abstract

Cancer Susceptibility Mutations in Patients With Urothelial Malignancies

Maria I. Carlo, MD¹; Vignesh Ravichandran, MS¹; Preethi Srinavasan, MS¹; Chaitanya Bandlamudi, PhD¹; Yelena Kemel, ScM¹; Ozge Ceyhan-Birsoy, PhD¹; Semanti Mukherjee, PhD¹; Diana Mandelker, MD, PhD¹; Joshua Chaim, DO¹; Andrea Knezevic, MS¹; Satshil Rana, MS¹; Zarina Fnu, MS¹; Kelsey Breen, MS¹; Angela G. Arnold, MS¹; Aliya Khurram, MBBS¹; Kaitlyn Tkachuk¹; Catharine K. Cipolla¹; Ashley Regazzi¹; A. Ari Hakimi, MD¹; Hikmat Al-Ahmadie, MD¹; Guido Dalbagni, MD¹; Karen A. Cadoo, MD¹; Michael F. Walsh, MD¹; Min-Yuen Teo, MBBCh¹; Samuel A. Funt, MD¹; Jonathan A. Coleman, MD¹; Bernard H. Bochner, MD¹; Gopa Iyer, MD¹; David B. Solit, MD¹; Zsofia K. Stadler, MD¹; Liying Zhang, MD¹; Jonathan E. Rosenberg, MD¹; Barry S. Taylor, PhD¹; Mark E. Robson, MD¹; Michael F. Berger, PhD¹; Joseph Vijai, PhD¹; Dean F. Bajorin, MD¹; and Kenneth Offit, MD, MPH¹

PURPOSE Urothelial cancers (UCs) have a substantial hereditary component, but, other than their association with Lynch syndrome, the contribution of genetic risk factors to UC pathogenesis has not been systematically defined. We sought to determine the prevalence of pathogenic/likely pathogenic (P/LP) germline variants in patients with UC and identify associated clinical factors.

PATIENTS AND METHODS Overall, 586 patients with UC underwent prospective, matched tumor-normal DNA sequencing. Seventy-seven genes associated with cancer predisposition were analyzed; allele frequencies were compared with publicly available database.

RESULTS P/LP germline variants were identified in 80 (14%) of 586 individuals with UC. The most common P/LP variants in high- or moderate-penetrance genes were *BRCA2* (n = 9; 1.5%), *MSH2* (n = 8; 1.4%), *BRCA1* (n = 8; 1.4%), *CHEK2* (n = 6; 1.0%), *ERCC3* (n = 4; 0.7%), and *NBN* and *RAD50* (n = 3; 0.5% each). Sixty-six patients (83%) had germline P/LP variants in DNA-damage repair (DDR) genes, of which 28 (42%) had biallelic inactivation. Patients with P/LP variants were more commonly diagnosed at an early age (22% v 6% in those without variants; P = .01). *BRCA2* and *MSH2* were significantly associated with an increased risk for UC (odds ratio, 3.7 [P = .004] and 4.6 [P = .001], respectively). Current clinical guidelines for referral for genetic testing failed to identify 6 (26%) patients with high-penetrance variants.

CONCLUSION Clinically significant P/LP germline variants in DDR genes frequently are present in patients with advanced UC. The presence of DDR germline variants could guide cancer screening for patients and their families and serve as predictive biomarkers of response to targeted or immunotherapies. Family history–based criteria to identify patients with hereditary UC susceptibility are insensitive. Broader germline testing in UC, particularly in those of young ages, should be considered.

J Clin Oncol 37. © 2019 by American Society of Clinical Oncology

INTRODUCTION

Urothelial cancer (UC) has a substantial hereditary component, with an estimated 30% heritable fraction according to epidemiologic studies.¹ Family history of UC confers a twofold increased risk, with numerous reports of multiple-case UC kindreds.²⁻⁶ The heritable mechanisms underlying familial aggregations and early-onset UC remain unknown, and there are few syndromic associations with known cancer susceptibility genes. Highly penetrant cancer susceptibility genes, such as those in the mismatch-repair (MMR) pathway, only account for a small fraction of inherited UC susceptibility; mutations in MMR-associated genes are found primarily in patients with tumors of the ureter and renal pelvis.^{7.8}

Beyond their role in tumor pathogenesis, germline variants can be predictors of response to cancer therapies that have activities enhanced by DNA repair

defects.9,10 Deficient MMR status and high microsatellite instability (MSI-H), hallmarks of tumors in patients with Lynch syndrome, are predictive of response to the PD-1 inhibitor pembrolizumab.¹¹ In UC, inactivating somatic mutations in ERCC2, ATM, RB1, and FANCC have been associated with response to cisplatin-based neoadjuvant chemotherapy.^{12,13} DNAdamage repair (DDR) mutations, both somatic and germline, may independently predict response to checkpoint inhibitors in patients with UC.¹⁴ The substantial heritability of UC, the incomplete understanding of the genes and pathways responsible for this increased heritable risk, and the potential that identification of germline variants could help guide therapeutic decisions provide a strong rationale to investigate putative UC susceptibility genes.

We have shown that germline mutations are commonly identified in individuals undergoing tumor-normal next-generation sequencing (NGS) and may reveal

ASSOCIATED CONTENT Appendix

Author affiliations and support information (if applicable) appear at the end of this article.

Accepted on November 6, 2019 and published at jco.org on December 3, 2019: D01 https://doi.org/10. 1200/JC0.19.01395



Journal of Clinical Oncology®

previously unknown genetic associations.^{15,16} Using a matched tumor-germline NGS platform, we determined the prevalence of pathogenic/likely pathogenic (P/LP) germline variants in 77 cancer-associated genes in patients with UC, and we examined associations between germline status and somatic mutational profile.

