

Evidence for the effectiveness of immunologic therapies in women with subfertility and/or undergoing assisted reproduction

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Implantation is a critical step in the establishment of a successful pregnancy, depending on a complex immune-endocrine dialogue between the developing embryo and maternal endometrium. Research suggests that altered immunity in the maternal decidua results in implantation impairment and failure. Immunomodulatory drugs have, thus, been widely used in assisted conception to aid embryo implantation, despite an absence of consensus on their effectiveness and safety. We conducted a systematic review and meta-analysis of interventional studies investigating the use of immunomodulators in women undergoing assisted reproduction. Evidence was uncertain of an effect for most of the included interventions, owing to heterogeneous findings and a paucity of high-quality studies. For certain patient subgroups, however, the use of specific immunomodulatory therapies may offer some benefit. There is a need for further large randomized controlled trials to corroborate these findings. (Fertil Steril® 2022;117:1144–59. ©2022 by American Society for Reproductive Medicine.)

Key Words: Immunomodulatory drugs, implantation, infertility



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The number of patients undergoing assisted reproductive technology (ART) for fertility treatment continues to increase. Yet, despite advances over the past decades, most ART cycles do not result in a live birth. Even when chromosomally normal embryos are transferred, only approximately 50% will implant (1). Implantation failure remains the likely outcome of most in vitro fertilization (IVF) cycles, and approximately 10% of women undergoing IVF are thought to experience recurrent implantation failure (RIF), although there remains no universally agreed definition (2).

Critical to successful implantation is the adequate decidualization of the endometrium. Endometrial immune cells are key to the transformation of the decidua into a receptive mucosa that will permit embryo attachment and implantation. This highly regulated response involving an influx of innate immune cells, including macrophages, dendritic cells, and neutrophils, in addition to a large population of natural killer (NK) cells, results in an inflammatory environment at the time of conception and implantation (3). However, this response is dynamic; recent data indicate that by the early first

trimester, maternal immune inflammatory responses are restrained and modulated by complex and diverse signaling between fetal trophoblast cells and decidual immune cells (4).

Given the complexity of the implantation process, several factors may contribute to the failure of an embryo to implant, not least embryo aneuploidy. However, there has been much focus both in the scientific community and from patients on how the maternal immune response influences implantation and whether modulating maternal immunity can improve the ART success rates. For some patients and clinicians, the concept of embryo rejection has been an attractive one, on the basis of the knowledge that a specialized type of NK cell is the predominant immune cell in the peri-implantation endometrium. This concept of embryo rejection has often been given unscientific credibility simply because of its name.

Immunomodulators may be offered empirically to patients with a history of

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implantation failure where a clinician believes that there may be an immune mechanism underlying this or because specific immune cell testing has been undertaken and found to be abnormal. Blood NK cell testing (with or without associated cytotoxicity assays) is commonly offered, despite the fact that circulating NK cells are phenotypically and functionally different from their specialized endometrial counterparts (5) and their numbers are not correlated with endometrial NK cells (6). Neither blood nor endometrial NK cells have been robustly demonstrated to be associated with subfertility or RIF (7). After some studies reported alterations in the expression of T helper (Th)1 and Th2 cytokines in circulating T cells, tests including measurement of Th1/Th2 cytokine ratios have also found their way into clinical practice (8–10). Others have reported an association between RIF and altered ratios of cytokine messenger ribonucleic acid within the endometrium, including interleukin (IL)-15, IL-18, and tumor necrosis factor (TNF)-like weak inducer of apoptosis, hypothesizing that immunomodulators may improve the implantation rates in such patients (11). Additionally, some clinics test for autoantibodies, such as antinuclear, antiphospholipid, and thyroid autoantibodies. Although there may be associations with autoantibodies and infertility, causal links remain to be established (12).

Immunologic treatments have been used for decades in ART. Recent data have shown that up to 1 in 4 women undergoing IVF treatment may use add-on therapies, such as aspirin, heparin, or corticosteroids (13–16). Although less commonly used, other drugs, such as intralipid, intravenous immunoglobulin (IVIG), granulocyte colony-stimulating factor (G-CSF) and TNF alpha (TNF- α) inhibitors, are also administered to ART patients (15).

We have systematically reviewed the literature to evaluate the effectiveness and safety of immunomodulatory therapies used commonly in clinical practice and consider their potential mechanisms of action in the context of subfertility, ART, and implantation failure. We included randomized and nonrandomized interventional full-text studies where outcome data were reported per participant. Trials investigating participants with recurrent pregnancy loss were excluded. The primary effectiveness outcome considered in this review was the composite rate of ongoing pregnancy rate (OPR) or live birth rate (LBR) per participant. Ongoing pregnancy was defined as a viable intrauterine pregnancy at ≥ 12 weeks of gestation identified on ultrasound, and live birth was defined as the delivery of a live fetus after 22 weeks of pregnancy (17). Where possible, we performed meta-analysis of randomized controlled trials (RCTs) using a random-effects model and considered heterogeneity to be substantial where $I^2 > 50\%$. Subgroup analyses were also undertaken where data were available for women with previous implantation failure, known autoimmunity, and conditions hypothesized to be associated with an altered inflammatory response (e.g., elevated NK cell activity or numbers).

ASPIRIN

Low-dose aspirin is one of the most widely used adjuvants in ART (15). Aspirin inhibits cyclooxygenase (COX) in platelets,

preventing the conversion of arachidonic acid into prostaglandins and, thus, inhibiting thromboxane production (18). There are 2 types of COX: 1 and 2. By irreversibly inhibiting both COXs 1 and 2, aspirin may improve uterine and ovarian blood flow and reduce inflammation, all of which may enhance fertility or the success of ART (19, 20).

There have been numerous studies investigating aspirin for women with subfertility undergoing ART (21–23). We identified 12 studies that reported the OPR or LBR comparing the use of aspirin vs. a placebo or no intervention (21, 24–34) (Table 1). Most of the 6 nonrandomized studies reported uncertain findings (21, 24–27) except for that of Frattarelli et al. (28), where the use of aspirin was associated with a 20% increase in the LBR compared with that of placebo (relative risk [RR], 1.20; 95% confidence interval [CI], 1.04–1.38). The 6 RCTs that reported the OPR or LBR included 1,319 participants (29–34). Meta-analysis of these RCTs showed that aspirin probably makes little or no difference to the OPR or LBR (RR, 1.04; 95% CI, 0.81–1.33; 6 RCTs; $I^2 = 28\%$; moderate-certainty evidence) (Fig. 1 and Table 2). None of these studies reported significant adverse events although aspirin is known to be associated with gastrointestinal disturbances and bleeding. Albeit very commonly used by women undergoing ART, we did not find evidence that aspirin improved pregnancy outcomes. In addition, the subgroup analyses of good-prognosis patients, women with at least 1 previous implantation failure or a thin endometrium did not alter these findings, and therefore, recommending aspirin to patients undergoing ART to date has no evidence-based justification.

HEPARIN

In addition to its antithrombotic effects, there are several mechanisms by which heparin has been hypothesized to influence implantation. Both unfractionated heparin and low-molecular-weight heparin (LMWH) modulate insulin-like growth factors 1 and 2, which in turn may facilitate trophoblast invasion. Heparin may also increase the decidual expression and secretion of heparin-binding epidermal growth factor, which has been shown to promote an invasive trophoblast phenotype (35). In addition, heparin may promote trophoblast invasion by inducing matrix metalloproteinase activity with a concomitant suppression of tissue inhibitors of matrix metalloproteinases (36, 37).

Three cohort studies reported uncertain findings as to the effect of heparin on the LBR (21, 38, 39). Additionally, we identified 3 RCTs including 386 women that reported the LBR (40–42). Our meta-analysis did not demonstrate an increase in the LBR in patients receiving subcutaneous LMWH, although the quality of the evidence was assessed as being of very low certainty (RR, 1.55; 95% CI, 0.80–3.00; 3 RCTs; $I^2 = 51\%$) (Fig. 1 and Table 2).

Heparin is a drug with a good safety profile in pregnancy. Nevertheless, the evidence to date does not support its use routinely in women with subfertility. Importantly, heparin may cause bruising and discomfort around the injection site as well as bleeding, although in the studies included in this analysis, adverse events were reported in

TABLE 1

Characteristics of included studies reporting on the ongoing pregnancy or live birth rate.

Interventions	Study	Country	Design	Duration	Population	Inclusion criteria	Subgroup	Total analyzed	Outcome
Aspirin	Chen et al. (2017) (24)	People's Republic of China	Prospective cohort	January 2015 to December 2015	IVF	Age of <40 y; serum autoantibodies; normal karyotype; no infectious diseases; normal basal level of hormones; normal uterine anatomy	Autoimmunity	76	LBR
	Dirckx et al. (2009) (29)	Belgium	RCT	Unstated	IVF	First IVF/ICSI cycle	Unselected	201	LBR
	Frattarelli et al. (2006) (28)	USA	Retrospective cohort	January 2000 to August 2003	IVF	Egg recipient cycles	Low ovarian response	460	LBR
	Frattarelli et al. (2008) (25)	USA	Retrospective cohort	January 2000 to July 2006	IVF	Low-responding women	Low ovarian response	1,250	LBR
	Gizzo et al. (2014) (26)	Italy	Prospective cohort	January 2010 to December 2012	IVF	Age of 25–45 y; normal ovarian response; previous 1–2 failed IVF/ICSI cycles	Previous IF (≥ 1)	206	OPR
	Haapsamo et al. (2010) (30)	Finland	RCT	Unstated	IVF	Age of <40 y; <4 ovarian stimulations; no contraindications for aspirin	Good prognosis	487	LBR
	Hurst et al. (2005) (27)	USA	Retrospective cohort	1995 to 2001	IVF	Unstated—consecutive cycles	Unselected	316	LBR
	Lambers et al. (2009) (31)	The Netherlands	RCT	Unstated	IVF	Age of <39 y; FSH level of ≤ 10 IU/L; ≥ 1 previous failed IVF/ICSI treatment	Previous IF (≥ 1)	169	OPR
	Madani et al. (2019) (32)	Iran	RCT	May 2012 to February 2015	IVF	Age of <40 y; FET; no history of uterine surgery; no uterine disorders, endometriosis, RPL or contraindications to aspirin	Good prognosis	60	LBR
	Pakkila et al. (2005) (33)	Finland	RCT	2000 to 2003	IVF	Age of <40 y; <4 previous ovarian stimulation cycles; no contraindication for aspirin	Good prognosis	374	LBR
LMWH (subcutaneous)	Weckstein et al. (1997) (34)	USA	RCT	September 1993 to January 1995	IVF	Oocyte recipient cycles; thin endometrium (<8 mm); normal uterine cavity	Thin endometrium	28	LBR
	Berker et al. (2011) (38)	Turkey	Prospective cohort	June 2007 to October 2009	IVF	2 previous IFs; normal uterine; normal hormone profile; negative thrombophilia screening	Previous IF (≥ 1)	207	LBR
	Noci et al. (2011) (40)	Italy	RCT	May 2008 to December 2008	IVF	Age of <40 y; no congenital or acquired thrombophilia; no recent treatment with LMWH; no endocrine/hematologic abnormalities, chronic diseases, or tubal or uterine pathology interfering with embryo implantation	Good prognosis	153	LBR
	Qublan et al. (2008) (41)	Jordan	RCT	October 2004 to March 2006	IVF	Age of 19–35 y; ≥ 3 previous IVF failures; basal FSH level of ≤ 10 IU/L; BMI of 19–29 kg/m ² ; presence of both ovaries; ≥ 3 previous IVF failures; good-quality embryos for transfer; endometrial thickness of 8–14 mm; GnRH agonist down-regulation protocol	Previous IF (≥ 1)	83	LBR
	Urman et al. (2009) (42)	Turkey	RCT	January 2006 to May 2008	IVF	≥ 2 failed previously fresh embryo transfer cycles; age of ≤ 38 y; fresh ejaculate sperm to be used for ICSI; no hormonal, coagulation, or immunologic disorders; normal uterine cavity as assessed by hysteroscopy or saline infusion sonography; normal female and male peripheral karyotype	Previous IF (≥ 1)	150	LBR
Siristatidis et al. (2018) (39)	Siristatidis et al. (2018) (39)	Greece	Retrospective cohort	February 2012 to June 2017	IVF	Age of 25–40 y; BMI of 19–35 kg/m ² ; ≥ 2 failed fresh IVF/ICSI cycles followed by embryo transfer of at least 2 very-good-quality embryos on day 3; basal FSH level of ≤ 12 mIU/ml; absence of coagulation and/or autoimmune disorders	Previous IF (≥ 1)	230	LBR

