Blood tests are performed in specialized laboratories using standard procedures. High-density lipoprotein (HDL) cholesterol and triglycerides are also present. Health-related factors included cardiovascular disease (CVD) and hypertension, which were defined based on a previous NHANES study [27].

2.6. Statistical analysis

As mentioned earlier, we performed all analyses according to the NHANES analysis guidelines [27]. Weighted means (standard errors [SE]) and percentages were employed for both continuous and categorical variables. Due to the small amount of missing covariates data (<5 %), we used multivariate interpolation with chained equations to deal with missing data [19].

The relationship between diabetes mellitus and biological ageing indicators (KDMAccel, PhenoAgeAccel) in different CRP levels, as well as the association between the combination of CRP (\leq or > 3 mg/L) and diabetes (no or yes) with biological ageing indicators, were investigated using survey-weighted multivariate linear regression models assessing the regression coefficients (β) and 95 % confidence intervals (CIs) and the significance of the interaction effect was assessed using the surveyweighted Wald test. Hazard ratios (HRs) and 95 % CIs were assessed by investigating the association of CRP, diabetes, and biological ageing status with all-cause mortality and cardiovascular mortality by weighted Cox proportional risk regression modeling. Two models were constructed to account for traditional risk factors: Model 1 adjusted for age, sex, and race/ethnicity. Model 2 built upon Model 1 by incorporating adjustments for education level, marital status, PIR, BMI, physical activity, alcohol consumption, smoking, HDL cholesterol, triglycerides, hypertension, and CVD.

Sensitivity analyses were conducted to mitigate reverse causality and bolster result stability. (1) Guidelines recommended [28] a threshold value of $\leq 1 \text{ mg/L}$ for low levels of CRP, and we investigated the relationship between diabetes mellitus and biological ageing indicators at a cutoff level of 1 mg/L for CRP; (2) excluded adults with CRP $\geq 10 \text{ mg/L}$, as abnormally high CRP levels may indicate active infections rather than chronic inflammation [29]; and (3) among persons with diagnosed DM, we evaluated trends in glycemic control defined by HbA1c <7.0 % or FBG 80–130 mg/dL [30] and investigated the relationship between glycemic control and biological ageing indicators; (4) hierarchical analysis by sex, age, hypertension and CVD.

Statistical analyses were performed using R (version 4.1.2) and STATA (version 15.1), and two-sided p values less than 0.05 indicated statistical significance.

3. Results

3.1. Participant characteristics

The study involved 41,634 participants aged 20 years or older. The characteristics of the participants were grouped according to CRP levels (Table 1). The weighted mean (SE) age was 48.7 [0.2] years, 39.1 % were men, and 16,488 (39.6 %) participants had serum CRP >3 mg/L. Participants with higher serum CRP were more likely to have a higher chronological age (48.7 years), KDM BioAge (49.5 years), and Phenotypic Age (49.6 years). Participants with elevated CRP had greater proportions of individuals with prediabetes, diabetes, CVD, and hypertension than did those with low CRP.

3.2. Relationship between diabetes mellitus, serum CRP, and biological ageing indicators

As expected, both diabetes mellitus and serum CRP levels were significantly associated with biological ageing (Table S1). After full adjustment for potential confounding factors, compared with those with no diabetes, KDMAccel increased by 0.86 years (95 % CI: 0.78–0.95), and 3.65 years (95 % CI: 3.43–3.87) in adults with prediabetes and diabetes, respectively, and PhenoAgeAccel increased by 1.60 years (95 % CI: 1.42–1.79), and 7.67 years (95 % CI: 7.30–8.04), respectively. KDMAccel increased by 0.63 years (95 % CI: 0.54–0.73) and the

Table 1

Baseline characteristics of participants in NHANES 1999–2020 by CRP levels.

