

The Pathobiology of Pulmonary Arterial Hypertension



Sudarshan Rajagopal, MD, PhD^{a,*}, Yen-Rei A. Yu, MD, PhD^b

KEYWORDS

• Pulmonary arterial hypertension • Endothelial cell • Smooth muscle cell • Fibroblast • Inflammation

KEY POINTS

- Pulmonary arterial hypertension is a disease of the pulmonary arterioles, characterized by abnormal remodeling and obstruction.
- Pulmonary arterial hypertension leads to increased resistance in the pulmonary vessels and right ventricular afterload, eventually resulting in right heart failure.
- Over the past decades, a number of important pathobiological mechanisms have been found to contribute to pulmonary arterial hypertension.
- These changes are in endothelial function, smooth muscle cell proliferation, fibroblast activation, inflammation, and metabolism.
- A number of molecular mechanisms underlie these pathophysiological changes, including alterations in type II bone morphogenetic protein receptor and transforming growth factor- β signaling, changes in vasoactive mediators, chemokine and cytokine signaling, ion channel activity, transcription factors, and microRNAs and other epigenetic changes.

THE PATHOLOGY OF PULMONARY ARTERIAL HYPERTENSION

Human Pathology

Pulmonary arterial hypertension (PAH) is characterized by lesions in the distal arterioles, ranging from 50 to 500 μm in size.^{1,2} Grossly, this results in a decrease in distal perfusion and the classic “pruning” of the distal pulmonary vasculature observed on a chest radiograph or pulmonary angiography.³ This in turn results in increased right ventricular (RV) afterload and right heart failure. The histopathologic findings of PAH include medial hypertrophy or hyperplasia, intimal and adventitial fibrosis, and plexiform lesions, the pathognomonic lesions for PAH.^{1,2} Recent detailed histologic analyses have demonstrated that plexiform lesions are complex lesions that arise from arteriolar obstruction followed by

collateralization of the vasa vasorum and bronchial arteries.^{4,5} Thus, these lesions represent a compensatory mechanism to maintain distal perfusion through the pulmonary arterial system. In addition to pulmonary arterioles, remodeling of the pulmonary capillaries and venules is also observed in all PH groups.⁶ Beyond the pulmonary vasculature, systemic vascular abnormalities in PAH support the emerging paradigm of PAH as a systemic vasculopathy.⁷ With their constant exposure to the entire cardiac output, the pulmonary arterioles are most susceptible to vascular remodeling.

Animal Models

Because clinical studies and patient sample studies are limited in offering insights into the pathogenesis of PAH, complementary animal models

^a Division of Cardiology, Department of Medicine, Duke University Medical Center, Room 128A Hanes House, 330 Trent Drive, Durham, NC 27710, USA; ^b Division of Pulmonary Sciences and Critical Care Medicine, University of Colorado, 12605 E. 16th Avenue, Aurora, CO 80045, USA

* Corresponding author.

E-mail address: sudarshan.rajagopal@duke.edu

are used to better understand the cellular and molecular basis of disease and for testing potential therapies. The earliest and simplest model is chronic, hypoxia-induced PAH. This state was first observed in cattle living at high altitude and was coined “brisket disease.”⁸ High altitude decreases oxygen tension, causes reactive hypoxic vasoconstriction, and results in abnormal medial hypertrophy of pulmonary arterioles. Although chronic hypoxia-induced PAH was observed initially in large animals, the observations has been extended to small animals (ie, mice and rats). Today, small animals are the most commonly used models for examining PAH.^{9,10} Because pathology induced by chronic hypoxia does not recapitulate all the features of human disease, the focus of many studies has been on 2-hit approaches to induce severe pulmonary vascular remodeling. For example, mice are relatively resistant to hypoxic stress and generally develop mild pulmonary remodeling and PH when exposed to chronic hypoxia. However, transgenic technology in mice allows the flexibility to examine the contribution of specific molecules, pathways, and cell types to disease pathogenesis. Genetic modifications in combination with hypoxia can lead to severe PH in mice, for example, exposing mice with lung-specific overexpression of IL-6 to hypoxia.¹¹ Other 2-hit approaches, including combining SU5416 injections, a vascular endothelial growth factor receptor 2 antagonist that can cause endothelial injury, with hypoxia, pneumonectomy, induction of allergic inflammation, or other genetic modifications,⁹ can also enhance the severity of PH.

Similar strategies have also been applied to rat models that, compared with mice, generally develop significantly more extensive vascular remodeling with severe pulmonary hypertension. Although mice are resistant to monocrotaline (MCT) toxicity, MCT administration is commonly used to induce PH in rats, where a single subcutaneous injection of MCT results in progressive, severe pulmonary remodeling, right heart failure, and fatal PH.¹⁰ MCT is thought to act as a toxin to the endothelium, although its effects are complex and not organ-specific. The SU5416-hypoxia model results in severe and sustained PH,¹² which can be fatal depending on the strain of rat used.¹³ Together, the use of these models has been invaluable in probing the pathobiology and pathophysiology of PAH.

THE PATHOPHYSIOLOGY OF PULMONARY ARTERIAL HYPERTENSION

The pathophysiology of PAH is complex, with contributions from multiple cell types in the pulmonary

vasculature and the right heart. Here we highlight some of the major mechanisms that contribute to pulmonary vascular remodeling (Fig. 1) and RV dysfunction in PAH.

Endothelial Dysfunction

PAH is characterized by microvascular rarefaction. Pulmonary artery endothelial cell (PAEC) injury leading to EC dysfunction and apoptosis are triggers for the development of PAH. Both animal models of PH (ie, MCT and SU5416-hypoxia models) and human type II bone morphogenetic protein receptor (BMPR2) mutation-associated PAH are characterized by lung PAEC apoptosis.¹⁴ Chronic PAEC apoptosis results in microvascular destruction and rarefaction. In addition, apoptosis leads to the selection of apoptosis resistance and proliferative PAECs causing occlusive arterial remodeling.¹⁵ Cultured PAECs isolated from human patients with PAH demonstrate increased proliferation compared with nondiseased controls.¹⁶ Idiopathic PAH (IPAH) PAECs also have metabolic abnormalities, with decreased mitochondrial numbers per cell and a higher glycolytic rate.¹⁷ It is unclear as to which of these processes—a primary loss of vessels owing to EC apoptosis versus a secondary loss of vessels owing to obliteration from apoptosis-resistant ECs—is central to PAH pathogenesis.¹⁸ Additionally, these processes may play roles in different phases of the disease.¹⁹ Another aspect by which the EC contributes to PAH is endothelial-to-mesenchymal transition, where ECs can transition to smooth muscle cell (SMC)-like cells that upregulate twist and vimentin.²⁰

Smooth Muscle Cell Hypertrophy and Proliferation

Hypertrophy, proliferation, migration, and apoptosis resistance of medial SMCs in the pulmonary arterioles (PASMCs) plays a central role in PAH. PASMCs in normal adult lung are quiescent with a contractile and nonmigratory phenotype. In the setting of tissue injury, SMCs can transition to a proliferative phenotype, resulting in vessel remodeling. Pathologic distal arteriole muscularization results from proliferation of preexisting SMCs and recapitulates many aspects of arterial wall development, including SMC dedifferentiation, distal migration, proliferation, and then redifferentiation.²¹ Some of the cues for this process are regulated by nonclassical monocytes and macrophages that sense hypoxia and are predicted to promote pulmonary vascular remodeling and SMC proliferation.²² For example, macrophage-derived factors such as platelet-derived growth