Melo. Immunomodulators in clinical practice. *Fertil Steril* 2022.

TABLE 1

Continued.

Interventions	Study	Country	Design	Duration	Population	Inclusion criteria	Subgroup	Total analyzed	Outcome
Corticosteroids	Bider et al. (1996) (49)	Israel	RCT	February 1993 to December 1994	IVF	FET; normal uterus; tubal occlusion; normal hormone profile; normal SFA	Good prognosis	99	LBR
	Bider et al. (1999) (48)	Israel	Retrospective cohort	Unstated	IVF	PCOS; irregular menstrual cycles; failure of clomiphene citrate or hMG/hCG treatment; LH/FSH ratio of >2; high DHEAS and testosterone levels	Good prognosis	71	LBR
	Litwicka et al. (2015) (51)	Italy	RCT	January 2011 to April 2012	IVF	Age of <40 y; BMI of 18–29 kg/m ² ; thyroid autoimmunity; regular (21–35 days) menstrual cycles; presence of both ovaries; normal ovarian reserve; normal ovaries on ultrasound	Autoimmunity	60	LBR
	Moffitt et al. (1995) (50)	USA	RCT	January to September 1993	IVF	Unselected	Unselected	206	LBR
	Ozmen et al. (2018) (47)	Turkey	Retrospective cohort	January 2010 to June 2013	IVF	Age of 18–40 y; first IVF cycle only; normal baseline hormone profile; tubal or unexplained infertility; ICSI treatment	Good prognosis	964	LBR
	Turi et al. (2010) (52)	Italy	RCT	January 2006 to August 2008	IUI	Age of 20–38 y; positive antithyroid antibodies; ≤ 2 previous ART cycles; regular menstrual cycles of 24–35 days; BMI of 17–29 kg/m ² ; no treatment with clomiphene citrate or gonadotropins within 1 month before the date of recruitment; normal uterine cavity; bilateral tubal patency	Autoimmunity	48	LBR
	van Kasteren et al. (1999) (53)	the Netherlands	RCT	Unstated	OI-TI	Age of 18–40 y; idiopathic premature ovarian failure; normal karyotype; no history of radiotherapy or chemotherapy.	Low ovarian response	36	LBR
Aspirin plus corticosteroid G-CSF (intrauterine)	Revelli et al. (2008) (56)	Italy	RCT	October 2002 to April 2006	IVF	Age of ≤ 40 y; first IVF/ICSI cycle; only fresh embryo transfer cycles	Good prognosis	395	OPR
	Eftekhari et al. (2016) (67)	Iran	RCT	March to September 2015	IVF	Age of 18–40 y; normal endometrial thickness	Good prognosis	113	OPR
	Huang et al. (2020) (68)	People's Republic of China	RCT	December 2015 to July 2017	IVF	Age of ≤ 38 y; BMI of 18–24 kg/m ² ; primary infertility; previous IF (≥ 2); normal endometrial thickness (8–16 mm); 2 or more frozen embryos available	Previous IF (≥ 1)	104	LBR
	Jain et al. (2018) (69)	India	RCT	Unstated	IVF	Age of 21–38 y; BMI of 18.5–29.9 kg/m ² ; normal hormone profile	Good prognosis	150	OPR
	Kalem et al. (2020) (70)	Turkey	RCT	March 2016 to December 2017	IVF	Age of <40 y; ≥ 3 previous IFs; FSH level of <15 mIU/mL	Previous IF (≥ 1)	173	LBR
	Kunicki et al. (2017) (65)	Poland	Prospective cohort	October 2011 to October 2014	IVF	Thin endometrium; FET; previously cancelled IVF due to thin endometrium (<7 mm); previous treatment of unresponsive endometrium with oral/vaginal estradiol, sildenafil citrate, or aspirin; lack of contraindications for G-CSF treatment; own embryo FET	Thin endometrium	62	LBR
	Mao et al. (2020) (71)	People's Republic of China	RCT	March 2017 to May 2018	IVF	Age of 20–42 y; BMI of 20–28 kg/m ² ; FET; regular menstrual cycles; normal hormone profile; thin endometrium (<7 mm) or mild to moderate intrauterine adhesions; no contraindications for G-CSF treatment; no fibroids, polyps or PCOS; availability of at least 2 good-quality embryos	Thin endometrium	304	LBR
Immunomodulators	Xu et al. (2015) (66)	People's Republic of China	Prospective cohort	July 2012 to July 2013	IVF	Age of <40 y; FSH level of <10 IU/L; endometrial thickness of <7 mm; no uterine malformations; no contraindications for G-CSF treatment	Thin endometrium	66	LBR

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TABLE 1

Continued.									
Interventions	Study	Country	Design	Duration	Population	Inclusion criteria	Subgroup	Total analyzed	Outcome
G-CSF (subcutaneous) Intralipid	Arefi et al. (2018) (73)	Iran	RCT	May 2010 to October 2015	IVF	Age of 22–44 y; previous IF (>2)	Previous IF (≥1)	52	LBR
	Al-Zebeidi et al. (2020) (79)	Saudi Arabia	RCT	January 2015 to December 2016	IVF	Age of <42 y; BMI of <30 kg/m ² ; 3 previous IFs; normal uterine cavity; normal hormone and lipid profile	Previous IF (≥1)	142	LBR
	Singh et al. (2019) (78)	India	RCT	January 2017 to January 2018	IVF	Age of 20–40 y; BMI of 20–32 kg/m ² ; nondonor oocyte IVF; ≥ 1 previous IF; normal baseline hormonal profile; morphologically normal oocytes and embryos	Previous IF (≥1) and thin endometrium	102	LBR
IVIG	Ahmadi et al. (2017) (84)	Iran	Prospective cohort	February 2015 to March 2016	IVF	3 IFs; preconception Th1/Th2 ratio and NK cell frequency and activity elevation	Previous IF (≥1) and high inflammation	72	LBR
	Chernyshov et al. (2016) (85)	Ukraine	Prospective cohort	Unstated	IVF	Age of 23–35 y; RIF (≥3); good-quality embryos and quantity	Previous IF (≥1)	238	LBR
	Ho et al. (2020) (86)	People's Republic of China	Prospective cohort	January 2007 to October 2011	IVF	RIF (≥2 previous failures) of unknown etiology	Previous IF (≥1)	283	LBR
	Moraru et al. (2012) (82)	Spain	Prospective cohort	May 2005 to May 2011	IVF	RIF (definition unstated); high NK or NK-like cell level	Previous IF (≥1) and high inflammation	40	LBR
	Sher et al. (1998) (87)	USA	Prospective cohort	January 1992 to 1996	IVF	Age of <40 y; antithyroid antibodies; negative APL antibodies	Autoimmunity	82	LBR
r-LIF	Stephenson and Fluker (2000) (88)	Canada	RCT	March 1995 to July 1998	IVF	Age of 18–44 y; ≥ 2 previous failed fresh or frozen embryo transfers resulting in IF, biochemical pregnancy loss, or clinical miscarriage at ≤ 8 weeks of gestation	Previous IF (≥1)	51	LBR
	Brinsden et al. (2009) (91)	UK	RCT	Unstated	IVF	Age of 21–37 y; 2 or more failed ART cycles; BMI of 20–30 kg/m ² ; normal hormone profile; normal SFA	Previous IF (≥1)	150	LBR
PBMCs	Li et al. (2017) (95)	People's Republic of China	Unstated	July 2013 to March 2015	IVF	Previous IF (≥1)	Previous IF (≥1)	633	LBR
	Okitsu et al. (2011) (93)	Japan	Prospective cohort	May 2007 to February 2010	IVF	Previous IF (≥1)	Previous IF (≥1)	253	LBR
	Pourmoghadam et al. (2020) (96)	Iran	RCT	October 2017 to September 2018	IVF	Age of <45 y; BMI of <30 kg/m ² ; previous IF (≥3); primary infertility; regular menstrual cycles; normal SFA	Previous IF (≥1)	100	LBR
	Yoshioka et al. (2006) (94)	Japan	Unstated	Unstated	IVF	Previous IF (≥4); normal ovarian reserve (FSH level of <15 mIU/mL)	Previous IF (≥1)	35	LBR
	Yu et al. (2016) (97)	People's Republic of China	RCT	September 2013 to May 2014	IVF	Age of <35 y; RIF (≥3); normal ovarian reserve (FSH level of <15 mIU/mL)	Previous IF (≥1)	212	LBR
Aspirin ± subcutaneous heparin	Akhtar et al. (2013) (21)	UK	Retrospective cohort	January 2005 to September 2010	IVF	Age of 23–44 y; previous failed implantation (≥1)	Previous IF (≥1)	206	LBR
	Kutteh et al. (1997) (22)	USA	Prospective cohort	June 1995 to August 1996	IVF	Age of <42 y; first IVF cycle; FSH level of <20 mIU/mL, estradiol level of <50 pg/mL; normal uterine cavity	Good prognosis	191	OPR
IVIG plus TNF-α inhibitor	Sher et al. (1994) (23)	USA	Cohort - unstated	January 1992 to June 1994	IVF	Antiphospholipid antibody positivity; normal uterine cavity as observed by hysteroscopy	Autoimmunity	194	OPR
	Winger et al. (2009) (83)	UK	Retrospective cohort	May 2003 to December 2006	IVF	Age of <38 y; elevated Th1/Th2 cytokine ratio before index fresh IVF cycle; negative tuberculin skin test; good embryo development (≥ 5 day 3 embryos with ≥ 5 cells)	High inflammation	75	LBR