Variables	$CRP \leq 3 mg/L$	CRP >3 mg/L
Age, year	46.3 ± 0.2	48.7 ± 0.2
Male, %	14000 (53.4)	6442 (39.1)
Race/ethnicity, %		
Non-Hispanic White	11604 (69.5)	7088 (66.5)
Non-Hispanic Black	4648 (9.4)	3792 (12.8)
Mexican American	4438 (7.8)	3327 (8.8)
Other	4420 (13.3)	2281 (11.9)
Education, %		
Less than high school	6506 (15.6)	4999 (19.6)
High school	5774 (24.0)	3998 (26.7)
More than high school	12866 (60.5)	7491 (53.7)
Marital status, %		
Married/Living with a partner	14188 (59.6)	8942 (58.3)
Widowed/Divorced/Separated	6008 (23.8)	3358 (20.5)
Never Married	4950 (16.7)	4188 (21.2)
Poverty income ratio, %		
≤ 1.30	8489 (16.7)	6564 (22.0)
1.31–3.5	8786 (32.5)	5840 (34.2)
>3.5	7811 (43.0)	4084 (35.8)
Body mass index, %		
>25 kg/m ²	9662 (40.0)	2887 (16.9)
25–30 kg/m ²	9345 (36.1)	4645 (27.4)
\geq 30 kg/m ²	6139 (24.0)	8956 (55.8)
Smoking status, %		
Never	18138 (55.1)	8601 (50.8)
Former	6252 (24.9)	4257 (26)
Current	5013 (20)	3630 (23.2)
Physical activity, %	9476 (43.8)	4298 (30.1)
Heavy drinking, g/d	1779 (8.5)	988 (7.3)
Diabetes, %		
Non-diabetes	14851 (65.1)	7443 (49.7)
Prediabetes	6906 (25.2)	5305 (31.9)
Diabetes	3389 (9.7)	3740 (18.4)
CVD, %	2255 (6.7)	2069 (10.6)
Hypertension, %	9384 (32.0)	8110 (45.2)
KDM Age, year	46.3 ± 0.2	49.5 ± 0.2
Phenotypic Age, year	43.7 ± 0.3	49.6 ± 0.2
Triglycerides, mmol/L	1.56 ± 0.1	1.87 ± 0.2
HDL-Cholesterol, mmol/L	1.42 ± 0.1	1.30 ± 0.1

Percentage, means and standard errors are adjusted for NHANES sampling weights. The observed numbers for categorical variables were unweighted. Abbreviations: KDM, Klemera-Doubal method; CVD, cardiovascular diseases; High-density Lipoprotein-Cholesterol, HDL-Cholesterol. PhenoAgeAccel increased by 2.52 years (95 % CI: 2.32–2.71) in the high CRP group compared with the low CRP group.

According to unadjusted and adjusted logistic regression models, the association between diabetes mellitus and biological ageing indicators KDMAccel or PhenoAgeAccel was especially significant in adults with elevated CRP levels compared to those with low CRP (Fig. 2). After adjustment for sociodemographic variables, among adults with serum CRP <3 mg/L, KDMAccel increased by 0.93 years (95 % CI: 0.82–1.05), and 3.35 years (95 % CI: 3.04-3.65) in adults with prediabetes and diabetes, respectively; and PhenoAgeAccel increased by 1.63 years (95 % CI. 1.42-1.84), and 7.15 years (95 % CI: 6.59-7.71) respectively, compared with those with no diabetes. Among adults with CRP >3 mg/ L, KDMAccel increased by 0.89 years (95 % CI: 0.73–1.05), and 4.54 years (95 % CI: 4.23-4.84) in adults with prediabetes and diabetes, respectively; and PhenoAgeAccel increased by 1.96 years (95 % CI: 1.67-2.26), and 9.66 years (95 % CI: 9.20-10.13), respectively, compared with those with no diabetes. According to the fully adjusted models, compared with those with no diabetes and CRP <3 mg/L, KDMAccel increased by 0.86 years (95 % CI: 0.75-0.98), and 3.11 years (95 % CI: 2.80–3.41) in adults with CRP <3 mg/L and prediabetes and diabetes, respectively, and PhenoAgeAccel increased by 1.43 years (95 % CI: 1.23-1.64) and 6.38 years (95 % CI: 5.83-6.92), respectively. Among adults with high CRP levels, compared with those with no diabetes, KDMAccel increased by 0.82 years (95 % CI: 0.67-0.98), and 4.08 years (95 % CI: 3.78-4.39), in adults with prediabetes and diabetes, respectively; and PhenoAgeAccel was increased by 1.65 years (95 % CI: 1.36-1.93), and 8.65 years (95 % CI: 8.16-9.14), respectively. A significant interaction effect was found between CRP and diabetes status on biological ageing acceleration in both the adjusted and unadjusted models (p for interaction <0.001).

We further investigated the relationships between the combinations of CRP and diabetes and biological ageing (Fig. 3). The population with diabetes and high CRP levels had the most severe biological ageing acceleration. In the fully adjusted models, using the CRP_{low}/non-DM group as a reference, adults in CRP_{high}/non-DM, CRP_{low}/DM, and CRP_{high}/DM groups had significantly advanced biological ageing with KDMAccel increasing by 0.40 years (95%CI: 0.30–0.49), 2.65 years (95%CI: 2.37–2.93) and 4.13 years (95%CI: 3.89–4.38), respectively; and PhenoAgeAccel increasing by 2.03 years (95%CI: 1.86–2.21), 5.66 years (95%CI: 5.14–6.18) and 9.88 years (95%CI: 9.46–10.13), respectively.