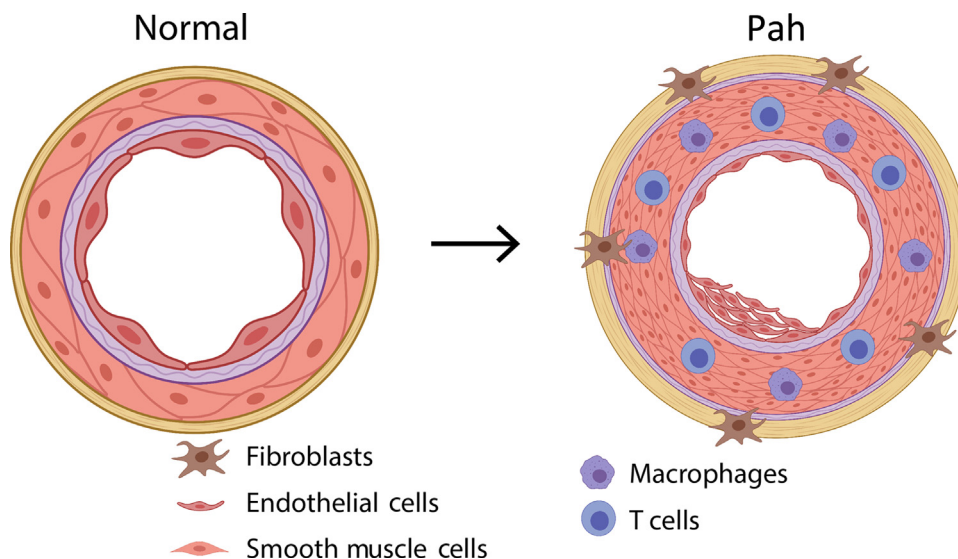


Fig. 1. Cell types that contribute to pulmonary vascular remodeling. Pulmonary vascular remodeling involves all layers of the vascular wall (the intima, primarily composed of ECs; the media, primarily composed of SMCs; and the adventitia, primarily composed of fibroblasts) along with infiltration with immune cells such as macrophages and T cells. This results in the selection of apoptosis-resistant ECs, the proliferation of SMCs, and the activation of fibroblasts. Much of this is driven by signaling from immune cells such as macrophages and T cells. See text for details. Created with [BioRender.com](https://www.biorender.com).

factor- β are upregulated in PAH and promote SMC proliferation.²³ There are also cell autonomous factors that contribute to SMC proliferation. This process is exemplified by SMC-specific knockout of *BMPR1A*, which resulted in attenuated SMC proliferation, decreased hypoxia-mediated muscularization of distal vessels, and the preservation of peripheral pulmonary arteries.²⁴

Fibroblasts and Fibrosis

Obliterative thickening of vessels in PAH are largely composed of SMCs and myofibroblasts. Frank fibrosis has been noted in PAH pathologic samples, especially in those with PAH associated with connective tissue disease.² Adventitial fibroblasts from calves with severe hypoxia-induced PH and humans with IPAH display increased aerobic glycolysis. Treatment of these mice with a pharmacologic agent for a nicotinamide adenine dinucleotide-sensitive transcription corepressor resulted in decreased glycolysis and expression of inflammatory genes, attenuated proliferation, and decreased remodeling of the distal pulmonary vasculature.²⁵ These fibroblasts also activate macrophages through paracrine signaling via IL-6-activated STAT3 to promote pulmonary vascular remodeling.²⁶

Immune Cells and Inflammation

Inflammation is known to be an important mediator of vascular dysfunction in the pathogenesis of PAH. Accumulation of immune cells, including T cells, B

cells, monocytes, macrophages, and mast cells, has been observed in all animal models of PH and human disease.^{27,28} The predominant cell types are T cells, monocytes, and macrophages. These cells are found mostly in the perivascular regions, but also observed within the plexiform lesions.²⁹ T cells, monocytes, and macrophages can directly or indirectly, via paracrine signaling, modulate the activation, proliferation, transformation, migration, and survival of vascular mural cells (ie, EC, vascular SMCs, and adventitial fibroblasts), modify extracellular matrix, and regulate remodeling of all parts of the vascular wall. Altering immune cell populations and balance in both adaptive and innate immune compartments modulates vascular remodeling and PH severity. For example, athymic animals develop severe pulmonary remodeling and PH spontaneously owing to the absence of the protective effect of CD4⁺ T regulatory cells.^{30,31} In contrast, Th1 and Th17 cells secrete proinflammatory cytokines such as IL-6, IL-1, IL-21, and tumor necrosis factor- α , and IFN- γ , and promote vascular remodeling.^{32,33} Additionally, Th17 cells also promote PH through the secretion of IL-17A.³⁴ Although Th1 cells and IFN- γ are required for the development of pneumocystis-induced PAH³²; Th2 cells, through production of IL-4 and IL-13, exacerbate schistosomiasis-induced and other forms of PAH.^{35–37} Thus, the balance of Th1 versus Th17 or Th2 versus T regulatory cell populations determine the immune milieu and its effects on vascular remodeling.³³

Monocytes and macrophages are also involved in multiple aspects of PAH pathogenesis. Recent studies have shown that macrophages are derived from both fetal precursors or circulating monocytes; the function of macrophages depends on their origin, anatomic microenvironment, and specific stimuli.^{38–41} Consistent with this paradigm, depletion strategies that differentially affect various populations of monocytes and macrophage can either promote or ameliorate PAH severity.^{42–44} Preferential polarization of macrophages to alternative phenotype (M2-like) spectrum, characterized by signaling through STAT3, and promote cellular proliferation and is believed to drive PAH. However, in animal models of PAH, both M1-like and M2-like macrophages have been described.⁴⁵ M1-like macrophages are potent producers of interferon, which is a known driver of vascular remodeling and PAH. Moreover, the M1/M2 classification is an oversimplification of macrophage involvement in PAH.⁴⁶ Macrophage recruitment and activation are modulated by many factors produced by both vascular mural and other immune cells, including a decrease in BMP2 signaling, secreted macrophage inhibitory factor by dysfunctional ECs, leukotriene T4, and chemokines (eg, CCL2, CCL1, and CX3CL1).^{33,47–50} These molecules and pathways are being explored as potential therapeutic targets for PAH. In addition to T cells and macrophages, mast cells and eosinophils have been implicated in PAH pathogenesis.^{51–55} With the complexity of different immune cell populations and responses, immune cell dysregulation generally promotes the abnormal vascular mural cell phenotype in PAH.

Metabolism

As noted elsewhere in this article, many of the cell types in pulmonary vasculature display changes in their metabolic state. Paulin and Michelakis⁵⁶ have proposed that many of the pathobiological abnormalities in PAH promote mitochondrial suppression, with an inhibition of glucose oxidation, which in turn results in many of the observed molecular abnormalities noted in PAH. It has been proposed that these metabolic abnormalities are similar to those seen in cancer and explain features of the PAH vascular phenotype, including the enhancement of proliferation and apoptosis resistance. Notably, insulin resistance is also common in PAH and is characterized by alterations in lipid and lipoprotein levels, and elevated levels of circulating medium- and long-chain acylcarnitines.^{57,58} These defects in fatty acid oxidation contribute to lipotoxicity in the RV in PAH.⁵⁹

Metabolic profiling has also been used to identify signatures of RV–pulmonary valve dysfunction, consistent with the pulmonary release of tryptophan metabolites.⁶⁰ Although it is likely an oversimplification to reduce all of the complex abnormalities in PAH to a single underlying cause of metabolic changes, these studies clearly demonstrate that metabolic abnormalities contribute to PAH pathogenesis and phenotype.