PATIENTS AND METHODS

Patient Cohort

Beginning in May 2014, patients with UC seen at Memorial Sloan Kettering Cancer Center were offered matched tumor-germline DNA sequencing at physician discretion under an institutional protocol (ClinicalTrials.gov identifier: NCT01775072), with only somatic variants reported. Baseline clinical characteristics for all enrolled patients were collected from institutional electronic medical records. Age \leq 45 years was prospectively defined as early onset (outside the 95% CI of the median age of UC diagnosis).¹⁷ Starting in May 2015, patients could opt in to receive results of a secondary germline analysis of genes associated with increased heritable cancer risk; 169 patients consented to disclosure of germline results. For these 169 patients (identified cohort), germline variants were associated with a broad range of clinical features. For the remaining 417 patients (anonymized cohort), analysis was performed in a permanently anonymized fashion with no clinical annotation beyond tumor subtype. The study was approved by the Memorial Sloan Kettering Cancer Center institutional review board.

Sequencing and Results Reporting

Paraffin-embedded tumor and blood from patients were obtained and sequenced using the MSK-IMPACT platform, a capture-based NGS assay capable of identifying mutations, copy number alterations, and select gene fusions involving 341 cancer-associated genes in the first iteration and 468 in the more recent iteration, as described previously (gene list; Appendix Table A1, online only).^{18,19} For the anonymized cases, sequence data were assigned a unique study identifier and irretrievably de-linked from personal identifiers before variant calling.

Variant Interpretation

Pathogenicity was determined according to American College of Medical Genetics criteria (updated as of January 2018).²⁰ Germline variants in 77 cancer predisposition genes were prioritized using PathoMAN, an automated germline variant classification tool for cancer; variants from the Exome Aggregation Consortium (ExAC) database without The Cancer Genome Atlas (TCGA) alleles were similarly prioritized.²¹ In addition, manual curation of the dataset was performed by a research genetic counselor (Y.K.), and any differences in variant calls were resolved with review by a molecular geneticist (O.C.B.) and a cancer geneticist (M.I.C.). For these 77 genes, all coding regions were sequenced in both the germline and the tumor. P/LP

germline variants (associated with disease causation) were included in this analysis; variants of unknown significance were reviewed but were not reported. According to known disease risks and prior modeling, P/LP variants were classified at the gene level as high penetrance (relative risk [RR] of disease, > 4), moderate penetrance (RR, 2-4), lowpenetrance (RR, < 2), or uncertain penetrance, or they were associated with an autosomal recessive condition.²² For *CHEK2*, *APC*, and *ERCC3*, classification was performed at the variant level: *APC* p.lle1307Lys and *CHEK2* p.lle157Thr were considered low or uncertain penetrance, and the *ERCC3* p.Arg109X, moderate penetrance.²³ Tumor sequencing results were available for all patients. The FACETS algorithm was used to evaluate loss of heterozygosity (LOH) at the locus of the germline variant.²⁴

We identified 34 genes within the MSK-IMPACT panel as related to DDR, as previously described (Appendix Table A2, online only).²⁵ Within DDR genes, *MSH2, MSH6, MLH1,* and *PMS2* were classified as MMR pathway genes; the remaining DDR genes were classified as other DDR.

Comparison of Guidelines Based Versus Agnostic Testing for Cancer Predisposition Syndromes in the Identified Cohort

Family history, religion, and race/ethnicity were selfreported and collected from the medical record or at time of genetic counseling. Published guidelines based on personal and family history were used to determine which genetic tests would be indicated according to the age at onset of cancer, personal or family history of cancer, and self-reported Ashkenazi Jewish ancestry.^{26,27} A pathogenic variant was considered incremental if it was detected by sequencing in this study but would not have been identified by genetic testing through application of current clinical guidelines.

Statistical Analysis

Allele frequencies of P/LP variants in the identified and anonymized cohorts were compared with allele frequencies of P/LP variants in the ExAC without cases from TCGA, as described previously.²¹ Ashkenazi Jewish founder mutations were excluded (Appendix Table A3, online only). Fisher's exact 2-sided test was performed to assess differences in frequency; odds ratios with 95% intervals were reported. The α used to determine statistical significance in the Fisher's test and the 95% confidence limits was adjusted using the Bonferroni correction for multiple comparisons. In the identified cohort, clinical characteristics of patients with germline P/LP variants were compared with those without them using Fisher's exact test. Statistical analysis was performed using SAS 9.4 (Cary, NC) and R version 3.3.3. Response to therapy in identified patients with MMR mutations treated with immunotherapy was assessed according to RECIST v1.1 criteria.

RESULTS

Patient Characteristics

Five hundred eighty-six patients had tumor-germline profiling using MSK-IMPACT; 417 consented to receive only tumor sequencing results (anonymized cohort), and 169 consented to receive both tumor and germline sequencing results (identified cohort). Clinical characteristics of patients are listed in Table 1. Patients were primarily men (74.1%) and had a median age of 63 years (range, 25-87 years); 42 (7.2%) were \leq 45 years of age. Only 42 patients (7.2%) had a family history of bladder or UC, and 112 (19.1%) had a history of a second malignancy other than UC. Most patients (79.0%) had bladder as the primary tumor site, and 59.7% had or developed metastatic disease during the period of clinical follow-up.

Frequency and Spectrum of Germline Variants

Eighty-six P/LP variants were identified in 80 individuals (13.7%), including 62 patients (13.4%) with bladder and 18 (15.8%) with upper tract (UT) tumors (Fig 1). Eleven patients had two P/LP variants each. The most frequently mutated moderate- or high-penetrance genes were BRCA2 (n = 9; 1.5%), *MSH2* (n = 8; 1.4%), *BRCA1* (n = 8; 1.4%), CHEK2 (n = 6; 1.0%), ERCC3 (n = 4; 0.7%), and NBN and RAD50 (n = 3; 0.5% each; Appendix Table A4, online only). The low-penetrance variant APC p. Ile1307Lys was the most prevalent overall (n = 11), and 6 (35.3%) of the 17 BRCA1/2 variants were Ashkenazi Jewish founder mutations. Of all variants, 35 (40.7%) were of high penetrance, 24 (27.9%) were of moderate penetrance, 18 (20.9%) were of low or uncertain penetrance, and 10 (11.6%) were in a gene associated with an autosomal recessive cancer-associated syndrome. Sixty-five variants (75.6%) were in genes associated with DDR; of these, 12 were in the MMRassociated genes MSH2 (n = 8), MSH6 (n = 2), and MLH1 (n = 2; Fig 2A).