3.3. The associations between CRP, diabetes, biological ageing acceleration, and mortality risk

There were 6534 deaths in the study population during a median of 11.9 years of follow-up. We calculated the unadjusted and adjusted HRs and 95%CIs for all-cause and cardiovascular mortality risk across the combination of binary CRP and diabetes (Fig. S1). According to the fully adjusted models, using the CRPlow/non-DM group as a reference, adults in CRP_{high}/non-DM, CRP_{low}/DM, and CRP_{high}/DM groups were significantly associated with all-cause mortality with HR 1.38(95%CI: 1.28-1.49), 1.69(95%CI: 1.53-1.86) and 2.15(95%CI: 1.93-2.39), respectively, and increased risk of cardiovascular mortality with HR 1.23(95%CI: 1.08-1.40), 1.72(95%CI: 1.45-2.04) and 2.20(95%CI: 1.85-2.63), respectively. We further considered the association of biological ageing acceleration with mortality risk (Fig. 4). We stratified binary CRP (low/high), DM (no/yes), and PhenoAge advancement (no/ yes) to yield 8 combinations. Compared to adults with non-DM, low CRP, and no ageing acceleration, the multivariable-adjusted HRs (95% CIs) of all-cause and cardiovascular mortality in those with DM, CRP, and ageing acceleration were 3.22 (2.79-3.72), and 3.57 (2.81-4.54), respectively.

3.4. Additional analysis

We repeated the analysis for the association between diabetes and



Fig. 2. The relationship between diabetes mellitus and biological ageing acceleration in different C-reactive protein levels

We found a significant interaction between CRP and diabetes status on biological ageing acceleration in both adjusted and unadjusted models (p for interaction <0.001).

Abbreviations: CRP, C-reactive protein; KDM, Klemera-Doubal method; KDMAccel, KDM Age acceleration; PhenoAgeAccel, Phenotypic Age acceleration. Model 1: age (continuous), sex (female, male), race/ethnicity (Mexican-American, other Hispanic, non-Hispanic white, non-Hispanic black).

Model 2: age (continuous), sex (female, male), race/ethnicity (Mexican-American, other Hispanic, non-Hispanic white, non-Hispanic black), education level (less than high school, high school, more than high school), marital status (married/cohabitating, divorced/widowed/separated, never married), Poverty income ratio (<1.3, 1.3–3.5, >3.5). body mass index (<25.0 kg/m², 25.0–29.9 kg/m², \geq 30.0 kg/m²), smoking status (never, former, current), heavy alcohol consumption (male \geq 20g/day, female \geq 10g/day), physical activity (moderate-equivalent exercise: at least 150 min per week to meet the recommended target), Triglycerides (continuous), HDL-Cholesterol (continuous), hypertension (Yes/no) and cardiovascular diseases (Yes/no).



Fig. 3. The relationship between the combination of diabetes mellitus and different CRP levels and biological ageing indicators

Abbreviations: CRP, C-reactive protein; DM, diabetes mellitus; KDM, Klemera-Doubal method; KDMAccel, KDM Age acceleration; PhenoAgeAccel, Phenotypic Age acceleration.

Model 1: age (continuous), sex (female, male), race/ethnicity (Mexican-American, other Hispanic, non-Hispanic white, non-Hispanic black). Model 2: age (continuous), sex (female, male), race/ethnicity (Mexican-American, other Hispanic, non-Hispanic white, non-Hispanic black), education level (less than high school, high school, more than high school), marital status (married/cohabitating, divorced/widowed/separated, never married), Poverty income ratio (<1.3, 1.3–3.5, >3.5). body mass index (<25.0 kg/m², 25.0–29.9 kg/m², \geq 30.0 kg/m²), smoking status (never, former, current), heavy alcohol consumption (male \geq 20g/day, female \geq 10g/day), physical activity (moderate-equivalent exercise: at least 150 min per week to meet the recommended target), Triglycerides (continuous), HDL-Cholesterol (continuous), hypertension (Yes/no) and cardiovascular diseases (Yes/no).