Right Ventricular Dysfunction

The majority of basic pathobiology studies in PAH focus on abnormalities in the pulmonary vasculature, but the RV in PAH also has an abnormal response to increased afterload.⁶¹ The increased afterload from pulmonary vascular remodeling results in right heart failure, characterized by decreased RV function that leads to insufficient cardiac output and/or increased filling pressure at rest or exercise. In response to the increased afterload, the RV displays an adaptive hypertrophic response, but over time this can transition to a maladaptive phenotype.⁶² Notably, patients who display preserved RV function in PH have significantly better survival than those who have decreased RV function at follow-up. Adaptive remodeling is characterized by the preservation of a normal cardiac output, RV ejection fraction, filling pressures, and exercise capacity. Adaptive remodeling consists primarily of concentric hypertrophy with minimal dilatation and fibrosis.⁶² Maladaptive remodeling is associated with increased filling pressures and a decreased cardiac output and RV ejection fraction; it consists primarily of RV dilatation and fibrosis. With these compensatory mechanisms, RV–pulmonary arterial coupling (typically quantified by the ratio of RV end-systolic elastance and arterial elastance) with efficient energy transfer is maintained initially in adaptive remodeling. However, it is then overwhelmed gradually⁶¹ and transitions from adaptive to maladaptive remodeling. In MCT-induced PH rat models, this transition is associated with a decrease in angiogenesis, a decrease in glucose uptake, and a reversion toward normal metabolism.⁶³ However, a detailed analysis of human samples with stereology⁶⁴ demonstrated that, in advanced PH, there was a significant increase in the RV vasculature in the setting of RV hypertrophy, consistent with compensatory angiogenesis in severe PAH. Similar results were observed in Su5416-hypoxia PH models,⁶⁵ where a compensatory angiogenic response was observed, but with a modest decrease in arterial delivery of metabolic substrates owing to an increase in the radius of tissue served per vessel. Metabolomics

revealed major metabolic alterations and reprogramming, but without evidence of tissue hypoxia or depletion of key metabolic substrates. This finding suggested that the major driver of RV maladaptation was related to direct changes in cardiomyocytes and not secondary to vascular rarefaction and decreased substrate delivery. Despite these findings supporting the importance of RV function on outcomes, there are currently no PAH therapies that directly target the RV.

MOLECULAR MECHANISMS OF PULMONARY ARTERIAL HYPERTENSION

Multiple molecular mechanisms contribute to the pathophysiological axes discussed elsewhere in this article. We highlight a number of these mechanisms in [Fig. 2](#) and elsewhere in this discussion.

Type II Bone Morphogenetic Protein Receptor and Transforming Growth Factor- β Signaling

BMPR2 mutations account for 70% of heritable PAH and are also found in 20% of patients with IPAH.⁶⁶ BMPR2 is a member of the transforming growth factor (TGF)- β receptor superfamily. While

TGF- β receptors promote signaling through Smad2/3 transcription factors, BMPR2 promotes signaling through Smad1/5 transcription factors.⁶⁷ A large meta-analysis also identified that patients with BMPR2 mutations, whether idiopathic, heritable, or anorexigen-associated, presented at a younger age with more severe disease and were at a higher risk of death compared with those without BMPR2 mutations.⁶⁸ Mutations of a number of proteins important in the TGF- β superfamily have been identified as potentially pathogenic in next-generation sequencing studies in PAH.⁶⁹ Although some of these genetic links have been known for some time,⁷⁰ the complex pharmacology of this system has made it difficult to identify the specific receptor heteroimers and their ligands that were responsible for signaling in the pulmonary vascular endothelium.⁷¹ A breakthrough was made with the discovery that BMP9 is the preferred ligand for preventing apoptosis and enhancing endothelial integrity.⁷² Additionally, the administration of BMP9 reversed established PAH in mice carrying a heterozygous knock-in allele of human BMPR2 (R899X) mutation.⁷² More recently, an approach of targeting the

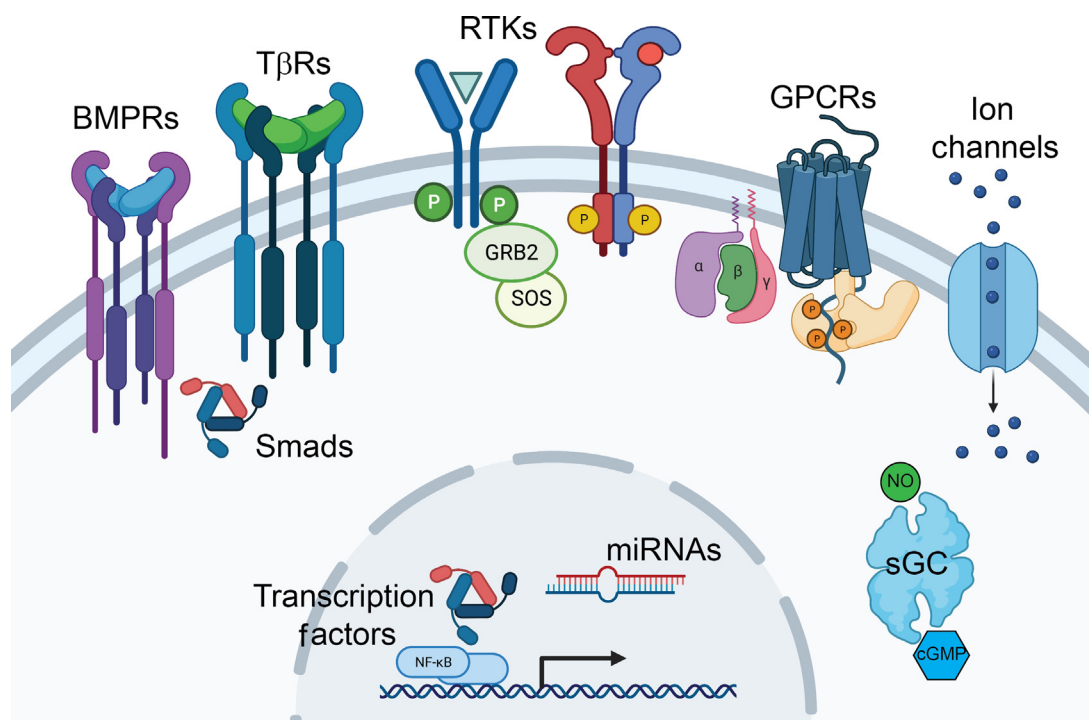


Fig. 2. Molecular mechanisms that contribute to PAH. Multiple molecular mechanisms contribute to the development of PAH, including signaling by BMPRs and TGF- β receptors through Smads, growth factor signaling via receptor tyrosine kinases (RTKs), growth factors and vasoactive mediator signaling via G protein-coupled receptors, the activation of ion channels, nitric oxide signaling via soluble guanylate cyclase (sGC), the activation of transcription factors and epigenetic mechanisms such as microRNAs (miRNAs). See text for details. Created with [BioRender.com](#).

balance of activin/growth differentiation factor and BMP signaling via BMPR2 has demonstrated pre-clinical efficacy.⁷³ Although activins and growth differentiation factors promote signaling via Smad2/3, BMPs promote signaling via Smad1/5/9. Treatment with a potent growth differentiation factor 8/11 and activin ligand trap inhibited Smad2/3 signaling, resulting in decreased proliferation and enhancement of apoptosis in the vascular wall, and attenuated PH.⁷³ These findings suggest that new approaches to target BMPR2 signaling will be useful as a therapeutic strategy in PAH.