Correlation Between Germline Genotype and Tumor Phenotype

Of 54 germline variants in DDR genes (excluding MMR genes), LOH/somatic mutation in the tumor was present in 18 patients (33.3%), including carriers of ATM (n = 2 of 3), ERCC2 (n = 3 of 3), BRCA2 (n = 6 of 9), and BRCA1 (n = 2 of 8), among others (Fig 2B). In the identified cohort, all tumors from patients with a germline MMR variant had either MSI-H status or immunohistochemistry showing deficient MMR protein staining.

Estimate of UC Risk Associated With Observed Variants

To estimate population frequencies of germline P/LP variants in genes seen in the UC cohort, we analyzed allele frequencies of these genes in the ExAC dataset. *BRCA2* and *MSH2* showed statistically significant increased risk in UC patients compared with ExAC (odds ratio, 3.7; 95% Cl, 1.5 to 7.8; P < .003 for *BRCA2*; and odds ratio, 4.6; 95% Cl, 1.8 to 9.8; P < .001 for *MSH2*; Table 2).

Clinical Characteristics Associated With P/LP Variants

The only clinical characteristic available for analysis for patients in the anonymized cohort was site of tumor. In patients with bladder primaries from both the anonymized and identified cohorts, 39 (8.4%) had moderate- or high-risk P/LP variants. Conversely, in patients with UT primaries, 16 (14.0%) had moderate- or high-risk P/LP variants, including 10 (8.8%) with MMR mutations.

In the identified cohort, patients with P/LP variants were more likely to have early age of onset (age \leq 45 years) compared with patients with no germline variant (22% v 6%; P = 0.01; Table 3). Six of 14 patients with early-onset UC had germline variants, including in *MSH2* (n = 2), *BRCA1* (n = 2), *MSH6* (n = 1), and *BRCA2* (n = 1). Presence of P/LP variants was not statistically significantly associated with a positive family history of UC, non-UC malignancy, smoking history, or metastasis at diagnosis, but presence was associated with Ashkenazi Jewish ancestry, reflective of founder mutations in *BRCA1*, *BRCA2*, and *CHEK2*.

Of 9 patients with MMR variants in the identified cohort, all but one had a UT tumor as the primary site of malignancy. The median age of diagnosis of UC was 58 years (range, 31-84 years), and UC was the first malignancy in 6 patients. Although none reported a family history of bladder or UT cancer, 4 reported relatives with cancers of unknown origin or kidney cancer, not otherwise specified.

Given the association of MSI-H status or deficient MMR tumors and response to immunotherapy in other malignancies, we explored response in the four patients in the identified cohort who had germline MMR variants (all MSI-H tumors) and who had received immunotherapy. All 4 had presented initially with metastatic disease and had received platinum chemotherapy as first-line treatment. Three experienced progression of disease after platinum chemotherapy followed by a complete response after immunotherapy (Appendix Fig A1, online only). The fourth patient experienced progression of disease on both platinum chemotherapy and immunotherapy.

Comparisons to Clinical Genetics Referral Criteria

In the identified cohort, detailed family history was obtained by interview for 29 (85.3%) of the patients with P/LP variants; for the remainder of patients, data were extracted from the electronic medical record. Only 9 of 27 patients with high- or moderate-penetrance variants had undergone clinical genetic testing or attended clinical genetic counseling before receiving genetic test results through the protocol. Of patients with high-penetrance P/LP variants, 6 patients (26.3%) would not have been referred for germline testing according to published guidelines (one each with *MSH2, MSH6, BRCA1, SDHA*, and *TP53*). The patient with a *TP53* variant, diagnostic of Li-Fraumeni syndrome, had a father with bladder cancer, but the patient did not meet criteria for any genetic testing. Of patients with

TABLE 1. Patient Characteristics

		No. (%)	
Characteristic	Identified Cohort (n = 169)	Anonymized Cohort $(n = 417)$	Total (N = 586)
Age, years			
Median (range)	63 (31-84)	64 (25-87)	63 (25-87)
\leq 45 years	14 (8.3)	28 (6.7)	42 (7.2)
≥ 46	155 (91.7)	388 (93.0)	543 (92.7)
Sex			
Female	41 (24.3)	111 (26.6)	152 (25.9)
Male	128 (75.7)	306 (73.4)	434 (74.1)
Race or ethnic background			
White	146 (86.4)	366 (87.8)	512 (87.4)
Hispanic	2 (1.2)	4 (1.0)	6 (1.0)
African American	5 (3.0)	13 (3.1)	18 (3.1)
Asian or Pacific Islander	5 (3.0)	12 (2.9)	17 (2.9)
Other or unknown	11 (6.5)	22 (5.3)	33 (5.6)
Ashkenazi Jewish ancestry			
Yes	31 (18.3)	76 (18.2)	107 (18.3)
No	97 (57.4)	220 (52.8)	317 (54.1)
Unknown	41 (24.3)	121 (29.0)	162 (27.7)
Other primary malignancy			
Yes	39 (23.1)	73 (17.5)	112 (19.1)
No	130 (76.9)	343 (82.3)	473 (80.7)
Unknown	0	1 (0.2)	1 (0.2)
Family history of urothelial cancer			
Yes	18 (10.7)	24 (5.8)	42 (7.2)
No	151 (89.3)	391 (93.8)	542 (92.5)
Unknown	0	2 (0.5)	2 (0.3)
Tobacco use history			
Ever	103 (60.9)	273 (65.5)	376 (64.2)
Never	66 (39.1)	142 (34.1)	208 (35.5)
Unknown	0	2 (0.5)	2 (0.3)
Site of primary malignancy			
Bladder/urethra	117 (69.2)	346 (83.0)	463 (79.0)
Renal pelvis/ureter	48 (28.4)	66 (15.8)	114 (19.5)
Both or unknown	4 (2.4)	5 (1.2)	9 (1.5)
Histologic subtype			
Urothelial carcinoma	169 (100.0)	393 (94.2)	562 (95.9)
Adenocarcinoma	0	16 (3.8)	16 (2.7)
Other	0	8 (1.9)	8 (1.4)
Stage at diagnosis			
Non-muscle invasive bladder	44 (26.0)	209 (50.1)	253 (43.2)
Muscle-invasive bladder	56 (33.1)	102 (24.5)	158 (27.0)
Localized upper tract	35 (20.7)	51 (12.2)	86 (14.7)
Metastatic	34 (20.1)	43 (10.3)	77 (13.1)
Unknown/other	0	12 (2.9)	12 (2.1)
	(continued on following	page)	