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TABLE 1

Interventions	Study	Country	Design	Duration	Population	Inclusion criteria	Total analyzed	Outcome
LMWH (subcutaneous) plus corticosteroid	Fawzy and El-Refaey (2014) (43)	Egypt	Prospective cohort	January 2008 to October 2012	IVF	Age of <39 y failed previously 1 or 2 implantations; fresh ejaculate fertile sperm used for ICSI; no hormonal disorders; normal uterine cavity	Previous IF (≥ 1)	295 OPR
TNF- α inhibitor	Sirsiatidis et al. (2018) (44)	Greece and Egypt	Retrospective cohort	February 2012 to July 2016	IVF	Age of <35 y; ≥ 2 failed IVF cycles; BMI of 19–35 kg/m ² ; FSH level of ≤ 15 mIU/ml; absence of coagulation and/or autoimmune disorders	Previous IF (≥ 1)	115 LBR
	Onalan et al. (2018) (98)	Turkey	Retrospective cohort (case-control analysis)	January 2014 to January 2015	IVF	Endometriomas	Good prognosis	66 LBR

Note: APL = antiphospholipid; ART = assisted reproductive technology; BMI = body mass index; DHEAS = dehydroepiandrosterone; FET = frozen embryo transfer; FSH = follicle-stimulating hormone; G-CSF = granulocyte colony-stimulating factor; GnRH = gonadotropin-releasing hormone; hCG = human chorionic gonadotropin; HMG = human menopausal gonadotropin; ICSC = intracytoplasmic sperm injection; IF = implantation failure; IU = international units; IVF = in vitro fertilization; IVG = intravenous immunoglobulin; LBR = live birth rate; LH = luteinizing hormone; LMWH = low-molecular-weight heparin; NK = natural killer; OI = ovulation induction; OPR = ongoing pregnancy rate; PRMC = peripheral blood mononuclear cell; PCOS = polycystic ovarian syndrome; RCT = randomized controlled trial; r-MLIF = recombinant human leukemia inhibitory factor; RF = recurrent implantation failure; RPL = recurrent pregnancy loss; SFA = seminal fluid analysis; Th = T helper; TI = timed intercourse; TNF = tumor necrosis factor.

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similar numbers in the heparin and no treatment/placebo groups (40, 41, 43, 44). Further trials are needed to determine whether patients with previous implantation failure and evidence of an acquired or inherited thrombophilia may benefit from LMWH.

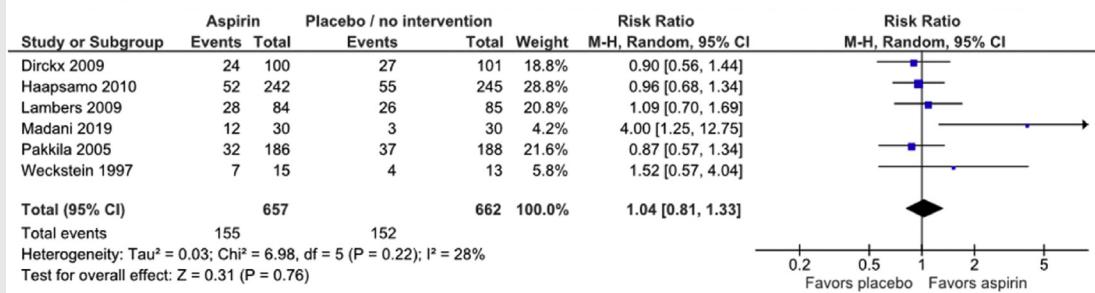
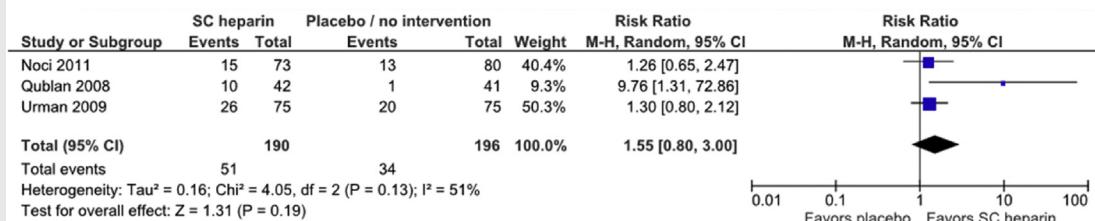
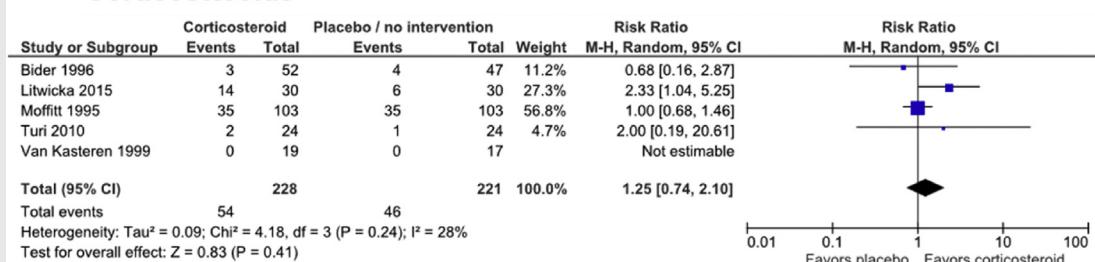
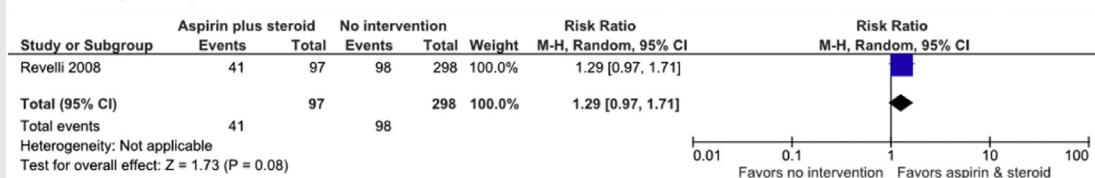
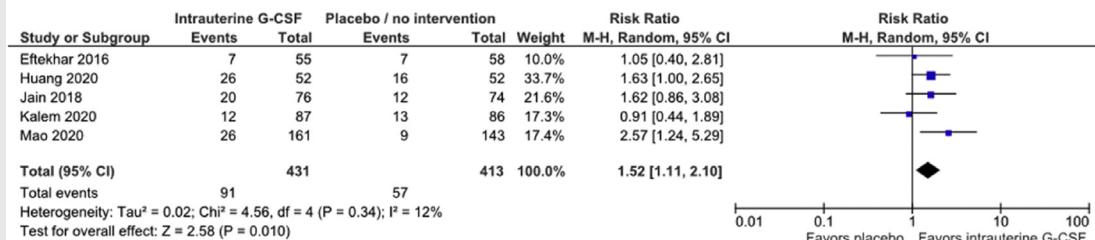
CORTICOSTEROIDS

Corticosteroids have been widely used in the context of IVF treatment. It has been proposed that steroids may improve the uterine immune environment by either modulating the uterine NK cell count or activity or altering the endometrial cytokine environment (45, 46). We identified 2 nonrandomized studies that investigated the effect of corticosteroids vs. no intervention on the LBR; both reported uncertain findings for this outcome (47, 48) (Table 1). In addition, our meta-analysis of 5 RCTs including 449 patients investigating corticosteroids vs. placebo or no treatment showed no clear evidence of an increased number of ongoing pregnancies or live births (RR, 1.25; 95% CI, 0.74–2.10; 5 RCTs; $I^2 = 0$; low-certainty evidence) (49–53) (Fig. 1 and Table 2).

An additional rationale for the use of corticosteroids has been to improve outcomes for women with subfertility who have autoantibodies. However, studies investigating the association between subfertility and autoantibodies have shown varying results (54). Although a recent meta-analysis has failed to demonstrate an association between thyroid autoantibodies and poor outcomes in patients undergoing IVF or intracytoplasmic sperm injection (55), our subgroup analysis including solely women with positive antithyroid antibodies showed that treatment with corticosteroids may result in improved LBR (RR, 2.29; 95% CI, 1.07–4.94; 2 RCTs; $I^2 = 0$; low-certainty evidence). Notably, however, only 108 women participated in these studies, and in 1 publication, participants underwent intrauterine insemination, not IVF (52).