Growth Factors

The dysregulation of a range of growth factors, many of which target receptor tyrosine kinases (RTKs), promote abnormal vascular proliferation and play a central role in PAH pathobiology.⁷⁴ Markers of angiogenesis, such as vascular endothelial growth factor, have been noted in plexiform lesions.⁷⁵ In the setting of vascular destruction and obstruction in PAH, this likely reflects an angiogenic response to form collaterals from bronchial/vasa vasorum to the pulmonary arterial circulation. KDR heterozygosity (the gene encoding vascular endothelial growth factor receptor 2) is strongly associated with PAH that occurs later in life.⁷⁶ Levels of fibroblast growth factor 2 (FGF2) are elevated in the remodeled pulmonary vascular endothelium of patients with IPAH; and FGF2 knockdown decreased PASMC growth.⁷⁷ In an animal model, the inhibition of FGF receptor 1, a receptor for FGF2, reverses established PH, suggesting this receptor as a potential therapeutic target. Similarly, platelet-derived growth factor (PDGF) promotes PASMC proliferation and migration; and expression of PDGF receptor (PDGFR) is increased in the lung tissue of patients with PAH.⁷⁸ The PDGFR antagonist, imatinib, can reverse advanced pulmonary vascular disease in MCT-PH rats and hypoxia mice.⁷⁸ Consistent with this finding, imatinib significantly improved exercise capacity and hemodynamics in a clinical trial. Unfortunately, it was also associated with a significant increase in subdural hematomas in patients who were on anticoagulation.⁷⁹ In PAH, epidermal growth factor (EGF) activates its receptor (EGFR) to promote cell proliferation and survival. However, the effects of different EGFR antagonists in MCT PH have been noted to be variable, with only some improving RV systolic pressure and hypertrophy.⁸⁰ Notably, none of them significantly improved RV systolic pressure or pulmonary vascular remodeling in mice with chronic hypoxic PH. Consistent with this finding, EGFR expression

in lungs was not altered in patients with IPAH. Targeting a range of tyrosine and serine/threonine kinases with sorafenib decreased RV hypertrophy and pulmonary arterial muscularization in MCT-treated rats.⁸¹ An approach using multiple agents to target RTKs (ie, FGFR, EGFR, and PDGFR), was successful in regressing established MCT PH. Treatment decreased the activation of the adapter protein p130(Cas), which works downstream of RTKs to promote cell migration and proliferation.⁸² With their central roles in regulating cell migration, proliferation and survival, these growth factors are excellent drug targets in PAH if their potential systemic effects can be limited.

Vasoactive Mediators: Prostacyclins, Endothelins, and Nitric Oxide

Vasoactive mediators have long been known to contribute to PAH pathophysiology, with low levels of vasodilators such as prostacyclin⁸³ and high levels of vasoconstrictors such as endothelin-1⁸⁴ noted in patients with PAH. Only a small percentage of patients with PAH are vasodilator responsive,⁸⁵ displaying an acute decrease in pulmonary pressures after treatment with a pulmonary vasodilator such as inhaled nitric oxide, adenosine or epoprostenol.⁸⁶ Vasodilator responders display an excellent long-term therapeutic response to calcium channel blockers.⁸⁷ Regardless of their acute response to vasodilators, patients with PAH demonstrated a positive response to treatments, such as prostacyclin infusion, in early studies.⁸⁸ These initial discoveries translated to the first therapy for PAH, intravenous epoprostenol.^{89,90} Although long-term treatment with therapies such as prostacyclins have been shown to be beneficial, they do not seem to reverse PAH pathology fully. As noted in an autopsy study of a PAH patient maintained on long-term therapy who died of a different cause, vascular abnormalities are still evident.⁹¹ All 3 pathways currently targeted by PAH-specific therapies—the prostacyclin pathway (targeted by prostacyclin receptor agonists), the nitric oxide/cyclic guanosine monophosphate axis (targeted by phosphodiesterase 5 inhibitors and soluble guanylate cyclase stimulators), and the endothelin pathway (targeted by endothelin receptor antagonists)—have direct vasoactive effects. It is possible that other vasoactive pathways, such as angiotensin^{92,93} and other mediators,⁹⁴ would also have beneficial effects in PAH, but they have not been tested in large clinical trials in the PAH population. Notably, although these drugs are thought to primarily act through vasodilation, with prolonged exposure, they likely also act as

antiproliferative agents, because vasoconstriction is closely linked to other complex changes in the vasculature.⁹⁵

Chemokines

Chemokines, also known as chemotactic cytokines, are a group of more than 40 small proteins that regulate cell migration and function. Chemokines bind to more than 20 chemokine receptors, which are G protein-coupled receptors. Chemokine receptors are categorized into 5 families based on their activating chemokine ligand (CXCR, CCR, CX3CR, or XCRs), with an additional group of atypical chemokine receptors that bind chemokines of different families, but primarily act as scavenger or decoy receptors. Chemokines bind to negatively charged glycosaminoglycans on the EC surface or extracellular matrix, resulting in a chemokine concentration gradient that promotes immune and inflammatory cell recruitment.^{96,97} Chemokine levels have been correlated with disease severity, pulmonary hemodynamics, and RV function in multiple studies. For example, CCL2 and CXCL10 levels are associated with disease severity.^{98–101} CCL2, CXCL8, CXCL10, CXCL12, and CXCL13 levels are correlated with hemodynamics such as pulmonary vascular resistance,^{100,102–104} right atrial pressure,^{98,102} cardiac output,¹⁰² and cardiac index.^{98,100,102} With respect to harder outcomes, CCL2, CCL21, and CXCL12 levels have been associated with adverse outcomes and mortality.^{105–108} Some of these chemokines may increase as part of an adaptive mechanism; for example, elevated levels of CXCL10 are associated with improved survival in patients with PAH.¹⁰⁹ In addition, some chemokines could serve as biomarkers for monitoring disease progression or treatment effect. For example, the beneficial effects of epoprostenol treatment on functional and hemodynamic status in patients with PAH were associated with increased levels of CCL2.¹⁰⁴ Despite these strong associations with disease severity, chemokines have been challenging to target owing to their complex pharmacology, but novel approaches may make these targets more druggable.¹¹⁰

Ion Channels

Ion channels play a central role in the membrane potential and regulate vascular tone of the pulmonary circulation. Voltage-gated potassium channels play a central role in hypoxic vasoconstriction.¹¹¹ Mice with mutation of Kv1.5 display impaired hypoxic vasoconstriction.¹¹² Fawn-hooded rats have a chromosomal

abnormality that disrupts a mitochondria–reactive oxygen species–hypoxia inducible factor–Kv pathway, resulting in a loss of voltage-gated potassium channel activity. These rats develop pulmonary hypertension spontaneously.¹¹³ A familial form of PAH is due to heterozygous loss-of-function mutations in KCNK3,¹¹⁴ which encodes the TASK-1 potassium channel. Consistent with this finding, MCT-treated rats have decreased expression of KCNK3, and the pharmacologic activation of KCNK3 improved PH.¹¹⁵ Such findings suggest that ion channels may serve as drug targets in PAH treatment.