 ${\bf 4} \, \, {\ensuremath{\mathbb C}}$ 2019 by American Society of Clinical Oncology

TABLE 1. Patient Characteristics (continued)

	NO. (%)			
Characteristic	Identified Cohort (n = 169)	Anonymized Cohort (n = 417)	Total (N = 586)	
Stage at time of analysis				
Nonmetastatic	53 (31.4)	182 (43.7)	235 (40.1)	
Metastatic	116 (68.6)	234 (56.1)	350 (59.7)	
Unknown	0	1 (0.2)	1 (0.2)	

NOTE. Cohort demographic and clinical characteristics are provided for the 586 patients who consented to tumor-normal testing. Patients in the identified cohort additionally consented to receive germline results.

moderate-penetrance variants, 7 (87.5%) would not have been referred (Appendix Table A5, online only).

DISCUSSION

This study shows that, in patients with UC, clinically significant P/LP germline variants, particularly in DDR genes, frequently are present. Multiple epidemiologic studies have identified an increased familial risk of UC. However, to date, the only identified hereditary cancer syndrome associated with increased UC risk is Lynch syndrome, which is caused by inactivating mutations in the MMR-associated genes MSH2, MSH6, MLH1, PMS2, and EPCAM.^{3,28,29} Patients with Lynch syndrome have an up to 12% cumulative risk of urinary tract cancer; although the risk is greater for UT UC, there also may be an increased risk for bladder UC.^{7,8,30,31} Studies estimating prevalence of Lynch syndrome in patients with UC, however, have been limited. In two studies looking at unselected patients with UT UC, 7% of tumors had deficient MMR protein expression or were MSI-H, usually a necessary but not sufficient biomarker for Lynch syndrome.^{32,33} In our study, 2.1% overall and 8.7% of patients with UT tumors had Lynch syndrome, and, for those in the identified cohort, 6 of 9 had UC as the first malignancy. Immunohistochemistry analysis of tumors for loss of MMR protein expression or tumor MSI analysis is standard practice for colorectal and endometrial cancers, for which the incidence of Lynch syndrome ranges from 2% to 5%.³⁴ A recent pan-cancer study showed that, among patients with MSI-high/intermediate UC, 37.5% had a germline MMR variant diagnostic of Lynch syndrome, the highest prevalence of all cancer types analyzed.³⁵ The increased incidence of Lynch syndrome in our cohort, and the high prevalence of Lynch syndrome in those with MSI-high/intermediate tumors, supports consideration of germline or tumor screening for those with UT UC.

N- (0/)

In our cohort, 9% of patients had a germline DDR mutation in a gene other than those in MMR. Dysregulation of DNA repair is implicated in the carcinogenesis of UC. DDR somatic mutations are frequent in UC.^{25,36} For example, the nucleotide excision repair pathway gene *ERCC2* was somatically mutated in 9% of UC in TCGA, and common polymorphisms in *ERCC2*, *NBN*, and *XPC* are associated



FIG 1. Frequency and penetrance of germline variants in patients with urothelial cancer. Frequency of pathogenic/likely pathogenic (P/LP) germline variants identified in 586 patients with UC. Pie chart shows that 13.7% (n = 80 patients) had a germline P/LP variant; 9.7% (n = 57) had high- or moderate-penetrance variants.

Journal of Clinical Oncology





FIG 2. Germline pathogenic/likely pathogenic variants by site of tumor origin and tumors with loss of heterozygosity (LOH) or second somatic mutation in the tumor in 586 patients. (A) Number of mutations in each gene by tumor type and (B) in germline-positive occurrences with LOH or second somatic mutation in same locus. DDR, DNA-damage repair; MMR, mismatch repair.

with bladder cancer risk.^{13,37,38} To further investigate the pathogenic role of DDR genes, we compared allele frequencies of the most frequently mutated genes in UC with frequencies in individuals in ExAC without cancer, and we found significantly increased frequencies of mutations in richment for second allele loss in tumors for germline

BRCA2 and MSH2 in UC. We also found LOH in 6 of 9 tumors in patients with germline variants in BRCA2 and 3 of 3 tumors in patients with germline ERCC2 variants. Of note, the current number of carriers is too small to analyze en-

Gene	UC Occurrence Allele Count	UC Occurrence Allele Number	Allele Count ExAC	Allele ExAC	Odds Ratio (95% CI)	Р
ATM	3	1,172	183	106,194	1.49 (0.15 to 5.7)	.46
BRCA1	4	1,164	85	106,206	4.29 (0.67 to 14.6)	.02
BRCA2	7	1,168	173	106,187	3.68 (1.01 to 9.5)	.004*
MSH2	7	1,172	133	102,035	4.58 (1.26 to 11.9)	.001*
NBN	3	1,172	65	106,189	4.18 (0.43 to 16.7)	.04
RAD50	3	1,172	392	106,179	0.69 (0.07 to 2.6)	.81

TABLE 2. Comparison of Allele Frequencies in All Patients Versus ExAC

NOTE. Odds ratio and CIs calculated with Bonferroni correction. Only genes with three or more occurrences in the UC cohort were considered. Ashkenazi Jewish BRCA1/2 founder mutations were excluded.

Abbreviation: UC, urothelial carcinoma.

*Statistically significant with Bonferroni-corrected $\alpha = .05/6 = .0083$.