biological ageing acceleration stratified by serum CRP with the new threshold ($\leq 1 \text{ mg/L}$ and >1 mg/L, Table S2). Consistently, biological ageing acceleration associated with diabetes was more significant in adults with CRP >1 mg/L compared to those with CRP $\leq 1 \text{ mg/L}$ (PhenoAgeAccel 7.98 versus 6.13 years). After excluding individuals with

CRP \geq 10 mg/L the findings were consistent with the primary analysis (Table S3). Taking the best glycemic control as a reference, after adjusting for confounders, KDMAccel increased by at least 2.20 years among adults with poorer glycemic control (Table S4). Subgroup analyses by age, sex, hypertension, and CVD were performed to further



Fig. 4. The relationship between the combinations of diabetes mellitus, different CRP levels, and ageing state and mortality risk Abbreviations: CRP, C-reactive protein; DM, diabetes mellitus; CVD, cardiovascular diseases.

Model 1: age (continuous), sex (female, male), race/ethnicity (Mexican-American, other Hispanic, non-Hispanic white, non-Hispanic black).

Model 2: age (continuous), sex (female, male), race/ethnicity (Mexican-American, other Hispanic, non-Hispanic white, non-Hispanic black), education level (less than high school, high school, more than high school), marital status (married/cohabitating, divorced/widowed/separated, never married), Poverty income ratio (<1.3, 1.3–3.5, >3.5). body mass index (<25.0 kg/m², 25.0–29.9 kg/m², \geq 30.0 kg/m²), smoking status (never, former, current), heavy alcohol consumption (male \geq 20g/day, female \geq 10g/day), physical activity (moderate-equivalent exercise: at least 150 min per week to meet the recommended target), hypertension (Yes/no) and cardiovascular diseases (Yes/no).

*Additionally based on binary CRP (low/high), DM (no/yes), and PhenoAge advancement (no/yes) to yield 8 combinations: CRP_{low}/non-DM/non-Ageing acceleration, CRP_{low}/non-DM/Ageing acceleration, CRP_{low}/DM/non-Ageing acceleration, CRP_{high}/DM/Ageing acceleration, CRP_{high}/non-DM/non-Ageing acceleration, CRP_{high}/non-DM/Ageing acceleration, CRP_{high}/non-DM/Ageing acceleration, CRP_{high}/non-DM/Ageing acceleration, CRP_{high}/non-DM/Ageing acceleration, CRP_{high}/DM/Ageing acceleration, CRP_{high}/DM/Ageing

explore the potential association of confounding factors on the association between diabetes and biological ageing indicators across various CRP levels (Tables S5–S8). The association between diabetes status and biological ageing was more significant among adults with aged less than 60 years and males.

4. Discussion

This study is the first to investigate the joint association of inflammation and diabetes mellitus on biological ageing in 41,634 US nationally representative participants. Our findings indicated that the association between diabetic status and accelerated biological ageing was more significant in adults with elevated CRP levels compared to those with low CRP. Notably, an accelerated rate of biological ageing was also observed in adults with prediabetes regardless of CRP levels, suggesting that biological ageing acceleration might be associated with impaired glycemic metabolism status before diabetes is diagnosed. Compared to the CRP_{low}/non-DM group, the CRP_{high}/DM group had the fastest rate of biological ageing and the highest risk of all-cause and cardiovascular mortality. The ageing acceleration phenotype might be increased the risk of mortality associated with diabetes, which was further amplified in populations with high CRP. Unexpectedly, the severity of biological ageing acceleration in populations with both diabetes and inflammation is more severe than that in those with diabetes or inflammation alone. Our finding underlined the importance of dual management of residual inflammation and blood glucose control which may substantially contribute to the improvement in healthy ageing.

Compared to young individuals, the number of SA- β -gal cells in isolated islets from healthy elderly individuals increases. Furthermore, compared to people with non-diabetes, individuals with T2D exhibit a further increase in the number of islets. This suggests that β -cell ageing may contribute to the pathogenesis of T2D [15]. Another model of chronic inflammation is based on the functional loss of the anti-inflammatory cytokine interleukin 10. In this scenario, the loss of anti-inflammatory signals leads to a state of low-grade proinflammatory status, accompanied by signs of accelerated ageing in middle age [31]. The research found that biological ageing is positively correlated with mortality in people with diabetes [32,33]. Our results support for the first time the population-based evidence to support the joint presence of low-grade inflammation and diabetes mellitus might be associated with an increased likelihood of biological ageing acceleration and premature mortality. Cribb et al. explored the association between inflammation and epigenetic ageing and found that the strongest associations were between CRP and GrimAgeAccel [34]. CRP-measured inflammation accounts for nearly one-third of the relationship between metabolic syndrome and GrimAge age acceleration [35]. Chiurazzi et al. reported that intermittent lipopolysaccharide-induced inflammation resulted in cognitive deficits [36].