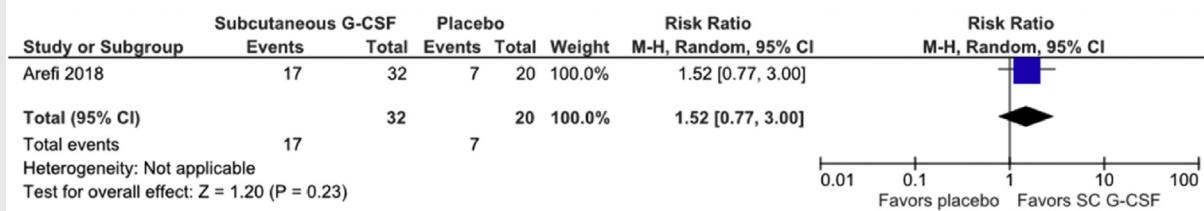
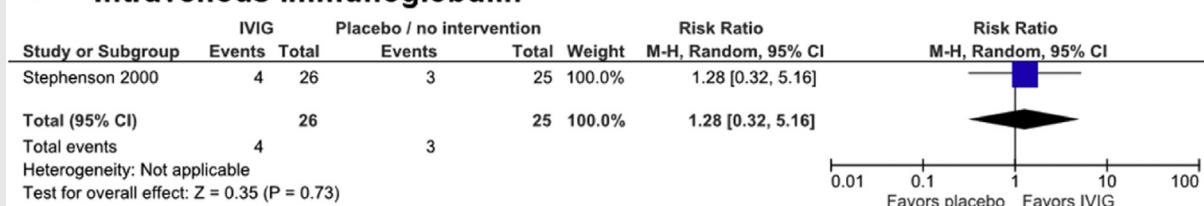
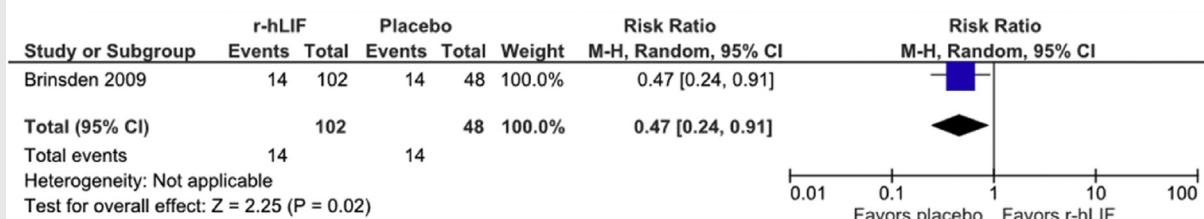
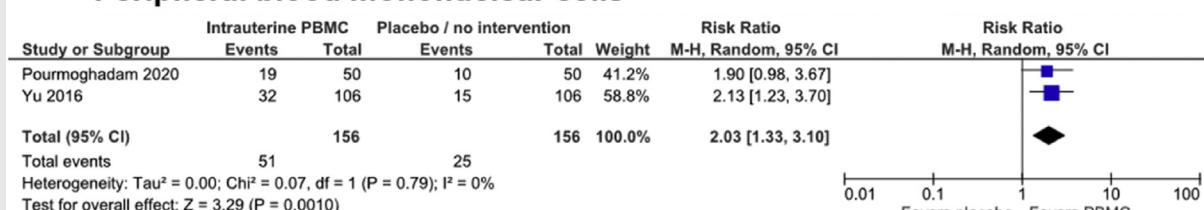
Corticosteroids are often prescribed in combination with other medications for women undergoing ART. We identified 1 RCT evaluating aspirin and corticosteroids vs. no intervention (56). The data were uncertain as to whether aspirin and corticosteroids in combination resulted in an increased OPR (RR, 1.29; 95% CI, 0.97–1.71; n = 395; low-certainty evidence) (Fig. 1 and Table 2).

Although generally considered to be safe in pregnancy, several studies have evaluated the possibility of teratogenicity with corticosteroid exposure. Initial evidence from the National Birth Defects Prevention Study in the United States suggested that corticosteroid exposure in the first trimester was associated with a significant, albeit small, increase in the risk of cleft lip and palate (57). However, more recent case-control data from the same registry including 1,577 children with cleft lip and palate no longer found this association (58). Adverse events reported from the trials included in this meta-analysis described typical effects of steroids, including insomnia and mild euphoria (53, 56). It is worth noting, however, that any use of corticosteroids beyond the first trimester of gestation should be restricted to women in whom the benefits are thought to outweigh the risks (e.g., asthma and autoimmune conditions). There is evidence suggesting that

FIGURE 1**A Aspirin****B Subcutaneous heparin****C Corticosteroids****D Aspirin plus corticosteroid****E Intrauterine G-CSF**

Meta-analysis of randomized controlled trials investigating different immunologic therapies in comparison with placebo or no intervention on the rate of ongoing pregnancy or live birth in women with subfertility and/or undergoing assisted conception. CI = confidence interval; df = degrees of freedom; G-CSF = granulocyte colony-stimulating factor; IVIG = intravenous immunoglobulin; M-H = Mantel-Haenszel test; PBMC = peripheral blood mononuclear cell; r-hLIF = recombinant human leukemia inhibitory factor; SC = subcutaneous.

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FIGURE 1**F Subcutaneous G-CSF****G Intralipid****H Intravenous immunoglobulin****I r-hLIF****J Peripheral blood mononuclear cells**

Continued

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TABLE 2

Summary of findings of included randomized controlled trials for the outcome of ongoing pregnancy or live birth.

Intervention	Anticipated absolute effects (95% CI)					Certainty of the evidence (GRADE)
	Risk with placebo or no intervention	Risk with intervention	Risk ratio (95% CI)	Number of participants (studies)		
Aspirin	230/1,000	239/1,000 (186–305)	RR, 1.04 (0.81–1.33)	1319 (6 RCTs)	⊕⊕⊕○	Moderate ^a
Subcutaneous heparin	173/1,000	269/1,000 (139–520)	RR, 1.55 (0.80–3.00)	386 (3 RCTs)	⊕○○○	Very low ^{a,b,c}
Corticosteroids	208/1,000	260/1,000 (154–437)	RR, 1.25 (0.74–2.10)	449 (5 RCTs)	⊕⊕○○	Low ^{a,b}
Aspirin plus corticosteroids	329/1,000	424/1,000 (319–562)	RR, 1.29 (0.97–1.71)	395 (1 RCT)	⊕⊕○○	Low ^{a,d}
Intrauterine G-CSF	138/1,000	210/1,000 (153–290)	RR, 1.52 (1.11–2.10)	844 (5 RCTs)	⊕⊕○○	Low ^{a,b}
Subcutaneous G-CSF	350/1,000	532/1,000 (269–1,000)	RR, 1.52 (0.77–3.00)	52 (1 RCT)	⊕○○○	Very low ^{a,b,e}
Intralipid	140/1,000	250/1,000 (133–469)	RR, 1.78 (0.95–3.34)	244 (2 RCTs)	⊕○○○	Very low ^{a,b,e}
IVIG	120/1,000	154/1,000 (38–619)	RR, 1.28 (0.32–5.16)	51 (1 RCT)	⊕⊕○○	Low ^{a,e}
r-hLIF	292/1,000	137/1,000 (70–265)	RR, 0.47 (0.24–0.91)	150 (1 RCT)	⊕⊕○○	Low ^{a,e}
PBMCs	160/1,000	325/1,000 (213–497)	RR, 2.03 (1.33–3.10)	312 (2 RCTs)	⊕○○○	Very low ^{a,b,e}

Note: Grading of Recommendations Assessment, Development and Evaluation (GRADE) Working Group grades of evidence

High certainty: we are very confident that the true effect lies close to that of the estimate of the effect.

Moderate certainty: we are moderately confident in the effect estimate: the true effect is likely to be close to the estimate of the effect, but there is a possibility that it is substantially different.

Low certainty: our confidence in the effect estimate is limited: the true effect may be substantially different from the estimate of the effect.

Very low certainty: we have very little confidence in the effect estimate: the true effect is likely to be substantially different from the estimate of effect.

CI = confidence interval; G-CSF = granulocyte colony-stimulating factor; IVIG = intravenous immunoglobulin; PBMCs = peripheral blood mononuclear cells; RCT = randomized controlled trial; r-hLIF = recombinant leukemia inhibitory factor; RR = risk ratio.

^a Downgraded once for serious imprecision due to the low number of events and/or wide confidence intervals.

^b Downgraded once for serious risk of bias in at least 1 of the included studies.

^c Downgraded once for serious inconsistency due to high heterogeneity.

^d Downgraded once for serious indirectness because the included study evaluated only participants with good prognosis, therefore limiting the applicability of its findings to subgroups.

^e Downgraded once for serious indirectness because the included study or studies evaluated only participants with recurrent implantation failure, thus limiting the applicability of the findings to other subgroups.

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repeated antenatal exposure to corticosteroids is associated with obstetric complications (e.g., increased risk of infection) as well as mental and behavioral disorders in children (59–61).

Evidence from RCTs to date does not support the generalized use of steroids in improving the OPR or LBR in women undergoing assisted conception. However, some groups of patients, including those with autoantibodies or autoimmune disease, may benefit from steroid treatment (51). There is a need for large RCTs to better define the effectiveness of corticosteroids to improve outcomes in women with infertility.

GRANULOCYTE COLONY-STIMULATING FACTOR

Granulocyte colony-stimulating factor is a cytokine produced by multiple cell types, including endothelium, epithelium, monocytes, macrophages, and bone marrow cells. Granulocyte colony-stimulating factor primarily stimulates the proliferation and survival of neutrophils (62). Recombinant G-CSF has been shown in an ex vivo model to alter endometrial gene expression, potentially influencing endometrial remodeling, cellular adhesion pathways, and local endometrial immune modulation (63). Intrauterine G-CSF treatment for patients with a thin endometrium was first reported in 2011

(64), and there have since then been a number of studies investigating its efficacy in the context of IVF.

Two cohort studies did not observe a difference in the LBR (65, 66) in patients treated with intrauterine G-CSF. In addition, we identified 5 RCTs including 844 participants treated with intrauterine G-CSF or a placebo/no intervention (67–71) (Table 1). Intrauterine G-CSF may result in a higher OPR or LBR than placebo or no intervention (RR, 1.52; 95% CI, 1.11–2.10; $I^2 = 12\%$), although the certainty of the evidence was found to be low (72) (Fig. 1 and Table 2). A subgroup analysis of women with a thin endometrium treated with intrauterine G-CSF suggested that this is the group in whom the increase in the LBR is most substantial (RR, 2.57; 95% CI, 1.24–5.29; 1 RCT; $n = 304$), although the evidence was judged to be of low certainty owing to the serious risk of bias and low number of events.

Although fatigue and bone and muscle pain are common side effects of G-CSF treatment, very few adverse events were reported in the included studies investigating the use of intrauterine G-CSF, presumably because the systemic dose of G-CSF is very low after intrauterine instillation (66, 74, 75).