TABLE 3.	Clinical	Characteristics	by	Moderate-/High-Penetrance	Variants	in
Identified	Cohort					

Characteristic	Any Moderate/High Penetrance (n = 27)	No Moderate/High Penetrance (n = 142)	Р
Age, years			
≤ 45	6 (22)	8 (6)	.01
≥ 46	21 (78)	134 (94)	
Sex			
Male	19 (70)	109 (77)	.47
Female	8 (30)	33 (23)	
Ashkenazi Jewish ancestry			
Yes	11 (41)	20 (14)	.005
No	13 (48)	84 (59)	
Unknown	3 (11)	38 (27)	
Tobacco use history			
Current or former	15 (56)	88 (62)	.53
Never	12 (44)	54 (38)	
Family history of UC			
Yes	2 (7)	17 (12)	.74
No	25 (93)	125 (88)	
History of second malignancy			
Yes	7 (26)	32 (23)	.80
No	20 (74)	110 (77)	
Site of primary malignancy			
Bladder/urethra	16 (59)	101 (72)	.32
Renal pelvis/ureter	11 (41)	37 (26)	
Both/unknown	0	2 (1)	
Stage at diagnosis			
Nonmetastatic	24 (89)	111 (78)	.30
Metastatic	3 (11)	31 (22)	
Stage at last follow-up*			
Nonmetastatic	12 (44)	41 (29)	.12
Metastatic	15 (56)	101 (71)	

Abbreviation: UC, urothelial carcinoma.

*Median length of follow-up was 4.1 years (range, 1.5-23.2 years).

events compared with the background loss of first somatic allele in these genes. Pathogenic nucleotide excision repair germline mutations have not been described previously in UC, although heterozygous germline mutations in *ERCC2* and *ERCC3* have been associated with increased risk of sarcoma and breast cancer.^{38,39} Additional studies will be needed to determine the role of these germline mutations in the pathogenicity of UC.

Germline MMR and other DDR mutations have potential implications for treatment selection. Somatic mutations in

ERCC2, ATM, RB1, FANCC, and other DDR genes are correlated with improved responses to platinum-based chemotherapy in patients with UC.^{12,13,40,41} Whether germline pathogenic mutations in these genes also are associated with improved outcomes must be explored. Preclinical models with ERCC2 and ERCC3 heterozygous knockouts show a hypomorphic functionality when exposed to DNA damaging agents, supporting a plausible mechanism for sensitivity to platinum therapies.^{23,25} Although tumor MMR deficiency predicts response to PD-1 blockade in other solid tumors, to date, no large studies correlating MMR deficiency and response to PD-1 or PD-L1 blockade have included patients with UC.¹¹ In this study, all patients with germline MMR mutations had MSI-H or MMRdeficient tumors, and 3 of 4 patients with Lynch syndrome had complete responses to immunotherapy after they experienced progression on chemotherapy.^{11,42} Our findings suggest that germline DDR alterations should be included with somatic alterations when assessing for correlations between therapeutic benefit.

Finally, a quarter of patients with high-penetrance germline variants would not have been detected by guidelinesdirected testing. Identification of germline mutations in these patients may allow for enhanced screening and early detection of hereditary cancers in those families for whom testing would not have been undertaken. Several individuals became the index cases (first detected cancer) in their families, which then led to cascade testing of other family members. Interestingly, among those with a family history of UC or personal history of other cancers, there was no increased incidence of P/LP germline variants. This may be explained in part by incomplete information available to patients—for example, of 8 patients with Lynch syndrome, none reported a family history of bladder or UT cancer, but 4 did report relatives with possible kidney cancer or of unclear origin, which may reflect UT cancers. Analysis in a larger cohort also may reveal whether personal history of cancer is associated with presence of germline alterations.

This study had several limitations. It was retrospective in nature and had a limited sample size; its findings will require validation in other cohorts. Given the smaller number of identified patients with clinically annotated data, associations between clinical features and prevalence of mutations may be limited by numbers. The study was conducted at a comprehensive cancer center with regional referral patterns, so the patient population may differ from that in the general community. For example, the median age of diagnosis in the cohort was 63 years, compared with approximately 70 years in the United States, and there was under-representation of nonwhite patients.43 We did observe a lower prevalence of germline mutations in the anonymized cohort, in which germline results were not returned. Although physicians were not instructed to select patients according to suspicion of an inherited syndrome, individuals consenting to return of germline results may have be more motivated to do so because of family history. We performed targeted exome sequencing of known cancerpredisposition genes; agnostic whole-exome sequencing could yield associations among novel genes or variants and risk of UC. LOH analysis was exploratory and was not corrected for possible increased background LOH in those genes, which also merits more study in larger cohorts.

This study demonstrates that germline mutations in patients with UC occur predominantly in DDR genes, with

AFFILIATION

¹Memorial Sloan Kettering Cancer Center, New York, NY

CORRESPONDING AUTHOR

Maria I. Carlo, MD, 353 East 68th St, New York, NY 10065; e-mail: carlom@mskcc.org.

EQUAL CONTRIBUTION

D.F.B. and K.O. contributed equally to this work.

PRIOR PRESENTATION

Presented at the 2018 Genitourinary Cancers Symposium, San Francisco, CA, February 8-10, 2018, and American Society of Clinical Oncology 2017 Annual Meeting, Chicago, IL, June 2-6, 2019.

SUPPORT

Supported by the Bladder Specialized Program of Research Excellence grant No. P50 CA221745-01A1 from the National Cancer Institute, The Robert and Kate Niehaus Center for Inherited Cancer Genomics at Memorial Sloan Kettering Cancer Center, the Marie-Josée and Henry R. Kravis Center for Molecular Oncology, the Andrew Sabin Cancer Research Fund, the Carmel Family Cancer Research Fund, and Cancer Center Support Grant No. P30 CA008748-50.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST AND DATA AVAILABILITY STATEMENT

Disclosures provided by the authors and data availability statement (if applicable) are available with this article at DOI https://doi.org/10.1200/JC0.19.01395.

increased frequency of *BRCA2* and *MSH2*. Traditional criteria to identify those at risk for hereditary syndromes only identified a fraction of patients. There is potential value of expanded germline analysis in UC, particularly in patients of young age at diagnosis and those with UT tumors. The genes found to be mutated were associated with increased risk for cancers other than UC; thus, their identification likely will have substantial implications for directed cancer screening in patients and their families.