Transcription Factors and Nuclear Receptors

Transcription factors are the downstream targets of a wide range of signals, such as hypoxia, growth factors, cytokines, and chemokines, that result in the reprogramming of cells to a PAH-promoting phenotype. Hypoxia inducible factor-1 α mediates much of the transcriptional response to hypoxia, and its activity in macrophages results in the activation of pro-proliferative signals to PASMCs. For example, FoxO1 is responsible for signaling downstream of many growth factors and inflammatory mediators, and inactivation of specific isoforms results in the pro-proliferative and antiapoptotic phenotype of PASMCs in PAH.¹¹⁶ Similarly, the related transcription factor FoxM1 promotes PASMC proliferation in PAH.¹¹⁷ PPAR- γ and β -catenin have been shown to promote BMPR2 signaling via apelin, resulting in improved PAEC survival. In mice, this effect was recapitulated with apelin treatment leading to PAH reversal.¹¹⁸

MicroRNAs and Long Noncoding RNAs

A number of epigenetic changes have been noted in PAH. Noncoding RNAs, such as microRNAs (miRNAs) and long noncoding RNAs (lncRNAs), have been identified as being dysregulated in PAH. The miRNAs are small noncoding RNAs that inhibit gene expression and are critical in gene regulation.¹¹⁹ Recently, a number of groups have identified changes in miRNA expression in PAH that seem to contribute to PAH pathobiology. Of these miRNAs, miR-29, -124, -140, and -204 have been reported as dysregulated in PAH, where they seem to play distinct roles.¹²⁰ miR-124, -140, and -240 inhibit cellular proliferation, and miR-29 promotes proliferation but inhibits vasoconstriction. However, it has been challenging to interpret some of these studies, because the effects are not always consistent between patients and different disease models.¹²⁰ More recently, lncRNAs have been implicated in PAH pathogenesis. The lncRNA H19 was described recently as a new biomarker

for right heart failure in PAH. The lncRNA H19 is upregulated in the decompensated RV of patients with PAH and also in rat disease models.¹²¹ Silencing H19 improved RV fibrosis and capillary rarefaction, suggesting that targeting these lncRNAs could be a potential therapeutic strategy. The lncRNA TYKRIL has been shown to be upregulated in pericytes and PASMCs in cells exposed to hypoxia and patients with IPAH.¹²² Expression of PDGFR β strongly correlated with TYKRIL expression and TYKRIL knockdown decreased PDGFR β expression, suggesting another approach for targeting pathogenic signaling in PAH.

SUMMARY

The pathobiology of PAH is complex, with contributions from multiple pathophysiologic processes that are regulated by a variety of molecular mechanisms. This finding likely explains the limited efficacy of our current therapies, which only target a small portion of the pathobiological mechanisms that underlie advanced disease. It is likely that different therapies that target these multiple axes will need to be used in combination, as is being used with our currently available therapies,⁸⁶ to improve outcomes in this devastating disease.

CLINICS CARE POINTS

- Multiple cellular and molecular mechanisms contribute to vascular remodeling in the development of PAH.
- Current PAH therapies focus on targeting vasoactive mechanisms, including prostacyclin receptor agonists, endothelin receptor antagonists, phosphodiesterase 5 inhibitors, and soluble guanylate cyclase stimulators.
- Novel PAH therapies that are being developed target other signaling pathways including the type II bone morphogenetic protein receptor and growth factor receptors.
- All of these pathways regulate endothelial function, smooth muscle cell proliferation, fibroblast activation, inflammation, and metabolism that underlie PAH pathogenesis.

DISCLOSURE

The authors have no commercial or financial conflicts of interest.

REFERENCES

1. Heath D, Edwards JE. The pathology of hypertensive pulmonary vascular disease; a description of six grades of structural changes in the pulmonary arteries with special reference to congenital cardiac septal defects. *Circulation* 1958;18:533–47.
2. Stacher E, Graham BB, Hunt JM, et al. Modern age pathology of pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2012;186:261–72.
3. Gray HH, Morgan JM, Kerr IH, et al. Clinical correlates of angiographically diagnosed idiopathic pulmonary hypertension. *Thorax* 1990;45:442–6.
4. Galambos C, Sims-Lucas S, Abman SH, et al. Intrapulmonary bronchopulmonary anastomoses and plexiform lesions in idiopathic pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2016;193:574–6.
5. Norvik C, Westoo CK, Peruzzi N, et al. Synchrotron-based phase-contrast micro-CT as a tool for understanding pulmonary vascular pathobiology and the 3-D microanatomy of alveolar capillary dysplasia. *Am J Physiol Lung Cell Mol Physiol* 2020;318:L65–75.
6. Pietra GG, Capron F, Stewart S, et al. Pathologic assessment of vasculopathies in pulmonary hypertension. *J Am Coll Cardiol* 2004;43:25S–32S.
7. Nickel NP, Yuan K, Dorfmueller P, et al. Beyond the lungs: systemic manifestations of pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2020;201:148–57.
8. Rhodes J. Comparative physiology of hypoxic pulmonary hypertension: historical clues from brisket disease. *J Appl Physiol* (1985) 2005;98:1092–100.
9. Gomez-Arroyo J, Saleem SJ, Mizuno S, et al. A brief overview of mouse models of pulmonary arterial hypertension: problems and prospects. *Am J Physiol Lung Cell Mol Physiol* 2012;302:L977–91.
10. Gomez-Arroyo JG, Farkas L, Alhussaini AA, et al. The monocrotaline model of pulmonary hypertension in perspective. *Am J Physiol Lung Cell Mol Physiol* 2012;302:L363–9.
11. Steiner MK, Syrkin OL, Kolliputi N, et al. Interleukin-6 overexpression induces pulmonary hypertension. *Circ Res* 2009;104:236–44, 28p following 244.
12. Abe K, Toba M, Alzoubi A, et al. Formation of plexiform lesions in experimental severe pulmonary arterial hypertension. *Circulation* 2010;121:2747–54.
13. Jiang B, Deng Y, Suen C, et al. Marked strain-specific differences in the SU5416 rat model of severe pulmonary arterial hypertension. *Am J Respir Cell Mol Biol* 2016;54:461–8.
14. Teichert-Kuliszewska K, Kutryk MJ, Kuliszewski MA, et al. Bone morphogenetic