The effect of subcutaneous G-CSF during ART treatment cycles has also been investigated, although we identified no

TABLE 3**Summary of recommendations on the use of immunotherapies in assisted reproduction.**

Intervention	ASRM	BFS	CFAS
Aspirin	"good evidence to recommend against the routine use of low-dose aspirin to improve the outcome of live birth in ART cycles in the general population"	"lack of proven efficacy for routine use of aspirin as an adjuvant in IVF cycles"	"aspirin should not be routinely offered in RIF" "In patients with thin endometrium undergoing embryo transfer cycles, we suggest against the use of aspirin to improve pregnancy rates" "empirical LMWH for RIF should be limited to research settings"
Heparin	—	"[the routine use of LMWH] in the wide population of women undergoing IVF treatment is not warranted" "[LMWH] should be carefully considered in women with thrombophilia"	
Corticosteroids	"good evidence to recommend against the routine use of corticosteroids during stimulation [and the implantation window] to improve the outcome of live birth in ART cycles in the general population" Additional evidence required on the effectiveness of corticosteroids in subpopulations	"lack of robust evidence to support the routine use of corticosteroids empirically as an adjuvant in IVF cycles" "limited evidence that corticosteroids may improve pregnancy rates in women undergoing conventional IVF and in the subgroup of women with autoimmunity or unexplained implantation failure"	"the use of glucocorticoids in RIF patients should be limited to research settings"
G-CSF	"insufficient evidence to routinely recommend for or against G-CSF administered locally or systemically to improve IVF outcomes"	—	"the use of G-CSF in RIF patients should be limited to research settings" "in patients with thin endometrium undergoing embryo transfer cycles, we suggest against the use of intrauterine infusion of G-CSF to improve pregnancy rates"
Intralipid	"insufficient evidence to routinely recommend intravenous fat emulsions for infertile women pursuing IVF" Additional evidence required on the effectiveness of intralipid in subpopulations	"lack of evidence to recommend intralipid infusion therapy as an adjuvant in IVF cycles. The use of intravenous lipids in this setting cannot therefore be supported"	"the use of intralipid in RIF patients should be limited to research settings"
IVIG	"insufficient evidence to recommend IVIG administration to improve IVF outcomes" Additional RCTs required to identify indications, risks, and benefits in subpopulations	"no convincing evidence for the use and safety of IVIg as adjuvants in women with recurrent implantation failure embarking on IVF. The use of IVIg in this setting cannot be supported"	"the use of immunotherapy in RIF patients should be limited to research settings"
r-hLIF PBMCs	— "insufficient evidence to recommend intrauterine infusion of autologous peripheral mononuclear cells before ET to improve IVF outcome"	—	—
TNF- α inhibitors	"insufficient evidence to recommend adalimumab treatment to improve IVF outcome"	"lack of evidence to indicate the effectiveness and safety of using anti-TNF- α agents as adjuvant in IVF cycles. The use of anti-TNF- α agents in this setting cannot therefore be supported"	—

Note: ART = assisted reproductive technology; ASRM = American Society for Reproductive Medicine; BFS = British Fertility Society; CFAS = Canadian Fertility & Andrology Society; ET = embryo transfer; G-CSF = granulocyte colony-stimulating factor; IVF = in vitro fertilization; IVIG = intravenous immunoglobulin; LMWH = low-molecular-weight heparin; PBMCs = peripheral blood mononuclear cells; RCT = randomized controlled trial; r-hLIF = recombinant leukemia inhibitory factor; RIF = recurrent implantation failure; TNF- α = tumor necrosis factor alpha.

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cohort studies and only 1 RCT including 52 patients that reported no difference in the LBR (73). Given the limited therapeutic options for treating women who have a persistently thin endometrium, further good-quality studies are needed to assess the efficacy of G-CSF.

INTRALIPID

Intralipid is an intravenous fat emulsion containing soybean oil, egg phospholipid, glycerin, and water and is licensed for parenteral nutrition. Its proposed mechanisms of action in the context of subfertility is the suppression of NK cell activity and proinflammatory cytokines (76). Interestingly, a recent study by Foyle et al. (77) investigated whether intralipids may modulate the adaptive immune response in women. The investigators identified no evidence of significant alterations to T cell populations (albeit in the blood rather than in the endometrium) after intralipid infusion. However, intralipid resulted in changes to circulating plasma cytokines, which the investigators suggested have the potential to enhance endometrial receptivity.

Despite the common use of intralipids in clinical practice, we identified only 2 RCTs including 244 patients in which the pooled effect of intralipid on the LBR was uncertain (RR, 1.78; 95% CI, 0.95–3.34; $I^2 = 26\%$) (Fig. 1 and Table 2). In both studies, women had experienced previous failed implantation after ART (78, 79). Clearly, further evidence for the efficacy of intralipid is required before using this treatment in patients outside of a research setting. No serious adverse events were described in these articles, yet at higher doses, intralipid has been associated with increased risks of infection, venous thromboembolism, fat embolism, acute kidney injury, and allergic reactions (80).

INTRAVENOUS IMMUNOGLOBULIN

Intravenous immunoglobulin is a pooled blood product licensed for several inflammatory and autoimmune conditions, such as chronic inflammatory demyelinating polyneuropathy, immune thrombocytopenia, and Kawasaki disease (81). We found a number of cohort studies assessing the effect of IVIG vs. no intervention. Most treated women considered to have high preconception Th1/Th2 ratios or a peripheral blood NK cell abnormality (82–84); others investigated participants who had previous implantation failure after ART treatment (85, 86), and 1 assessed IVIG use in women with antithyroid autoantibodies (87). Four of these studies found that treatment with IVIG led to a higher LBR (82, 84, 86, 87). The largest of these studies was a prospective analysis by Ho et al. (86) of 283 women with previous implantation failure (RR, 1.43; 95% CI, 1.05–1.95). In contrast, Chernyshov et al. (85) did not show a benefit to IVIG treatment, whereas the data from a retrospective cohort by Winger et al. (83) are difficult to interpret given multiple group comparators with a very small number of patients receiving no treatment. Despite the apparently encouraging results in some of these cohort studies, it is notable that we only identified 1 RCT evaluating 51 patients that demonstrated no clear effect of IVIG on the LBR (RR, 1.28; 95% CI, 0.32–5.16; low-certainty evidence) (88) (Fig. 1 and Table 2).

Intravenous immunoglobulin is commonly used as an adjuvant to ART, particularly in women with RIF or apparent immune dysregulation, despite the lack of high-quality evidence of a benefit in the OPR or LBR. Those studies that reported adverse events did not describe significant side effects (86, 88). In view of the positive data presented in several cohort studies, further large trials of this intervention are warranted, but the current evidence does not support routine clinical use in any specific patient population.

RECOMBINANT HUMAN LEUKEMIA INHIBITORY FACTOR

Leukemia inhibitory factor is a cytokine that was first identified as critical in mouse implantation (89). It has subsequently been investigated in humans, where it is expressed in the endometrium in the luteal phase of the menstrual cycle and thought to play a critical role in endometrial receptivity (90). Only 1 RCT has evaluated recombinant human leukemia inhibitory factor (r-hLIF) in a population of women with previous implantation failure undergoing ART (91). That study found that r-hLIF may result in a lower LBR (RR, 0.47; 95% CI, 0.24–0.91; n = 150; low-certainty evidence) (Fig. 1 and Table 2). This trial also reported more serious adverse events in the r-hLIF group, although the evidence was uncertain due to the relatively low numbers of patients in the study (RR, 4.24; 95% CI, 0.55–32.48; n = 150; low-certainty evidence). The apparent negative impact of r-hLIF on the LBR has undoubtedly discouraged others from further investigating this treatment.

PERIPHERAL BLOOD MONONUCLEAR CELLS

Peripheral blood mononuclear cells (PBMCs) include T and B lymphocytes, NK cells, as well as monocytes and dendritic cells. It has been hypothesized that intrauterine infusion of autologous PBMCs may enhance endometrial receptivity and improve the implantation rates. In most studies, PBMCs are cultured with human chorionic gonadotropin or corticotropin-releasing hormone before transfer (92). Some cohort studies have identified no difference in the LBR using intrauterine PBMCs (93), whereas others suggest an increase in the LBR (94). Most studies have investigated this treatment in patients with RIF, and 1 large cohort study demonstrated an increase in the LBR in participants with higher-order (≥ 4) implantation failure (95).

We identified 2 RCTs including 312 participants comparing intrauterine PBMC treatment with a placebo or no intervention that reported on the LBR (96, 97). In both studies, PBMCs were cocultured with human chorionic gonadotropin before transfer. Meta-analysis of these studies demonstrated a pooled RR of 2.03 (CI, 1.33–3.10; $I^2 = 0$) (Fig. 1), with anticipated absolute effects of achieving an LBR of 325 ongoing pregnancies or live births per 1,000 treated women compared with 160 patients receiving no treatment or placebo (Table 2). However, it must be noted that the certainty of the evidence was very low in these studies. Further, both trials investigated participants with previous implantation failure and, therefore, may not be generalizable to other subgroups of women with subfertility.

The putative mechanism of action of intrauterine PBMC therapy remains unclear. It is unlikely that the cells will breach the uterine mucosa, but *in vitro* culture of PBMCs activate cells and may stimulate the production of cytokines or factors beneficial to implantation (92). Endeavoring to identify which specific factors produced by cocultured PBMC may be ultimately more beneficial is the quest for studies investigating therapies for women with RIF.

TNF- α INHIBITORS

Tumor necrosis factor alpha is an inflammatory cytokine produced mainly by activated macrophages, T lymphocytes, and NK cells as well as nonimmune cells. Tumor necrosis factor alpha is synthesized within minutes of proinflammatory stress or injury, and chronic elevation of TNF- α levels is associated with several inflammatory conditions. Tumor necrosis factor alpha inhibitors are monoclonal antibody drugs licensed for the treatment of several of these conditions, including severe rheumatoid arthritis and other arthropathies, as well as inflammatory bowel disease. Some studies have described the ratios of Th1 and Th2 cytokines as being discordant or abnormal in the context of subfertility and RIF (8–10). This has led to the hypothesis that these medications may be beneficial, particularly in ART patients with RIF. Our systematic review identified no RCTs evaluating anti-TNF- α therapies on the OPR or LBR and found only 2 cohort studies where the data were expressed per participant (83, 98) (Table 1). In the study by Winger et al. (83), patients undergoing ART who were identified by the investigators as having an abnormal TNF- α /IL-10 ratio were administered adalimumab alone, IVIG alone, a combination of adalimumab and IVIG, or no treatment. The treatment was based on patient choice, and only 5 patients received no treatment. Although the investigators identified a higher LBR using the combination therapy than that in no treatment, the study design precludes drawing meaningful conclusions from these data. In another small retrospective study, Onalan et al. (98) evaluated the effectiveness of etanercept in women with endometrioma undergoing ART and found no difference in the LBR. Tumor necrosis factor alpha inhibitors should not be recommended to patients outside the context of a well-designed RCT. Most patients using this medication experience only mild side effects; however, biologic medications can result in more serious adverse events, such as an increased risk of severe infections (99).

CONCLUSION

Despite the widespread use of immunomodulatory drugs in clinical practice, this systematic review of the literature once again demonstrates a real lack of good-quality evidence for the use of immunologic treatments in women with subfertility or who are undergoing ART. In both research and clinical practice, patient selection remains controversial. Some clinicians administer immunologic therapies exclusively to women with known risk factors for implantation failure, for

example, RIF or the existence of concomitant autoimmune conditions, such as antiphospholipid syndrome (100). Others apply less stringent criteria and use these drugs in the absence of known pathology that may impair implantation. Table 3 summarizes existing recommendations from different international societies on the use of immunotherapies in ART, highlighting that for most interventions, the evidence is scarce (101–104).