AUTHOR CONTRIBUTIONS

Conception and design: Maria I. Carlo, Vignesh Ravichandran, A. Ari Hakimi, Guido Dalbagni, Samuel A. Funt, David B. Solit, Jonathan E. Rosenberg, Mark E. Robson, Joseph Vijai, Dean F. Bajorin, Kenneth Offit **Collection and assembly of data:** Maria I. Carlo, Vignesh Ravichandran, Chaitanya Bandlamudi, Yelena Kemel, Joshua Chaim, Zarina Fnu, Angela G. Arnold, Aliya Khurram, Kaitlyn Tkachuk, Catharine K. Cipolla, Ashley Regazzi, Hikmat Al-Ahmadie, Michael F. Walsh, Min-Yuen Teo, Samuel A. Funt, Bernard H. Bochner, David B. Solit, Liying Zhang, Jonathan E. Rosenberg, Barry S. Taylor, Mark E. Robson, Michael F. Berger, Joseph Vijai, Dean F. Bajorin, Kenneth Offit

Administrative support: Aliya Khurram, Catharine K. Cipolla, Ashley Regazzi

Data analysis and interpretation: Maria I. Carlo, Vignesh Ravichandran, Preethi Srinavasan, Chaitanya Bandlamudi, Yelena Kemel, Ozge Ceyhan-Birsoy, Semanti Mukherjee, Diana Mandelker, Joshua Chaim, Andrea Knezevic, Satshil Rana, Kelsey Breen, Hikmat Al-Ahmadie, Karen A. Cadoo, Samuel A. Funt, Jonathan A. Coleman, Bernard H. Bochner, Gopa Iyer, David B. Solit, Zsofia K. Stadler, Liying Zhang, Jonathan E. Rosenberg, Mark E. Robson, Michael F. Berger, Joseph Vijai, Dean F. Bajorin, Kenneth Offit

Provision of study material or patients: Guido Dalbagni, Min-Yuen Teo, Bernard H. Bochner, David B. Solit, Liying Zhang, Jonathan E. Rosenberg, Kenneth Offit

Administrative support: David B. Solit, Dean F. Bajorin, Kenneth Offit Financial support: David B. Solit, Kenneth Offit

Manuscript writing: All authors

Final approval of manuscript: All authors Accountable for all aspects of the work: All authors

REFERENCES

- 1. Mucci LA, Hjelmborg JB, Harris JR, et al: Familial risk and heritability of cancer among twins in Nordic countries. JAMA 315:68-76, 2016
- 2. Kramer AA, Graham S, Burnett WS, et al: Familial aggregation of bladder cancer stratified by smoking status. Epidemiology 2:145-148, 1991
- 3. Lin J, Spitz MR, Dinney CP, et al: Bladder cancer risk as modified by family history and smoking. Cancer 107:705-711, 2006
- 4. Lynch HT, Walzak MP, Fried R, et al: Familial factors in bladder carcinoma. J Urol 122:458-461, 1979
- 5. Mahboubi AO, Ahlvin RC, Mahboubi EO: Familial aggregation of urothelial carcinoma. J Urol 126:691-692, 1981
- 6. McCullough DL, Lamma DL, McLaughlin AP III, et al: Familial transitional cell carcinoma of the bladder. J Urol 113:629-635, 1975
- Joost P, Therkildsen C, Dominguez-Valentin M, et al: Urinary tract cancer in Lynch syndrome: Increased risk in carriers of MSH2 mutations. Urology 86: 1212-1217, 2015
- van der Post RS, Kiemeney LA, Ligtenberg MJ, et al: Risk of urothelial bladder cancer in Lynch syndrome is increased, in particular among MSH2 mutation carriers. J Med Genet 47:464-470, 2010
- 9. Robson M, Im SA, Senkus E, et al: Olaparib for metastatic breast cancer in patients with a germline BRCA mutation. N Engl J Med 377:523-533, 2017
- 10. Mateo J, Carreira S, Sandhu S, et al: DNA-repair defects and olaparib in metastatic prostate cancer. N Engl J Med 373:1697-1708, 2015
- 11. Le DT, Durham JN, Smith KN, et al: Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science 357:409-413, 2017
- 12. Plimack ER, Dunbrack RL, Brennan TA, et al: Defects in DNA repair genes predict response to neoadjuvant cisplatin-based chemotherapy in muscle-invasive bladder cancer. Eur Urol 68:959-967, 2015
- 13. Van Allen EM, Mouw KW, Kim P, et al: Somatic *ERCC2* mutations correlate with cisplatin sensitivity in muscle-invasive urothelial carcinoma. Cancer Discov 4: 1140-1153, 2014