Inflammation and immune-mediated injury are thought to play pathogenic roles in many diseases. Elevated CRP levels in plasma are associated with inflammation, injury, or bacterial infection [37]. Previous research has demonstrated an association between low-grade inflammation and the development of diabetes, cardiovascular disease, and malignant neoplasms [22,38]. Elevated CRP is linked to the

Diabetes & Metabolic Syndrome: Clinical Research & Reviews 18 (2024) 103050

accelerated development of diabetes mellitus, which is attributed to the biological mechanisms of low-grade chronic inflammation in glucose metabolism disorders [39]. Beyond elevating the risk of diabetes development, heightened inflammation may worsen the progression of diabetes by inducing the destruction of pancreatic tissue through inflammation-induced amyloid deposits [40,41]. In addition, increased inflammation (e.g., elevated CRP levels), including diabetic retinopathy [42,43] and cardiovascular disease [10], may increase diabetic end-organ damage. Our study further demonstrated the significant interaction of CRP and diabetes status on biological ageing acceleration and premature mortality. Although the mechanisms underlying the role of chronic inflammation in diabetes development remain unknown, the observation that adipose tissue synthesizes significant proinflammatory cytokines such as tumor necrosis factor, interleukins-1, and -6 and that inflammatory biomarkers correlate with body fat quantity indicates that activated innate immunity and inflammation are crucial biological factors in the pathogenesis of diabetes and diabetes mellitus [44].

Diabetes is a risk factor associated with CVD [45,46]. A mechanistic study in diabetic rats revealed that throughout the progression of diabetes, there are gradual changes in cardiac function that are closely associated with alterations in two groups of proteins, neurotrophic cascade proteins (NTF4) and electron transport chain cascade proteins (ETFBs) [47]. Multiple epidemiological studies have explored increased mortality associated with cardiovascular disease in individuals with diabetes. Evidence from the Japanese Observational Cohort Cardiovascular Prevention Study suggested that diabetes mellitus represents a significant risk factor for all-cause mortality and deaths specific to CVD, with a 2- to 4-fold increased risk of cardiovascular death [48]. A meta-analysis revealed a nonlinear association between CRP and both all-cause mortality and CVD mortality [49]. Previous studies demonstrated the predictive value of CRP for the risk of both all-cause and cardiovascular mortality [50,50], and a study conducted using NHANES data from 1999 to 2011 demonstrated that elevated CRP levels were associated with reduced overall and CVD survival [51]. In people with diabetes [52], CRP might be associated with all-cause mortality, cardiovascular mortality, and malignant neoplasm mortality, and the results are similar in nondiabetic, but risk levels differ. Our study revealed that the risk of all-cause and cardiovascular death was greater in the CRP_{high}/DM group than in the CRP_{low}/non-DM group. These findings highlight the need for joint management of inflammation and blood glucose to prevent ageing acceleration and premature death.

5. Clinical relevance

The clinical implications of our study lie in assessing the association of different inflammatory states and diabetes with biological ageing and premature death. It was found that diabetes might accelerate biological ageing in a low-grade inflammatory state, and the severity of accelerated biological ageing in the population with diabetes and inflammation is more severe than that in the population with diabetes or inflammation. Our findings highlight the importance of dual management of residual inflammation and glycemic control toward improving healthy ageing in future practice.

6. Strengths

There are several strengths to this study. First, utilizing a nationally representative dataset with a large sample size (41,634) facilitates the generalization of the findings to a large population and enhances the external validity of our results. Second, utilizing rigorous statistical methods, we controlled for potential confounding variables, thereby enhancing the internal validity of the findings. Finally, multiple chained interpolations were employed to fill in missing covariate data, thereby enhancing the statistical efficiency.

7. Limitations

This study has limitations. First, as a cross-sectional study, it cannot establish causal conclusions because of inherent flaws. Therefore, a welldesigned cohort study is essential to address this limitation. Second, in exploring the relationship between glycemic status and biological ageing across different CRP levels in the general population, special populations such as children were excluded from this study. In future research, we intend to incorporate specific populations for further analysis. Third, a definition of distinguishing Type 1 Diabetes and T2D was lacking in this study. Fourth, although we collected data from 11 cycles of the NHANES from 1999 to 2000 to 2017-2020, we did not track the data of the same individuals. Therefore, it is difficult to conduct longitudinal research or trajectory analysis on NHANES data. Further longitudinal research is needed in the future. Finally, CRP measurements and analyses were conducted at different stages of the NHANES employing various analytical methods. Despite efforts to statistically harmonize, the results do not match the precision of simultaneous using consistent experimental methods.