The use of immunologic treatments often accrues significant financial burden to patients and their partners (14, 105). Commonly prescribed drugs, such as aspirin and LMWH, are relatively inexpensive, yet the added cost of other immunotherapies can be substantial. For example, 1 injection of recombinant G-CSF solution (300 mcg) costs upward of US\$360, with some centers administering more than 1 injection per treatment cycle (106, 107). Treatment with IVIG (approximately 1 g/kg) is even more expensive and can add up to US\$6,000 to the cost of 1 ART cycle (108). It is also worth noting that IVIG is a blood product derived from the plasma of healthy donors and, thus, constitutes a finite resource (109). Clinicians have a duty to thoroughly counsel patients on the absence of conclusive evidence demonstrating a benefit of these therapies in women undergoing assisted conception.

Crucially, embryo aneuploidy represents the leading cause of implantation failure, abnormal implantation, and miscarriage, especially with advancing female age. In cases of embryo aneuploidy, the use of immunotherapy drugs remains futile (110). However, none of the included studies in this review restricted their samples to euploid embryos. There is a need for additional trials investigating immunomodulatory treatments in women having ART with embryos known to be chromosomally normal.

Recent studies have begun to elucidate the very complex cellular composition of the human endometrium and early decidua, including multiple subtypes of NK cells with distinctive immunomodulatory profiles (4, 111). In light of these data, current testing for immune dysfunction is likely to be far too simplistic. For example, the population of CD3+CD56+ endometrial cells identified and quantified by several clinical tests will in fact encompass a diverse range of cells, including subtypes of innate lymphoid cells and not just NK cells (112). Further, there is emerging evidence on various other etiologic pathways in RIF, including the role of killer immunoglobulin-like receptors/human leukocyte antigen incompatibility. However, our searches found no interventional studies evaluating the effectiveness and safety of immunotherapy drugs in such cases (113).

Despite the lack of certainty in the existing evidence, it would be premature to dismiss a future for immunologic treatments to improve outcomes for patients undergoing ART. Perhaps before embarking on major RCTs, clinicians and researchers need to agree on what pathology is being treated. The first challenge for the scientific community is to define tests that identify a true population of patients with an immune-mediated pathology and then undertake well-designed RCTs in suitably phenotyped patients who have the potential to benefit from immune treatments. In an era of personalized

medicine, there remain the scope and opportunity for reproductive immunology to improve outcomes for patients.



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REFERENCES

1. Munne S, Kaplan B, Frattarelli JL, Child T, Nakhuda G, Shamma FN, et al. Pre-implantation genetic testing for aneuploidy versus morphology as selection criteria for single frozen-thawed embryo transfer in good-prognosis patients: a multicenter randomized clinical trial. *Fertil Steril* 2019;112:1071–9.e7.
2. Cimadomo D, Craciunas L, Vermeulen N, Vornstein K, Toth B. Definition, diagnostic and therapeutic options in recurrent implantation failure: an international survey of clinicians and embryologists. *Hum Reprod* 2021;36:305–17.
3. Mor G, Cardenas I, Abrahams V, Guller S. Inflammation and pregnancy: the role of the immune system at the implantation site. *Ann N Y Acad Sci* 2011;1221:80–7.
4. Vento-Tormo R, Efremova M, Botting RA, Turco MY, Vento-Tormo M, Meyer KB, et al. Single-cell reconstruction of the early maternal-fetal interface in humans. *Nature* 2018;563:347–53.
5. Huhn O, Zhao X, Esposito L, Moffett A, Colucci F, Sharkey AM. How do uterine natural killer and innate lymphoid cells contribute to successful pregnancy? *Front Immunol* 2021;12:607669.
6. Laird SM, Mariee N, Wei L, Li TC. Measurements of CD56+ cells in peripheral blood and endometrium by flow cytometry and immunohistochemical staining in situ. *Hum Reprod* 2011;26:1331–7.
7. Seshadri S, Sunkara SK. Natural killer cells in female infertility and recurrent miscarriage: a systematic review and meta-analysis. *Hum Reprod Update* 2014;20:429–38.
8. Kalu E, Bhaskaran S, Thum MY, Vishwanatha R, Croucher C, Sherriff E, et al. Serial estimation of Th1:th2 cytokines profile in women undergoing in-vitro fertilization-embryo transfer. *Am J Reprod Immunol* 2008;59:206–11.
9. Kwak-Kim JY, Chung-Bang HS, Ng SC, Ntrivalas EI, Mangubat CP, Beaman KD, et al. Increased T helper 1 cytokine responses by circulating T cells are present in women with recurrent pregnancy losses and in infertile women with multiple implantation failures after IVF. *Hum Reprod* 2003;18:767–73.
10. Kuroda K, Nakagawa K, Horikawa T, Moriyama A, Ojiri Y, Takamizawa S, et al. Increasing number of implantation failures and pregnancy losses associated with elevated Th1/Th2 cell ratio. *Am J Reprod Immunol* 2021;86:e13429.
11. Ledee N, Petitbarat M, Prat-Ellenberg L, Dray G, Cassuto GN, Chevrier L, et al. Endometrial immune profiling: a method to design personalized care in assisted reproductive medicine. *Front Immunol* 2020;11:1032.
12. Deroux A, Dumestre-Perard C, Dunand-Faure C, Bouillet L, Hoffmann P. Female infertility and serum auto-antibodies: a systematic review. *Clin Rev Allergy Immunol* 2017;53:78–86.
13. Lensen S, Shreeve N, Barnhart KT, Gibrel A, Ng EHY, Moffett A. In vitro fertilization add-ons for the endometrium: it doesn't add-up. *Fertil Steril* 2019;112:987–93.
14. Harper J, Jackson E, Sermon K, Aitken RJ, Harbottle S, Mocanu E, et al. Adjuncts in the IVF laboratory: where is the evidence for 'add-on' interventions? *Hum Reprod* 2017;32:485–91.
15. Lensen S, Hammarberg K, Polyakov A, Wilkinson J, Whyte S, Peate M, et al. How common is add-on use and how do patients decide whether to use them? A national survey of IVF patients. *Hum Reprod* 2021;36:1854–61.
16. Armstrong SC, Lensen S, Vaughan E, Wainwright E, Peate M, Balen AH, et al. VALUE study: a protocol for a qualitative semi-structured interview study of IVF add-ons use by patients, clinicians and embryologists in the UK and Australia. *BMJ Open* 2021;11:e047307.
17. Zegers-Hochschild F, Adamson GD, Dyer S, Racowsky C, de Mouzon J, Sokol R, et al. The International Glossary on Infertility and Fertility Care, 2017. *Fertil Steril* 2017;108:393–406.
18. Vane JR, Botting RM. The mechanism of action of aspirin. *Thromb Res* 2003;110:255–8.
19. Haapsamo M, Martikainen H, Rasanen J. Low-dose aspirin and uterine haemodynamics on the day of embryo transfer in women undergoing IVF/ICSI: a randomized, placebo-controlled, double-blind study. *Hum Reprod* 2009;24:861–6.
20. Rubinstein M, Marazzi A, Polak de Fried E. Low-dose aspirin treatment improves ovarian responsiveness, uterine and ovarian blood flow velocity, implantation, and pregnancy rates in patients undergoing in vitro fertilization: a prospective, randomized, double-blind placebo-controlled assay. *Fertil Steril* 1999;71:825–9.
21. Akhtar MA, Eljabu H, Hopkisson J, Raine-Fenning N, Quenby S, Jayaprakasan K. Aspirin and heparin as adjuvants during IVF do not improve live birth rates in unexplained implantation failure. *Reprod Biomed Online* 2013;26:586–94.
22. Kutteh WH, Yetman DL, Chantilis SJ, Crain J. Effect of antiphospholipid antibodies in women undergoing in-vitro fertilization: role of heparin and aspirin. *Hum Reprod* 1997;12:1171–5.
23. Sher G, Feinman M, Zouves C, Kuttner G, Maassarani G, Salem R, et al. High fecundity rates following in-vitro fertilization and embryo transfer in antiphospholipid antibody seropositive women treated with heparin and aspirin. *Hum Reprod* 1994;9:2278–83.
24. Chen X, Mo M-L, Huang C-Y, Diao L-H, Li G-G, Li Y-Y, et al. Association of serum autoantibodies with pregnancy outcome of patients undergoing first IVF/ICSI treatment: a prospective cohort study. *J Reprod Immunol* 2017;122:14–20.
25. Frattarelli JL, McWilliams GDE, Hill MJ, Miller KA, Scott RT Jr. Low-dose aspirin use does not improve in vitro fertilization outcomes in poor responders. *Fertil Steril* 2008;89:1113–7.
26. Gizzo S, Capuzzo D, Zicchina C, Di Gangi S, Coronella ML, Andrisani A, et al. Could empirical low-dose-aspirin administration during IVF cycle affect both the oocytes and embryos quality via COX 1-2 activity inhibition? *J Assist Reprod Genet* 2014;31:261–8.
27. Hurst BS, Bhojwani JT, Marshburn PB, Papadakis MA, Loeb TA, Matthews ML. Low-dose aspirin does not improve ovarian stimulation, endometrial response, or pregnancy rates for in vitro fertilization. *J Exp Clin Assist Reprod* 2005;2:8.
28. Frattarelli JL, Miller BT, Scott RT Jr. Adjuvant therapy enhances endometrial receptivity in patients undergoing assisted reproduction. *Reprod Biomed Online* 2006;12:722–9.
29. Dirckx K, Cabri P, Merien A, Galajdova L, Gerris J, Dhont M, et al. Does low-dose aspirin improve pregnancy rate in IVF/ICSI? A randomized double-blind placebo controlled trial. *Hum Reprod* 2009;24:856–60.
30. Haapsamo M, Martikainen H, Tinkanen H, Heinonen S, Nuoja-Huttunen S, Rasanen J. Low-dose aspirin therapy and hypertensive pregnancy complications in unselected IVF and ICSI patients: a randomized, placebo-controlled, double-blind study. *Hum Reprod* 2010;25:2972–7.
31. Lambers MJ, Hoozemans DA, Schats R, Homburg R, Lambalk CB, Hompes PG. Low-dose aspirin in non-tubal IVF patients with previous failed conception: a prospective randomized double-blind placebo-controlled trial. *Fertil Steril* 2009;92:923–9.
32. Madani T, Ahmadi F, Jahangiri N, Bahmanabadi A, Bagheri Lankarani N. Does low-dose aspirin improve pregnancy rate in women undergoing frozen-thawed embryo transfer cycle? A pilot double-blind, randomized placebo-controlled trial. *J Obstet Gynaecol Res* 2019;45:156–63.
33. Pakkila M, Ralsanen J, Heinonen S, Tinkanen H, Tuomiavaara L, Makikallio K, et al. Low-dose aspirin does not improve ovarian responsiveness or pregnancy rate in IVF and ICSI patients: a randomized, placebo-controlled double-blind study. *Hum Reprod* 2005;20:2211–4.
34. Weckstein LN, Jacobson A, Galen D, Hampton K, Hammel J. Low-dose aspirin for oocyte donation recipients with a thin endometrium: prospective, randomized study. *Fertil Steril* 1997;68:927–30.