- protein receptor-2 signaling promotes pulmonary arterial endothelial cell survival: implications for loss-of-function mutations in the pathogenesis of pulmonary hypertension. *Circ Res* 2006;98:209–17.
15. Sakao S, Taraseviciene-Stewart L, Lee JD, et al. Initial apoptosis is followed by increased proliferation of apoptosis-resistant endothelial cells. *FASEB J* 2005;19:1178–80.
 16. Xu W, Erzurum SC. Endothelial cell energy metabolism, proliferation, and apoptosis in pulmonary hypertension. *Compr Physiol* 2011;1:357–72.
 17. Xu W, Koeck T, Lara AR, et al. Alterations of cellular bioenergetics in pulmonary artery endothelial cells. *Proc Natl Acad Sci U S A* 2007;104:1342–7.
 18. Kuebler WM, Nicolls MR, Olschewski A, et al. A pro-con debate: current controversies in PAH pathogenesis at the American Thoracic Society International Conference in 2017. *Am J Physiol Lung Cell Mol Physiol* 2018;315:L502–16.
 19. Michelakis ED. Spatio-temporal diversity of apoptosis within the vascular wall in pulmonary arterial hypertension: heterogeneous BMP signaling may have therapeutic implications. *Circ Res* 2006;98:172–5.
 20. Ranchoux B, Antigny F, Rucker-Martin C, et al. Endothelial-to-mesenchymal transition in pulmonary hypertension. *Circulation* 2015;131:1006–18.
 21. Sheikh AQ, Lighthouse JK, Greif DM. Recapitulation of developing artery muscularization in pulmonary hypertension. *Cell Rep* 2014;6:809–17.
 22. Yu YA, Malakhau Y, Yu CA, et al. Nonclassical monocytes sense hypoxia, regulate pulmonary vascular remodeling, and promote pulmonary hypertension. *J Immunol* 2020;204:1474–85.
 23. Ntokou A, Dave JM, Kauffman AC, et al. Macrophage-derived PDGF-B induces muscularization in murine and human pulmonary hypertension. *JCI Insight* 2021;6(6):e139067.
 24. El-Bizri N, Wang L, Merklinger SL, et al. Smooth muscle protein 22alpha-mediated patchy deletion of *Bmpr1a* impairs cardiac contractility but protects against pulmonary vascular remodeling. *Circ Res* 2008;102:380–8.
 25. Li M, Riddle S, Zhang H, et al. Metabolic reprogramming regulates the proliferative and inflammatory phenotype of adventitial fibroblasts in pulmonary hypertension through the transcriptional corepressor C-terminal binding protein-1. *Circulation* 2016;134:1105–21.
 26. El Kasmi KC, Pugliese SC, Riddle SR, et al. Adventitial fibroblasts induce a distinct proinflammatory/profibrotic macrophage phenotype in pulmonary hypertension. *J Immunol* 2014;193:597–609.
 27. Tuder RM, Voelkel NF. Pulmonary hypertension and inflammation. *J Lab Clin Med* 1998;132:16–24.
 28. Dorfmueller P, Perros F, Balabanian K, et al. Inflammation in pulmonary arterial hypertension. *Eur Respir J* 2003;22:358–63.
 29. Tuder RM, Groves B, Badesch DB, et al. Exuberant endothelial cell growth and elements of inflammation are present in plexiform lesions of pulmonary hypertension. *Am J Pathol* 1994;144:275–85.
 30. Taraseviciene-Stewart L, Nicolls MR, Kraskauskas D, et al. Absence of T cells confers increased pulmonary arterial hypertension and vascular remodeling. *Am J Respir Crit Care Med* 2007;175:1280–9.
 31. Chu Y, Xiangli X, Xiao W. Regulatory T cells protect against hypoxia-induced pulmonary arterial hypertension in mice. *Mol Med Rep* 2015;11:3181–7.
 32. Swain SD, Siemsen DW, Pullen RR, et al. CD4+ T cells and IFN-gamma are required for the development of Pneumocystis-associated pulmonary hypertension. *Am J Pathol* 2014;184:483–93.
 33. Rabinovitch M, Guignabert C, Humbert M, et al. Inflammation and immunity in the pathogenesis of pulmonary arterial hypertension. *Circ Res* 2014;115:165–75.
 34. Schuler R, Efentakis P, Wild J, et al. T cell-derived IL-17a induces vascular dysfunction via perivascular fibrosis formation and dysregulation of (.)NO/cGMP signaling. *Oxid Med Cell Longev* 2019;2019:6721531.
 35. Kumar R, Mickael C, Kassa B, et al. Th2 CD4(+) T cells are necessary and sufficient for schistosoma-pulmonary hypertension. *J Am Heart Assoc* 2019;8:e013111.
 36. Chen G, Zuo S, Tang J, et al. Inhibition of CRTH2-mediated Th2 activation attenuates pulmonary hypertension in mice. *J Exp Med* 2018;215:2175–95.
 37. Daley E, Emson C, Guignabert C, et al. Pulmonary arterial remodeling induced by a Th2 immune response. *J Exp Med* 2008;205:361–72.
 38. Tan SY, Krasnow MA. Developmental origin of lung macrophage diversity. *Development* 2016;143:1318–27.
 39. Mould KJ, Barthel L, Mohning MP, et al. Cell origin dictates programming of resident versus recruited macrophages during acute lung injury. *Am J Respir Cell Mol Biol* 2017;57:294–306.
 40. Gosselin D, Link VM, Romanoski CE, et al. Environment drives selection and function of enhancers controlling tissue-specific macrophage identities. *Cell* 2014;159:1327–40.
 41. Lavin Y, Winter D, Blecher-Gonen R, et al. Tissue-resident macrophage enhancer landscapes are shaped by the local microenvironment. *Cell* 2014;159:1312–26.
 42. Zawia A, Arnold ND, West L, et al. Altered macrophage polarization induces experimental pulmonary hypertension and is observed in patients with pulmonary arterial hypertension. *Arterioscler Thromb Vasc Biol* 2021;41:430–45.