- 14. Teo MY, Seier K, Ostrovnaya I, et al: Alterations in DNA damage response and repair genes as potential marker of clinical benefit from PD-1/PD-L1 blockade in advanced urothelial cancers. J Clin Oncol 36:1685-1694, 2018
- 15. Schrader KA, Cheng DT, Joseph V, et al: Germline variants in targeted tumor sequencing using matched normal DNA. JAMA Oncol 2:104-111, 2016
- Mandelker D, Zhang L, Kemel Y, et al: Mutation detection in patients with advanced cancer by universal sequencing of cancer-related genes in tumor and normal DNA vs guideline-based germline testing. JAMA 318:825-835, 2017
- Howlader N, Noone AM, Krapcho M (eds): SEER Cancer Statistics Review, 1975-2016, Bethesda, MD, National Cancer Institute. https://seer.cancer.gov/csr/ 1975_2016/
- Cheng DT, Mitchell TN, Zehir A, et al: Memorial Sloan Kettering-integrated mutation profiling of actionable cancer targets (MSK-IMPACT): A hybridization capture-based next-generation sequencing clinical assay for solid tumor molecular oncology. J Mol Diagn 17:251-264, 2015
- 19. Zehir A, Benayed R, Shah RH, et al: Mutational landscape of metastatic cancer revealed from prospective clinical sequencing of 10,000 patients. Nat Med 23: 703-713, 2017
- Richards S, Aziz N, Bale S, et al: Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med 17:405-424, 2015
- 21. Ravichandran V, Shameer Z, Kemel Y, et al: Toward automation of germline variant curation in clinical cancer genetics. Genet Med 21:2116-2125, 2019
- 22. Tung N, Domchek SM, Stadler Z, et al: Counselling framework for moderate-penetrance cancer-susceptibility mutations. Nat Rev Clin Oncol 13:581-588, 2016
- 23. Vijai J, Topka S, Villano D, et al: A Recurrent ERCC3 truncating mutation confers moderate risk for breast cancer. Cancer Discov 6:1267-1275, 2016
- 24. Shen R, Seshan VE: FACETS: Allele-specific copy number and clonal heterogeneity analysis tool for high-throughput DNA sequencing. Nucleic Acids Res 44: e131, 2016
- Teo MY, Bambury RM, Zabor EC, et al: DNA damage response and repair gene alterations are associated with improved survival in patients with platinumtreated advanced urothelial carcinoma. Clin Cancer Res 23:3610-3618, 2017
- 26. National Comprehensive Cancer Network: Guidelines: Genetic/familial high-risk assessment—breast and ovarian, 2019. https://www.nccn.org/professionals/ physician_gls/pdf/genetics_screening.pdf
- 27. National Comprehensive Cancer Network: Guidelines: Genetic/familial high-risk assessment—Colorectal, 2018. https://www.nccn.org/professionals/ physician_gls/pdf/genetics_colon.pdf
- Murta-Nascimento C, Silverman DT, Kogevinas M, et al: Risk of bladder cancer associated with family history of cancer: Do low-penetrance polymorphisms account for the increase in risk? Cancer Epidemiol Biomarkers Prev 16:1595-1600, 2007
- 29. Plna K, Hemminki K: Familial bladder cancer in the National Swedish Family Cancer Database. J Urol 166:2129-2133, 2001
- 30. Vasen HF, Stormorken A, Menko FH, et al: *MSH2* mutation carriers are at higher risk of cancer than MLH1 mutation carriers: A study of hereditary nonpolyposis colorectal cancer families. J Clin Oncol 19:4074-4080, 2001
- 31. Aarnio M, Sankila R, Pukkala E, et al: Cancer risk in mutation carriers of DNA-mismatch-repair genes. Int J Cancer 81:214-218, 1999
- Harper HL, McKenney JK, Heald B, et al: Upper tract urothelial carcinomas: Frequency of association with mismatch repair protein loss and lynch syndrome. Mod Pathol 30:146-156, 2017
- Audenet F, Isharwal S, Cha EK, et al: Clonal relatedness and mutational differences between upper tract and bladder urothelial carcinoma. Clin Cancer Res 25: 967-976, 2018
- 34. Kwon JS, Scott JL, Gilks CB, et al: Testing women with endometrial cancer to detect Lynch syndrome. J Clin Oncol 29:2247-2252, 2011
- 35. Latham A, Srinivasan P, Kemel Y, et al: Microsatellite instability is associated with the presence of Lynch syndrome pan-cancer. J Clin Oncol 37:286-295, 2019
- 36. Yap KL, Kiyotani K, Tamura K, et al: Whole-exome sequencing of muscle-invasive bladder cancer identifies recurrent mutations of UNC5C and prognostic importance of DNA repair gene mutations on survival. Clin Cancer Res 20:6605-6617, 2014
- 37. Robertson AG, Kim J, Al-Ahmadie H, et al: Comprehensive molecular characterization of muscle-invasive bladder cancer. Cell 171:540-556.e25, 2017
- 38. Stern MC, Lin J, Figueroa JD, et al: Polymorphisms in DNA repair genes, smoking, and bladder cancer risk: Findings from the international consortium of bladder cancer. Cancer Res 69:6857-6864, 2009
- 39. Cancer Genome Atlas Research Network: Comprehensive molecular characterization of urothelial bladder carcinoma. Nature 507:315-322, 2014
- 40. Liu D, Plimack ER, Hoffman-Censits J, et al: Clinical validation of chemotherapy response biomarker *ERCC2* in muscle-invasive urothelial bladder carcinoma. JAMA Oncol 2:1094-1096, 2016
- 41. Teo MY, Seier K, Ostrovnaya I, et al: Alterations in DNA damage response and repair genes as potential marker of clinical benefit from PD-1/PD-L1 blockade in advanced urothelial cancers. J Clin Oncol 36:1685-1694, 2018
- 42. Le DT, Uram JN, Wang H, et al: PD-1 blockade in tumors with mismatch-repair deficiency. N Engl J Med 372:2509-2520, 2015
- 43. Scosyrev E, Noyes K, Feng C, et al: Sex and racial differences in bladder cancer presentation and mortality in the US. Cancer 115:68-74, 2009

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Cancer Susceptibility Mutations in Patients With Urothelial Malignancies

The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated unless otherwise noted. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to www.asco.org/rwc or ascopubs.org/jco/site/ifc

Open Payments is a public database containing information reported by companies about payments made to US-licensed physicians (Open Payments).