35. Quaranta M, Erez O, Mastrolia SA, Koifman A, Leron E, Eshkoli T, et al. The physiologic and therapeutic role of heparin in implantation and placentation. *PeerJ* 2015;3:e691.
36. Arpino PA, Goeller AJ, Fatalo A, Van Cott EM. Evaluation of 2 nomogram-based strategies for dosing argatroban in patients with known or suspected heparin-induced thrombocytopenia. *Clin Appl Thromb Hemost* 2015;21:260–5.
37. Di Simone N, Di Nicuolo F, Sanguineti M, Ferrazzani S, D'Alessio MC, Castellani R, et al. Low-molecular weight heparin induces in vitro trophoblast invasiveness: role of matrix metalloproteinases and tissue inhibitors. *Placenta* 2007;28:298–304.
38. Berker B, Kahraman K, Taskin EA, Atabekoglu C, Sonmezler M, Taskin S. The role of low-molecular-weight heparin in recurrent implantation failure: a prospective, quasi-randomized, controlled study. *Fertil Steril* 2011;95: 2499–502.
39. Siristatidis C, Dafopoulos K, Salamalekis G, Galazios G, Christoforidis N, Moustakarias T, et al. Administration of low-molecular-weight heparin in patients with two or more unsuccessful IVF/ICSI cycles: a multicenter cohort study. *Gynecol Endocrinol* 2018;34:747–51.
40. Noci I, Milanini MN, Fuzzi B, Ruggiero M, Papini F, Artini PG. Effect of dalteparin sodium administration on IVF outcome in non-thrombophilic young women: a pilot study. *Reprod Biomed Online* 2011;22:615–20.
41. Qublan H, Amarin Z, Dabbas M, Farraj AE, Beni-Merei Z, Al-Akash H, et al. Low-molecular-weight heparin in the treatment of recurrent IVF-ET failure and thrombophilia: a prospective randomized placebo-controlled trial. *Hum Fertil (Camb)* 2008;11:246–53.
42. Urman B, Ata B, Yakin K, Alatas C, Aksoy S, Mercan R, et al. Luteal phase empirical low molecular weight heparin administration in patients with failed ICSI embryo transfer cycles: a randomized open-labeled pilot trial. *Hum Reprod* 2009;24:1640–7.
43. Fawzy M, El-Refaeey AA. Does combined prednisolone and low molecular weight heparin have a role in unexplained implantation failure? *Arch Gynecol Obstet* 2014;289:677–80.
44. Siristatidis C, Dafopoulos K, El-Khayat W, Salamalekis G, Anifandis G, Vranta T, et al. Administration of prednisolone and low molecular weight heparin in patients with repeated implantation failures: a cohort study. *Gynecol Endocrinol* 2018;34:136–9.
45. Ledee N, Prat-Ellenberg L, Petitbarat M, Chevrier L, Simon C, Irani EE, et al. Impact of prednisone in patients with repeated embryo implantation failures: beneficial or deleterious? *J Reprod Immunol* 2018;127:11–5.
46. Cooper S, Laird SM, Mariee N, Li TC, Metwally M. The effect of prednisolone on endometrial uterine NK cell concentrations and pregnancy outcome in women with reproductive failure. A retrospective cohort study. *J Reprod Immunol* 2019;131:1–6.
47. Ozmen B, Pabuccu EG, Sukur YE, Ulubasoglu H, Ates C, Sonmezler M, et al. A retrospective comparative study of prednisolone use in antagonist co-treated assisted reproductive technology cycles for patients with good prognosis. *Turk J Obstet Gynecol* 2018;15:147–51.
48. Bider D, Hourvitz A, Tur Kaspa I, Dirnfeld M, Dor J. Dexamethasone supplementation to gonadotropin stimulation for in vitro fertilization in polycystic ovarian disease. *J Assist Reprod Genet* 1999;16:233–5.
49. Bider D, Amoday I, Yonesh M, Yemini Z, Mashiach S, Dor J. Glucocorticoid administration during transfer of frozen-thawed embryos: a prospective, randomized study. *Fertil Steril* 1996;66:154–6.
50. Moffitt D, Queenan JT Jr, Veeck LL, Schoolcraft W, Miller CE, Muasher SJ. Low-dose glucocorticoids after in vitro fertilization and embryo transfer have no significant effect on pregnancy rate. *Fertil Steril* 1995;63:571–7.
51. Litwicka K, Arrivi C, Varricchio MT, Mencacci C, Greco E. In women with thyroid autoimmunity, does low-dose prednisolone administration, compared with no adjuvant therapy, improve in vitro fertilization clinical results? *J Obstet Gynaecol Res* 2015;41:722–8.
52. Turi A, Giannibilo SR, Zanconi S, Mascetti A, Tranquilli AL. Preconception steroid treatment in infertile women with antithyroid autoimmunity undergoing ovarian stimulation and intrauterine insemination: a double-blind, randomized, prospective cohort study. *Clin Ther* 2010;32:2415–21.
53. van Kasteren YM, Braat DDM, Hemrika DJ, Lambalk CB, Rekers-Mombarg LTM, von Blomberg BM, et al. Corticosteroids do not influence ovarian responsiveness to gonadotropins in patients with premature ovarian failure: a randomized, placebo-controlled trial. *Fertil Steril* 1999; 71:90–5.
54. Khizroeva J, Nalli C, Bitsadze V, Lojacono A, Zatti S, Andreoli L, et al. Infertility in women with systemic autoimmune diseases. *Best Pract Res Clin Endocrinol Metab* 2019;33:101369.
55. Venables A, Wong W, Way M, Homer HA. Thyroid autoimmunity and IVF/ICSI outcomes in euthyroid women: a systematic review and meta-analysis. *Reprod Biol Endocrinol* 2020;18:120.
56. Revelli A, Dolfin E, Gennarelli G, Lantieri T, Massobrio M, Holte JG, et al. Low-dose acetylsalicylic acid plus prednisolone as an adjuvant treatment in IVF: a prospective, randomized study. *Fertil Steril* 2008;90:1685–91.
57. Carmichael SL, Shaw GM, Ma C, Werler MM, Rasmussen SA, Lammer EJ, et al. Maternal corticosteroid use and orofacial clefts. *Am J Obstet Gynecol* 2007;197:585, e1–7; discussion 683–4, e1–7.
58. Skuladottir H, Wilcox AJ, Ma C, Lammer EJ, Rasmussen SA, Werler MM, et al. Corticosteroid use and risk of orofacial clefts. *Birth Defects Res A Clin Mol Teratol* 2014;100:499–506.
59. Räikkönen K, Gissler M, Kajantie E. Associations between maternal antenatal corticosteroid treatment and mental and behavioral disorders in children. *J Am Med Assoc* 2020;323:1924–33.
60. Mariotti V, Marconi AM, Pardi G. Undesired effects of steroids during pregnancy. *J Matern Fetal Neonatal Med* 2004;16(Suppl 2):5–7.
61. Kemp MW, Newnham JP, Challis JG, Jobe AH, Stock SJ. The clinical use of corticosteroids in pregnancy. *Hum Reprod Update* 2016;22:240–59.
62. Mehta HM, Corey SJ, CSFG-. the guardian of granulopoiesis. *Semin Immunol* 2021;54:101515.
63. Rahmati M, Petitbarat M, Dubanchet S, Bensussan A, Chaouat G, Ledee N. Granulocyte-colony stimulating factor related pathways tested on an endometrial ex-vivo model. *PLoS One* 2014;9:e102286.
64. Gleicher N, Vidali A, Barad DH. Successful treatment of unresponsive thin endometrium. *Fertil Steril* 2011;95:2123, e13–7.
65. Kunicki M, Lukaszuk K, Liss J, Skowronska P, Szczypanska J. Granulocyte colony stimulating factor treatment of resistant thin endometrium in women with frozen-thawed blastocyst transfer. *Syst Biol Reprod Med* 2017;63:49–57.
66. Xu B, Zhang Q, Hao J, Xu D, Li Y. Two protocols to treat thin endometrium with granulocyte colony-stimulating factor during frozen embryo transfer cycles. *Reprod Biomed Online* 2015;30:349–58.
67. Eftekhar M, Hosseini Sadat R, Baradaran R, Naghshineh E. Effect of granulocyte colony stimulating factor (G-CSF) on IVF outcomes in infertile women: an RCT. *Int J Reprod Biomed* 2016;14:341–6.
68. Huang P, Yao C, Wei L, Lin Z. The intrauterine perfusion of granulocyte-colony stimulating factor (G-CSF) before frozen-thawed embryo transfer in patients with two or more implantation failures. *Hum Fertil* 2020;1–5.
69. Jain S, Mahey R, Malhotra N, Kalaivani M, Sangeeta P, Bhatt A, et al. Effect of intrauterine perfusion of granulocyte colony-stimulating factor on endometrial parameters and in vitro fertilization outcome in women undergoing in vitro fertilization/intracytoplasmic sperm injection cycles: a randomized controlled trial. *J Hum Reprod Sci* 2018;11:254–60.
70. Kalem Z, Namli Kalem M, Bakirarar B, Kent E, Makrigianakis A, Gurgan T. Intrauterine G-CSF administration in recurrent implantation failure (RIF): an Rct. *Sci Rep* 2020;10:5139.
71. Mao X, Zhang J, Cai R, Tao Y, Gao H, Kuang Y, et al. Therapeutic role of granulocyte macrophage colony-stimulating factor (GM-CSF) in patients with persistent thin endometrium: a prospective and randomized study. *Int J Gynaecol Obstet* 2020;150:194–9.
72. Guyatt GH, Oxman AD, Vist GE, Kunz R, Falck-Ytter Y, Alonso-Coello P, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ* 2008;336:924–6.
73. Arefi S, Fazeli E, Esfahani M, Borhani N, Yamini N, Hosseini A, et al. Granulocyte-colony stimulating factor may improve pregnancy outcome in patients with history of unexplained recurrent implantation failure: an RCT. *Int J Reprod Biomed* 2018;16:299–304.
74. Barad DH, Yu Y, Kushnir VA, Shohat-Tal A, Lazzaroni E, Lee HJ, et al. A randomized clinical trial of endometrial perfusion with granulocyte colony-