43. Frid MG, Brunetti JA, Burke DL, et al. Hypoxia-induced pulmonary vascular remodeling requires recruitment of circulating mesenchymal precursors of a monocyte/macrophage lineage. *Am J Pathol* 2006;168:659–69.
44. Zaloudikova M, Vytasek R, Vajnerova O, et al. Depletion of alveolar macrophages attenuates hypoxic pulmonary hypertension but not hypoxia-induced increase in serum concentration of MCP-1. *Physiol Res* 2016;65:763–8.
45. Schweitzer F, Tarantelli R, Rayens E, et al. Monocyte and alveolar macrophage skewing is associated with the development of pulmonary arterial hypertension in a primate model of HIV infection. *AIDS Res Hum Retroviruses* 2019;35:63–74.
46. Pugliese SC, Kumar S, Janssen WJ, et al. A time- and compartment-specific activation of lung macrophages in hypoxic pulmonary hypertension. *J Immunol* 2017;198:4802–12.
47. Sawada H, Saito T, Nickel NP, et al. Reduced BMPR2 expression induces GM-CSF translation and macrophage recruitment in humans and mice to exacerbate pulmonary hypertension. *J Exp Med* 2014;211:263–80.
48. Jalce G, Guignabert C. Multiple roles of macrophage migration inhibitory factor in pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 2020;318:L1–9.
49. Le Hiress M, Tu L, Ricard N, et al. Proinflammatory signature of the dysfunctional endothelium in pulmonary hypertension. Role of the macrophage migration inhibitory factor/CD74 complex. *Am J Respir Crit Care Med* 2015;192:983–97.
50. Groth A, Vrugt B, Brock M, et al. Inflammatory cytokines in pulmonary hypertension. *Respir Res* 2014;15:47.
51. Farha S, Sharp J, Asosingh K, et al. Mast cell number, phenotype, and function in human pulmonary arterial hypertension. *Pulm Circ* 2012;2:220–8.
52. Hoffmann J, Yin J, Kukucka M, et al. Mast cells promote lung vascular remodelling in pulmonary hypertension. *Eur Respir J* 2011;37:1400–10.
53. Bartelds B, van Loon RLE, Mohaupt S, et al. Mast cell inhibition improves pulmonary vascular remodeling in pulmonary hypertension. *Chest* 2012;141:651–60.
54. Weng M, Baron DM, Bloch KD, et al. Eosinophils are necessary for pulmonary arterial remodeling in a mouse model of eosinophilic inflammation-induced pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 2011;301:L927–36.
55. Medoff BD, Okamoto Y, Leyton P, et al. Adiponectin deficiency increases allergic airway inflammation and pulmonary vascular remodeling. *Am J Respir Cell Mol Biol* 2009;41:397–406.
56. Paulin R, Michelakis ED. The metabolic theory of pulmonary arterial hypertension. *Circ Res* 2014;115:148–64.
57. Hemnes AR, Luther JM, Rhodes CJ, et al. Human PAH is characterized by a pattern of lipid-related insulin resistance. *JCI Insight* 2019;4:e123611.
58. Luo N, Craig D, Ilkayeva O, et al. Plasma acylcarnitines are associated with pulmonary hypertension. *Pulm Circ* 2017;7:211–8.
59. Brittain EL, Talati M, Fessel JP, et al. Fatty acid metabolic defects and right ventricular lipotoxicity in human pulmonary arterial hypertension. *Circulation* 2016;133:1936–44.
60. Lewis GD, Ngo D, Hemnes AR, et al. Metabolic profiling of right ventricular-pulmonary vascular function reveals circulating biomarkers of pulmonary hypertension. *J Am Coll Cardiol* 2016;67:174–89.
61. Vonk Noordegraaf A, Chin KM, Haddad F, et al. Pathophysiology of the right ventricle and of the pulmonary circulation in pulmonary hypertension: an update. *Eur Respir J* 2019;53:1801900.
62. Ryan JJ, Archer SL. The right ventricle in pulmonary arterial hypertension: disorders of metabolism, angiogenesis and adrenergic signaling in right ventricular failure. *Circ Res* 2014;115:176–88.
63. Sutendra G, Dromparis P, Paulin R, et al. A metabolic remodeling in right ventricular hypertrophy is associated with decreased angiogenesis and a transition from a compensated to a decompensated state in pulmonary hypertension. *J Mol Med* 2013;91:1315–27.
64. Graham BB, Koyanagi D, Kandasamy B, et al. Right ventricle vasculature in human pulmonary hypertension assessed by stereology. *Am J Respir Crit Care Med* 2017;196:1075–7.
65. Graham BB, Kumar R, Mickael C, et al. Vascular adaptation of the right ventricle in experimental pulmonary hypertension. *Am J Respir Cell Mol Biol* 2018;59:479–89.
66. Morrell NW, Aldred MA, Chung WK, et al. Genetics and genomics of pulmonary arterial hypertension. *Eur Respir J* 2019;53:1801899.
67. Wharton K, Derynck R. TGFbeta family signaling: novel insights in development and disease. *Development* 2009;136:3691–7.
68. Evans JD, Girerd B, Montani D, et al. BMPR2 mutations and survival in pulmonary arterial hypertension: an individual participant data meta-analysis. *Lancet Respir Med* 2016;4:129–37.
69. Graf S, Haimel M, Bleda M, et al. Identification of rare sequence variation underlying heritable pulmonary arterial hypertension. *Nat Commun* 2018;9:1416.
70. International PPHC, Lane KB, Machado RD, et al. Heterozygous germline mutations in BMPR2, encoding a TGF-beta receptor, cause familial primary pulmonary hypertension. *Nat Genet* 2000;26:81–4.
71. Upton PD, Morrell NW. TGF-beta and BMPR-II pharmacology—implications for pulmonary vascular diseases. *Curr Opin Pharmacol* 2009;9:274–80.

72. Long L, Ormiston ML, Yang X, et al. Selective enhancement of endothelial BMPRII with BMP9 reverses pulmonary arterial hypertension. *Nat Med* 2015;21:777–85.
73. Yung LM, Yang P, Joshi S, et al. ACTRIIA-Fc rebalances activin/GDF versus BMP signaling in pulmonary hypertension. *Sci Transl Med* 2020;12:eaz5660.
74. Hassoun PM, Mouthon L, Barbera JA, et al. Inflammation, growth factors, and pulmonary vascular remodeling. *J Am Coll Cardiol* 2009;54:S10–9.
75. Tuder RM, Chacon M, Alger L, et al. Expression of angiogenesis-related molecules in plexiform lesions in severe pulmonary hypertension: evidence for a process of disordered angiogenesis. *J Pathol* 2001;195:367–74.
76. Swietlik EM, Greene D, Zhu N, et al. Bayesian inference associates rare KDR Variants with specific phenotypes in pulmonary arterial hypertension. *Circ Genom Precis Med* 2020;14(1):e003155.
77. Izikki M, Guignabert C, Fadel E, et al. Endothelial-derived FGF2 contributes to the progression of pulmonary hypertension in humans and rodents. *J Clin Invest* 2009;119:512–23.
78. Schermuly RT, Dony E, Ghofrani HA, et al. Reversal of experimental pulmonary hypertension by PDGF inhibition. *J Clin Invest* 2005;115:2811–21.
79. Hoepfer MM, Barst RJ, Bourge RC, et al. Imatinib mesylate as add-on therapy for pulmonary arterial hypertension: results of the randomized IMPRES study. *Circulation* 2013;127:1128–38.
80. Dahal BK, Cornitescu T, Tretyn A, et al. Role of epidermal growth factor inhibition in experimental pulmonary hypertension. *Am J Respir Crit Care Med* 2010;181:158–67.
81. Klein M, Schermuly RT, Ellinghaus P, et al. Combined tyrosine and serine/threonine kinase inhibition by sorafenib prevents progression of experimental pulmonary hypertension and myocardial remodeling. *Circulation* 2008;118:2081–90.
82. Tu L, De Man FS, Girerd B, et al. A critical role for p130Cas in the progression of pulmonary hypertension in humans and rodents. *Am J Respir Crit Care Med* 2012;186:666–76.
83. Christman BW, McPherson CD, Newman JH, et al. An imbalance between the excretion of thromboxane and prostacyclin metabolites in pulmonary hypertension. *N Engl J Med* 1992;327:70–5.
84. Stewart DJ, Levy RD, Cernacek P, et al. Increased plasma endothelin-1 in pulmonary hypertension: marker or mediator of disease? *Ann Intern Med* 1991;114:464–9.
85. Brittain EL, Hemnes AR. Vasodilator-responsive idiopathic pulmonary arterial hypertension: evidence for a new disease? *Ann Intern Med* 2015;162:148–9.
86. Galie N, Humbert M, Vachiery JL, et al. 2015 ESC/ERS guidelines for the diagnosis and treatment of pulmonary hypertension: the Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS): endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). *Eur Heart J* 2016;37:67–119.
87. Rich S, Brundage BH. High-dose calcium channel-blocking therapy for primary pulmonary hypertension: evidence for long-term reduction in pulmonary arterial pressure and regression of right ventricular hypertrophy. *Circulation* 1987;76:135–41.
88. Rubin LJ, Groves BM, Reeves JT, et al. Prostacyclin-induced acute pulmonary vasodilation in primary pulmonary hypertension. *Circulation* 1982;66:334–8.
89. Rubin LJ, Mendoza J, Hood M, et al. Treatment of primary pulmonary hypertension with continuous intravenous prostacyclin (epoprostenol). Results of a randomized trial. *Ann Intern Med* 1990;112:485–91.
90. Barst RJ, Rubin LJ, Long WA, et al. A comparison of continuous intravenous epoprostenol (prostacyclin) with conventional therapy for primary pulmonary hypertension. *N Engl J Med* 1996;334:296–301.
91. Pogoriler JE, Rich S, Archer SL, et al. Persistence of complex vascular lesions despite prolonged prostacyclin therapy of pulmonary arterial hypertension. *Histopathology* 2012;61:597–609.
92. Morrell NW, Morris KG, Stenmark KR. Role of angiotensin-converting enzyme and angiotensin II in development of hypoxic pulmonary hypertension. *Am J Physiol* 1995;269:H1186–94.
93. de Man FS, Tu L, Handoko ML, et al. Dysregulated renin-angiotensin-aldosterone system contributes to pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2012;186:780–9.
94. Iyinkkel J, Murray F. GPCRs in pulmonary arterial hypertension: tipping the balance. *Br J Pharmacol* 2018;175:3063–79.
95. Humbert M, Guignabert C, Bonnet S, et al. Pathology and pathobiology of pulmonary hypertension: state of the art and research perspectives. *Eur Respir J* 2019;53:1801887.
96. Rot A. Endothelial cell binding of NAP-1/IL-8: role in neutrophil emigration. *Immunol Today* 1992;13:291–4.
97. Tanaka Y, Adams DH, Shaw S. Proteoglycans on endothelial cells present adhesion-inducing cytokines to leukocytes. *Immunol Today* 1993;14:111–5.
98. Yang T, Li ZN, Chen G, et al. Increased levels of plasma CXC-Chemokine Ligand 10, 12 and 16 are associated with right ventricular function in patients with idiopathic pulmonary arterial hypertension. *Heart Lung* 2014;43:322–7.