8. Conclusions

These findings suggest that the joint presence of low-grade inflammation and dysfunctional glycemic metabolism conditions might be associated with higher odds of biological ageing acceleration and premature mortality. The dual management of residual inflammation and blood glucose in the people with diabetes may contribute to improvements in healthy ageing.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.dsx.2024.103050.

References

- Beard JR, Officer A, de Carvalho IA, Sadana R, Pot AM, Michel JP, et al. The World Report on ageing and health: a policy framework for healthy ageing. Lancet (N AM ED) 2016;387:2145–54.
- [2] Ahadi S, Zhou W, Schussler-Fiorenza RS, Sailani MR, Contrepois K, Avina M, et al. Personal aging markers and genotypes revealed by deep longitudinal profiling. Nat Med 2020;26:83–90.
- [3] Ferrucci L, Gonzalez-Freire M, Fabbri E, Simonsick E, Tanaka T, Moore Z, et al. Measuring biological aging in humans: a quest. Aging Cell 2020;19:e13080.
- [4] Rutledge J, Oh H, Wyss-Coray T. Measuring biological age using omics data. Nat Rev Genet 2022;23:715–27.
- [5] Horvath S. DNA methylation age of human tissues and cell types. Genome Biol 2013;14:R115.
- [6] Klemera P, Doubal S. A new approach to the concept and computation of biological age. Mech Ageing Dev 2006;127:240–8.
- [7] Levine ME, Lu AT, Quach A, Chen BH, Assimes TL, Bandinelli S, et al. An epigenetic biomarker of aging for lifespan and healthspan. Aging (Albany NY) 2018;10: 573–91.
- [8] 3. Prevention or delay of type 2 diabetes: Standards of medical Care in diabetes-2020. Diabetes Care 2020;43:S32–6.
- [9] Jeong H, Baek SY, Kim SW, Park EJ, Lee J, Kim H, et al. C reactive protein level as a marker for dyslipidemia, diabetes, and metabolic syndrome: results from the Korea National Health and Nutrition Examination Survey. BMJ Open 2019;9:e029861.
- [10] Soinio M, Marniemi J, Laakso M, Lehto S, Ronnemaa T. High-sensitivity C-reactive protein and coronary heart disease mortality in patients with type 2 diabetes: a 7year follow-up study. Diabetes Care 2006;29:329–33.
- [11] Akinboboye O, Williams JS, Garacci E, Egede LE. The relationship between C-Reactive protein and mortality in adults with diabetes: Influences of demographic characteristics, lifestyle behaviors, and medications. Nutr Metab Cardiovas 2022; 32:176–85.
- [12] Sharif S, Van der Graaf Y, Cramer MJ, Kapelle LJ, de Borst GJ, Visseren F, et al. Low-grade inflammation as a risk factor for cardiovascular events and all-cause mortality in patients with type 2 diabetes. Cardiovasc Diabetol 2021;20:220.
- [13] Chuengsamarn S, Rattanamongkolgul S, Sittithumcharee G, Jirawatnotai S. Association of serum high-sensitivity C-reactive protein with metabolic control and diabetic chronic vascular complications in patients with type 2 diabetes. Diabetes Metab Synd 2017;11:103–8.
- [14] Palmer AK, Xu M, Zhu Y, Pirtskhalava T, Weivoda MM, Hachfeld CM, et al. Targeting senescent cells alleviates obesity-induced metabolic dysfunction. Aging Cell 2019;18:e12950.