- stimulating factor in in vitro fertilization cycles: impact on endometrial thickness and clinical pregnancy rates. *Fertil Steril* 2014;101:710–5.
75. Eftekhari M, Sayadi M, Arabjahani F. Transvaginal perfusion of G-CSF for infertile women with thin endometrium in frozen ET program: a non-randomized clinical trial. *Iran J Reprod Med* 2014;12:661–6.
 76. Roussev RG, Ng SC, Coulam CB. Natural killer cell functional activity suppression by intravenous immunoglobulin, intralipid and soluble human leukocyte antigen-G. *Am J Reprod Immunol* 2007;57:262–9.
 77. Foyle KL, Sharkey DJ, Moldenhauer LM, Green ES, Wilson JJ, Roccisano CJ, et al. Effect of Intralipid infusion on peripheral blood T cells and plasma cytokines in women undergoing assisted reproduction treatment. *Clin Transl Immunology* 2021;10:e1328.
 78. Singh N, Davis AA, Kumar S, Kriplani A. The effect of administration of intravenous intralipid on pregnancy outcomes in women with implantation failure after IVF/ICSI with non-donor oocytes: a randomised controlled trial. *Eur J Obstet Gynecol Reprod Biol* 2019;240:45–51.
 79. Al-Zebeidi J, Agdi M, Lary S, Al-Obaid S, Salim G, Al-Jaroudi D. Effect of empiric intravenous intralipid therapy on pregnancy outcome in women with unexplained recurrent implantation failure undergoing intracytoplasmic sperm injection-embryo transfer cycle: a randomized controlled trial. *Gynecol Endocrinol* 2020;36:131–4.
 80. Hayes BD, Gosselin S, Calelio DP, Nacca N, Rollins CJ, Abourbih D, et al. Systematic review of clinical adverse events reported after acute intravenous lipid emulsion administration. *Clin Toxicol (Phila)* 2016;54:365–404.
 81. Perez EE, Orange JS, Bonilla F, Chinen J, Chinn IK, Dorsey M, et al. Update on the use of immunoglobulin in human disease: a review of evidence. *J Allergy Clin Immunol* 2017;139:S1–46.
 82. Moraru M, Carbone J, Alecsandru D, Castillo-Rama M, Garcia-Segovia A, Gil J, et al. Intravenous immunoglobulin treatment increased live birth rate in a Spanish cohort of women with recurrent reproductive failure and expanded CD56(+) cells. *Am J Reprod Immunol* 2012;68:75–84.
 83. Winger EE, Reed JL, Ashoush S, Ahuja S, El-Toukhy T, Taranissi M. Treatment with adalimumab (Humira) and intravenous immunoglobulin improves pregnancy rates in women undergoing IVF. *Am J Reprod Immunol* 2009;61:113–20.
 84. Ahmadi M, Abdolmohammadi-Vahid S, Ghaebi M, Aghebati-Maleki L, Dolati S, Farzadi L, et al. Regulatory T cells improve pregnancy rate in RIF patients after additional IVIG treatment. *Syst Biol Reprod Med* 2017;63:350–9.
 85. Chernyshov VP, Dons'koi BV, Sudoma IO, Goncharova YO. Multiple immune deviations predictive for IVF failure as possible markers for IVIG therapy. *Immunol Lett* 2016;176:44–50.
 86. Ho YK, Chen HH, Huang CC, Lee CI, Lin PY, Lee MS, et al. Peripheral CD56⁺ CD16⁺ NK cell populations in the early follicular phase are associated with successful clinical outcomes of intravenous immunoglobulin treatment in women with repeated implantation failure. *Front Endocrinol (Lau-sanne)* 2019;10:937.
 87. Sher G, Maassarani G, Zouves C, Feinman M, Sohn S, Matzner W, et al. The use of combined heparin/aspirin and immunoglobulin G therapy in the treatment of in vitro fertilization patients with antithyroid antibodies. *Am J Reprod Immunol* 1998;39:223–5.
 88. Stephenson MD, Fluker MR. Treatment of repeated unexplained in vitro fertilization failure with intravenous immunoglobulin: a randomized, placebo-controlled Canadian trial. *Fertil Steril* 2000;74:1108–13.
 89. Stewart CL, Kaspar P, Brunet LJ, Bhatt H, Gadi I, Kontgen F, et al. Blastocyst implantation depends on maternal expression of leukaemia inhibitory factor. *Nature* 1992;359:76–9.
 90. Suman P, Malhotra SS, Gupta SK. LIF-STAT signaling and trophoblast biology. *JAKSTAT* 2013;2:e25155.
 91. Brinsden PR, Alam V, de Moustier B, Engrand P. Recombinant human leukemia inhibitory factor does not improve implantation and pregnancy outcomes after assisted reproductive techniques in women with recurrent unexplained implantation failure. *Fertil Steril* 2009;91:1445–7.
 92. Turocy J, Williams Z. Novel therapeutic options for treatment of recurrent implantation failure. *Fertil Steril* 2021;116:1449–54.
 93. Okitsu O, Kiyokawa M, Oda T, Miyake K, Sato Y, Fujiwara H. Intrauterine administration of autologous peripheral blood mononuclear cells increases clinical pregnancy rates in frozen/thawed embryo transfer cycles of patients with repeated implantation failure. *J Reprod Immunol* 2011;92:82–7.
 94. Yoshioka S, Fujiwara H, Nakayama T, Kosaka K, Mori T, Fujii S. Intrauterine administration of autologous peripheral blood mononuclear cells promotes implantation rates in patients with repeated failure of IVF-embryo transfer. *Hum Reprod* 2006;21:3290–4.
 95. Li S, Wang J, Cheng Y, Zhou D, Yin T, Xu W, et al. Intrauterine administration of hCG-activated autologous human peripheral blood mononuclear cells (PBMC) promotes live birth rates in frozen/thawed embryo transfer cycles of patients with repeated implantation failure. *J Reprod Immunol* 2017;119:15–22.
 96. Pourmoghadam Z, Soltani-Zangbar MS, Sheikhsari G, Azizi R, Eghbal-Fard S, Mohammadi H, et al. Intrauterine administration of autologous hCG- activated peripheral blood mononuclear cells improves pregnancy outcomes in patients with recurrent implantation failure; a double-blind, randomized control trial study. *J Reprod Immunol* 2020;142:103182.
 97. Yu N, Zhang B, Xu M, Wang S, Liu R, Wu J, et al. Intrauterine administration of autologous peripheral blood mononuclear cells (PBMCs) activated by HCG improves the implantation and pregnancy rates in patients with repeated implantation failure: a prospective randomized study. *Am J Reprod Immunol* 2016;76:212–6.
 98. Onalan G, Tohma YA, Zeyneloglu HB. Effect of etanercept on the success of assisted reproductive technology in patients with endometrioma. *Gynecol Obstet Inves* 2018;83:358–64.
 99. Singh JA, Cameron C, Noorbaloochi S, Cullis T, Tucker M, Christensen R, et al. Risk of serious infection in biological treatment of patients with rheumatoid arthritis: a systematic review and meta-analysis. *Lancet* 2015;386:258–65.
 100. Han AR, Han JW, Lee SK. Inherited thrombophilia and anticoagulant therapy for women with reproductive failure. *Am J Reprod Immunol* 2021;85:e13378.
 101. Practice Committee of the American Society for Reproductive Medicine. The role of immunotherapy in in vitro fertilization: a guideline. *Fertil Steril* 2018;110:387–400.
 102. Nardo LG, El-Toukhy T, Stewart J, Balen AH, Potdar N. British Fertility Society Policy and Practice Committee: adjuvants in IVF: evidence for good clinical practice. *Hum Fertil (Camb)* 2015;18:2–15.
 103. Shaoul T, Sierra S, Sylvestre C. Recurrent implantation failure in IVF: a Canadian Fertility and Andrology Society clinical practice guideline. *Reprod Biomed Online* 2020;41:819–33.
 104. Liu KE, Hartman M, Hartman A. Management of thin endometrium in assisted reproduction: a clinical practice guideline from the Canadian Fertility and Andrology Society. *Reprod Biomed Online* 2019;39:49–62.
 105. Katz P, Showstack J, Smith JF, Nachtigall RD, Millstein SG, Wing H, et al. Costs of infertility treatment: results from an 18-month prospective cohort study. *Fertil Steril* 2011;95:915–21.
 106. Drugs.com. Neupogen prices. Available at: <https://www.drugs.com/price-guide/neupogen>. Accessed April 6, 2022.
 107. Farimani M, Mehrabi N, Pirdehghan A, Bahmanzadeh M. The effects of subcutaneous administration of granulocyte colony-stimulating factor on pregnancy outcome after assisted reproductive technology: clinical trial. *Avicenna J Med Biochem* 2018;6:31–6.
 108. MacIsaac J, Siddiqui R, Jamula E, Li N, Baker S, Webert KE, et al. Systematic review of rituximab for autoimmune diseases: a potential alternative to intravenous immune globulin. *Transfusion* 2018;58:2729–35.
 109. Saab W, Seshadri S, Huang C, Alsukbi L, Sung N, Kwak-Kim J. A systemic review of intravenous immunoglobulin G treatment in women with recurrent implantation failures and recurrent pregnancy losses. *Am J Reprod Immunol* 2021;85:e13395.

110. Ata B, Kalafat E, Somigliana E. A new definition of recurrent implantation failure on the basis of anticipated blastocyst aneuploidy rates across female age. *Fertil Steril* 2021;116:1320–7.
111. Garcia-Alonso L, Handfield LF, Roberts K, Nikolakopoulou K, Fernando RC, Gardner L, et al. Mapping the temporal and spatial dynamics of the human endometrium *in vivo* and *in vitro*. *Nat Genet* 2021;53:1698–711.
112. Huhn O, Ivarsson MA, Gardner L, Hollinshead M, Stinchcombe JC, Chen P, et al. Distinctive phenotypes and functions of innate lymphoid cells in human decidua during early pregnancy. *Nat Commun* 2020;11:381.
113. Piekarzka K, Radwan P, Tarnowska A, Wiśniewski A, Radwan M, Wilczyński JR, et al. ERAP, KIR, and HLA-C profile in recurrent implantation failure. *Front Immunol* 2021;12:755624.