99. Hashimoto K, Nakamura K, Fujio H, et al. Epoprostenol therapy decreases elevated circulating levels of monocyte chemoattractant protein-1 in patients with primary pulmonary hypertension. *Circ J* 2004;68:227–31.
100. George PM, Oliver E, Dorfmueller P, et al. Evidence for the involvement of type I interferon in pulmonary arterial hypertension: novelty and significance. *Circ Res* 2014;114:677–88.
101. Zabini D, Heinemann A, Foris V, et al. Comprehensive analysis of inflammatory markers in chronic thromboembolic pulmonary hypertension patients. *Eur Respir J* 2014;44:951–62.
102. Olsson KM, Olle S, Fuge J, et al. CXCL13 in idiopathic pulmonary arterial hypertension and chronic thromboembolic pulmonary hypertension. *Respir Res* 2016;17:21.
103. Kimura H, Okada O, Tanabe N, et al. Plasma monocyte chemoattractant protein-1 and pulmonary vascular resistance in chronic thromboembolic pulmonary hypertension. *Am J Respir Crit Care Med* 2001;164:319–24.
104. Damas JK, Otterdal K, Yndestad A, et al. Soluble CD40 ligand in pulmonary arterial hypertension: possible pathogenic role of the interaction between platelets and endothelial cells. *Circulation* 2004;110:999–1005.
105. Duncan M, Wagner BD, Murray K, et al. Circulating cytokines and growth factors in pediatric pulmonary hypertension. *Mediators Inflamm* 2012;2012:143428.
106. McCullagh BN, Costello CM, Li L, et al. Elevated plasma CXCL12 α is associated with a poorer prognosis in pulmonary arterial hypertension. *PLoS One* 2015;10:e0123709.
107. Kazimierczyk R, Blaszczak P, Jasiewicz M, et al. Increased platelet content of SDF-1 α is associated with worse prognosis in patients with pulmonary arterial hypertension. *Platelets* 2018;30(4):445–51.
108. Hoffmann-Vold AM, Hesselstrand R, Fretheim H, et al. CCL 21 as a potential serum biomarker for pulmonary arterial hypertension in Systemic Sclerosis. *Arthritis Rheumatol* 2018;70(10):1644–53.
109. Heresi GA, AYTEKIN M, Newman J, et al. CXC-chemokine ligand 10 in idiopathic pulmonary arterial hypertension: marker of improved survival. *Lung* 2010;188:191–7.
110. Mamazhakypov A, Viswanathan G, Lawrie A, et al. The role of chemokines and chemokine receptors in pulmonary arterial hypertension. *Br J Pharmacol* 2021;178(1):72–89.
111. Post JM, Hume JR, Archer SL, et al. Direct role for potassium channel inhibition in hypoxic pulmonary vasoconstriction. *Am J Physiol* 1992;262:C882–90.
112. Archer SL, London B, Hampf V, et al. Impairment of hypoxic pulmonary vasoconstriction in mice lacking the voltage-gated potassium channel Kv1.5. *FASEB J* 2001;15:1801–3.
113. Bonnet S, Michelakis ED, Porter CJ, et al. An abnormal mitochondrial-hypoxia inducible factor-1 α -Kv channel pathway disrupts oxygen sensing and triggers pulmonary arterial hypertension in fawn hooded rats: similarities to human pulmonary arterial hypertension. *Circulation* 2006;113:2630–41.
114. Ma L, Roman-Campos D, Austin ED, et al. A novel channelopathy in pulmonary arterial hypertension. *N Engl J Med* 2013;369:351–61.
115. Antigny F, Hautefort A, Meloche J, et al. potassium channel subfamily K member 3 (KCNK3) contributes to the development of pulmonary arterial hypertension. *Circulation* 2016;133:1371–85.
116. Savai R, Al-Tamari HM, Sedding D, et al. Pro-proliferative and inflammatory signaling converge on FoxO1 transcription factor in pulmonary hypertension. *Nat Med* 2014;20:1289–300.
117. Bourgeois A, Lambert C, Habbout K, et al. FOXM1 promotes pulmonary artery smooth muscle cell expansion in pulmonary arterial hypertension. *J Mol Med* 2018;96:223–35.
118. Alastalo TP, Li M, Perez Vde J, et al. Disruption of PPAR γ /beta-catenin-mediated regulation of apelin impairs BMP-induced mouse and human pulmonary arterial EC survival. *J Clin Invest* 2011;121:3735–46.
119. Chun HJ, Bonnet S, Chan SY. Translational advances in the field of pulmonary hypertension. Translating MicroRNA biology in pulmonary hypertension. It will take more than "miR" words. *Am J Respir Crit Care Med* 2017;195:167–78.
120. Santos-Ferreira CA, Abreu MT, Marques CI, et al. Micro-RNA analysis in pulmonary arterial hypertension: current knowledge and challenges. *JACC Basic Transl Sci* 2020;5:1149–62.
121. Omura J, Habbout K, Shimauchi T, et al. Identification of long noncoding RNA H19 as a new biomarker and therapeutic target in right ventricular failure in pulmonary arterial hypertension. *Circulation* 2020;142:1464–84.
122. Zehendner CM, Valasarajan C, Werner A, et al. Long noncoding RNA TYKRIL plays a role in pulmonary hypertension via the p53-mediated regulation of PDGFR β . *Am J Respir Crit Care Med* 2020;202:1445–57.