Diabetes & Metabolic Syndrome: Clinical Research & Reviews 18 (2024) 103050

- [15] Aguayo-Mazzucato C, Andle J, Lee TJ, Midha A, Talemal L, Chipashvili V, et al. Acceleration of beta cell aging Determines diabetes and Senolysis Improves disease Outcomes. Cell Metab. 2019;30:129–42.
- [16] Stepanova M, Rodriguez E, Birerdinc A, Baranova A. Age-independent rise of inflammatory scores may contribute to accelerated aging in multi-morbidity. Oncotarget 2015;6:1414–21.
- [17] Goto M, Hayata K, Chiba J, Matsuura M, Iwaki-Egawa S, Watanabe Y. Multiplex cytokine analysis of Werner syndrome. Intractable Rare Dis 2015;4:190–7.
- [18] Jurk D, Wilson C, Passos JF, Oakley F, Correia-Melo C, Greaves L, et al. Chronic inflammation induces telomere dysfunction and accelerates ageing in mice. Nat Commun 2014;2:4172.
- [19] Liu Y, Huang Z, Qiu H, Tang F, Liu F, Zhang Y, et al. The association between serum methylmalonic acid, cobalamin-related biomarkers, and long-term mortality risk in cancer survivors: a prospective cohort study. Am J Clin Nutr 2024;119:1122–32.
- [20] Wang S, Wang Y, Wan X, Guo J, Zhang Y, Tian M, et al. Cobalamin Intake and related biomarkers: Examining associations with mortality risk among adults with type 2 diabetes in NHANES. Diabetes Care 2022;45:276–84.
- [21] Wang S, Guo J, Liu X, Tian W, Zhang Y, Wang Y, et al. Sexual dimorphism in mitochondrial dysfunction and diabetes mellitus: evidence from a populationbased cohort study. Diabetol Metab Syndr. 2023;15:114.
- [22] Ong KL, Allison MA, Cheung BM, Wu BJ, Barter PJ, Rye KA. Trends in C-reactive protein levels in US adults from 1999 to 2010. Am J Epidemiol 2013;177:1430–42.
- [23] Pearson TA, Mensah GA, Alexander RW, Anderson JL, Cannon RR, Criqui M, et al. Markers of inflammation and cardiovascular disease: application to clinical and public health practice: a statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. Circulation 2003;107:499–511.
- [24] Kwon D, Belsky DW. A toolkit for quantification of biological age from blood chemistry and organ function test data: BioAge. Geroscience. 2021;43:2795–808.
- [25] Xu Y, Wang X, Belsky DW, McCall WV, Liu Y, Su S. Blunted rest-activity circadian rhythm is associated with increased rate of biological aging: an analysis of NHANES 2011-2014. J Gerontol A-Biol 2023;78:407–13.
- [26] Thomas A, Belsky DW, Gu Y. Healthy lifestyle behaviors and biological aging in the U.S. National health and nutrition examination surveys 1999-2018. J Gerontol A-Biol 2023;78:1535–42.
- [27] Wang S, Liu Y, Liu J, Tian W, Zhang X, Cai H, et al. Mitochondria-derived methylmalonic acid, a surrogate biomarker of mitochondrial dysfunction and oxidative stress, predicts all-cause and cardiovascular mortality in the general population. Redox Biol 2020;37:101741.
- [28] Wu L, Wu M, Zhao D, Chen S, Wang G, Xu L, et al. Elevated high-sensitivity Creactive protein levels increase the risk of new-onset cardiac conduction disorders. Cardiovasc Diabetol 2023;22:268.
- [29] Melbye H, Hvidsten D, Holm A, Nordbo SA, Brox J. The course of C-reactive protein response in untreated upper respiratory tract infection. Br J Gen Pract 2004;54:653–8.
- [30] Fang M, Wang D, Coresh J, Selvin E. Trends in diabetes treatment and control in U. S. Adults, 1999-2018. New Engl J Med 2021;384:2219–28.
- [31] Ko F, Abadir P, Marx R, Westbrook R, Cooke C, Yang H, et al. Impaired mitochondrial degradation by autophagy in the skeletal muscle of the aged female interleukin 10 null mice. Exp Gerontol 2016;73:23–7.
- [32] Chen L, Yin X, Zhao Y, Chen H, Tan T, Yao P, et al. Biological ageing and the risks of all-cause and cause-specific mortality among people with diabetes: a prospective cohort study. J Epidemiol Commun H 2022;76:771–8.
- [33] Liu Y, Feng X, Yang J, Sun I, Zhai G, Qianyun G, et al. Prognostic significance of HbA 1c level in Asian patients with prediabetes and coronary Artery disease. CVIA 2022;6(3).
- [34] McCarthy K, O'Halloran AM, Fallon P, Kenny RA, McCrory C. Metabolic syndrome accelerates epigenetic ageing in older adults: findings from the Irish Longitudinal Study on Ageing (TILDA). Exp Gerontol 2023;183:112314.

- [35] Cribb L, Hodge AM, Yu C, Li SX, English DR, Makalic E, et al. Inflammation and epigenetic aging are largely independent markers of biological aging and mortality. J Gerontol A-Biol. 2022;77:2378–86.
- [36] Engler-Chiurazzi EB, Russell AE, Povroznik JM, McDonald KO, Porter KN, Wang DS, et al. Intermittent systemic exposure to lipopolysaccharide-induced inflammation disrupts hippocampal long-term potentiation and impairs cognition in aging male mice. Brain Behav Immun 2023;108:279–91.
- [37] Shi L, Tan GS, Zhang K. Relationship of the serum CRP level with the Efficacy of metformin in the treatment of type 2 diabetes mellitus: a meta-analysis. J Clin Lab Anal 2016;30:13–22.
- [38] Wang J, Zhang F, Gao M, Wang Y, Song X, Li Y, et al. The systemic immune inflammatory index predicts No-reflow phenomenon after primary percutaneous coronary intervention in older patients with STEMI. Cardiovasc Innov App. 2023;7.
- [39] Grossmann V, Schmitt VH, Zeller T, Panova-Noeva M, Schulz A, Laubert-Reh D, et al. Profile of the immune and inflammatory response in individuals with prediabetes and type 2 diabetes. Diabetes Care 2015;38:1356–64.
- [40] Khin PP, Lee JH, Jun HS. A brief review of the mechanisms of beta-cell dedifferentiation in type 2 diabetes. Nutrients 2021;13.
- [41] Zatterale F, Longo M, Naderi J, Raciti GA, Desiderio A, Miele C, et al. Chronic adipose tissue inflammation linking obesity to insulin resistance and type 2 diabetes. Front Physiol 2019;10:1607.
- [42] Mesquida M, Drawnel F, Fauser S. The role of inflammation in diabetic eye disease. Semin Immunopathol 2019;41:427–45.
- [43] Lin KY, Hsih WH, Lin YB, Wen CY, Chang TJ. Update in the epidemiology, risk factors, screening, and treatment of diabetic retinopathy. J Diabetes Invest 2021; 12:1322–5.
- [44] Lemieux I, Pascot A, Prud'Homme D, Almeras N, Bogaty P, Nadeau A, et al. Elevated C-reactive protein: another component of the atherothrombotic profile of abdominal obesity. Arterioscl Throm Vas 2001;21:961–7.
- [45] Peng Y, Zhou G, Guo M, Cheng Z, Luo S, Guo Y. Inhibition of stimulator of interferon genes protects against myocardial ischemia-reperfusion injury in diabetic mice. CVIA 2023;8(1).
- [46] Wang H, Yang Z, Qi Y, Huang Y, Xiao L, Hao Y, et al. Addition of risk-enhancing factors improves risk assessment of atherosclerotic cardiovascular disease in middle-aged and older Chinese adults: findings from the Chinese multi-provincial cohort study. CVIA 2023;8(1).
- [47] Karthik D, Vijayakumar R, Pazhanichamy K, Ravikumar S. A proteomics approach to identify the differential protein level in cardiac muscle of the diabetic rat. Acta Biochim Pol 2014;61:285–93.
- [48] Hirakawa Y, Ninomiya T, Kiyohara Y, Murakami Y, Saitoh S, Nakagawa H, et al. Age-specific impact of diabetes mellitus on the risk of cardiovascular mortality: an overview from the evidence for Cardiovascular Prevention from Observational Cohorts in the Japan Research Group (EPOCH-Japan). J Epidemiol 2017;27:123–9.
- [49] Ni P, Yu M, Zhang R, Cheng C, He M, Wang H, et al. Dose-response association between C-reactive protein and risk of all-cause and cause-specific mortality: a systematic review and meta-analysis of cohort studies. Ann Epidemiol 2020;51: 20–7.
- [50] Li Y, Zhong X, Cheng G, Zhao C, Zhang L, Hong Y, et al. Hs-CRP and all-cause, cardiovascular, and cancer mortality risk: a meta-analysis. Atherosclerosis 2017; 259:75–82.
- [51] Liu J, Zhang Y, Lavie CJ, Tabung FK, Xu J, Hu Q, et al. Associations of C-reactive protein and fibrinogen with mortality from all-causes, cardiovascular disease and cancer among U.S. adults. Prev Med 2020;139:106044.
- [52] Liu Y, Yang D, Shi F, Wang F, Liu X, Wen H, et al. Association of serum 25(OH)D, cadmium, CRP with all-cause, cause-specific mortality: a prospective cohort study. Front Nutr 2022;9:803985.

Descargado para Lucia Angulo (lu.maru26@gmail.com) en National Library of Health and Social Security de ClinicalKey.es por Elsevier en junio 14, 2024. Para uso personal exclusivamente. No se permiten otros usos sin autorización. Copyright ©2024. Elsevier Inc. Todos los derechos reservados.