Maria I. Carlo

Consulting or Advisory Role: Pfizer Other Relationship: Prostate Cancer Foundation, Robert Wood Johnson Foundation

Semanti Mukherjee Stock and Other Ownership Interests: Regeneron

Catharine K. Cipolla Stock and Other Ownership Interests: Roche Travel, Accommodations, Expenses: Flatiron Health

Hikmat Al-Ahmadie

Consulting or Advisory Role: Bristol-Myers Squibb, EMD Serono, AstraZeneca/ MedImmun

Karen A. Cadoo

Research Funding: AstraZeneca (Inst) Research Funding: Syndax (Inst)

Travel, Accommodations, Expenses: AstraZeneca

Min-Yuen Teo

Research Funding: Bristol-Myers Squibb, Clovis Oncology

Samuel A. Funt

Stock and Other Ownership Interests: Kite Pharma, Urogen Pharma (I), Hubble (I), Second Science, Allogene Therapeutics, Neogene Therapeutics, Kronos Bio (I). Vida Ventures (I)

Consulting or Advisory Role: AstraZeneca (Inst), MedImmune (Inst) Research Funding: Genentech (Inst), Roche (Inst), AstraZeneca (Inst), Decibel Therapeutics (Inst)

Travel, Accommodations, Expenses: Bristol-Myers Squibb, AstraZeneca, MedImmune

Jonathan A. Coleman Travel, Accommodations, Expenses: Digital Angiography Reading Center (I) Other Relationship: Steba Biotech

Bernard H. Bochner

Honoraria: Genentech, Roche Consulting or Advisory Role: Genentech, Roche, Olympus

Gopa Iver

Consulting or Advisory Role: Bayer, Janssen, Mirati Therapeutics Research Funding: Mirati Therapeutics (Inst), Novartis (Inst), Debiopharm Group (Inst), Bayer (Inst)

David B. Solit

Stock and Other Ownership Interests: Loxo

Consulting or Advisory Role: Pfizer, Loxo, Illumina, Intezyne Technologies, Vivideon Therapeutics, Lilly Oncology

Travel, Accommodations, Expenses: Merck KGaA

Zsofia K. Stadler

Consulting or Advisory Role: Allergan (I), Genentech (I), Roche (I), Regeneron (I), Optos (I), Adverum (I), Biomarin (I), Alimera Sciences (I), Novartis (I), Spark Therapeutics (I), Fortress Biotech (I), Regenxbio (I)

Living Zhang

Employment: Shanghai Genome Center (I)

Leadership: Shanghai Genome Center (I)

Stock and Other Ownership Interests: Shanghai Genome Center (I)

Honoraria: Future Technology Research, BGI, Illumina, Roche Diagnostics Asia Pacific

Travel, Accommodations, Expenses: Shanghai Genome Center (I), Roche Diagnostics Asia Pacific

Jonathan E. Rosenberg

Stock and Other Ownership Interests: Merck, Illumina Honoraria: UpToDate, Bristol-Myers Squibb, AstraZeneca, Medscape, Vindico, Peerview, Chugai Pharma, Research to Practice, Intellisphere, Clinical Care Options, Clinical Mind

Consulting or Advisory Role: Lilly, Merck, Agensys, Roche, Genentech, Sanofi, AstraZeneca, Medimmune, Bristol-Myers Squibb, EMD Serono, Seattle Genetics, Bayer, Inovio Pharmaceuticals, BioClin Therapeutics, QED Therapeutics, Adicet Bio, Sensei Biotherapeutics, Fortress Biotech, Pharmacyclics, Western Oncolytics, GlaxoSmithKline, Janssen Oncology, Astellas Pharma

Research Funding: Oncogenex (Inst), Agensys (Inst), Mirati Therapeutics (Inst), Novartis (Inst), Viralytics (Inst), Genentech (Inst), Roche (Inst), Incyte (Inst), Seattle Genetics (Inst), Bayer (Inst), AstraZeneca (Inst), QED Therapeutics (Inst), Astellas Pharma (Inst)

Patents, Royalties, Other Intellectual Property: Predictor of platinum sensitivity (Inst)

Travel, Accommodations, Expenses: Genentech, Roche, Bristol-Myers Squibb Barry S. Taylor

Consulting or Advisory Role: Boehringer Ingelheim Research Funding: Genentech

Mark E. Robson

Honoraria: AstraZeneca Consulting or Advisory Role: McKesson, AstraZeneca Research Funding: AstraZeneca (Inst), Myriad Genetics (Inst), InVitae (Inst), AbbVie (Inst), Tesaro (Inst), Medivation (Inst) Travel, Accommodations, Expenses: AstraZeneca, Pfizer

Other Relationship: Research to Practice, Clinical Care Options, Physician Education Resource

Uncompensated Relationships: Merck, Pfizer, Daiichi Sankyo Open Payments Link: https://openpaymentsdata.cms.gov/physician/612669/

summary

Michael F. Berger Consulting or Advisory Role: Roche Research Funding: Grail

Joseph Viiai

Patents, Royalties, Other Intellectual Property: Title: Diagnosis & treatment of ERCC3-mutant cancer; Inventors: Vijai Joseph, Sabine Topka, Kenneth Offit; US National Stage Patent Application No.: 16/493,214; Filing Date: September 11, 2019 (Inst)

Dean F. Bajorin

Honoraria: Merck Sharp & Dohme

Consulting or Advisory Role: Bristol-Myers Squibb, Novartis, Roche, Genentech, Merck, Lilly, Fidia Farmaceutici, Urogen Pharma, Pfizer, EMD Serono

Research Funding: Novartis (Inst), Genentech (Inst), Roche (Inst), Merck (Inst), Bristol-Myers Squibb (Inst), AstraZeneca (Inst), Astellas Pharma (Inst), Seattle Genetics (Inst), Astellas (Inst)

Travel, Accommodations, Expenses: Roche, Genentech, Merck, Bristol-Myers Squibb, Lilly, Urogen Pharma

No other potential conflicts of interest were reported.



FIG A1. Response to chemotherapy and immunotherapy in two patients with Lynch syndrome. (A) 18-Fluorodeoxyglucose positron emission tomography/computer tomography (PET/CT) image reveals a left para-aortic lymph node measuring 1.9 cm in largest dimension (red arrow) before initiation of paclitaxel. (B) Post-treatment CT after six infusions of paclitaxel shows enlargement of lymph node to 2.4 cm. (C) Post-treatment CT after six infusions of atezolizumab shows reduction in lymph node size to 0.9 cm, considered complete response by RECIST v 1.1 criteria. (D) CT image in another patient reveals right and left external iliac lymph nodes measuring 2.8 cm and 2.3 cm in largest dimensions, respectively (red arrows), before initiation of platinum-based chemotherapy. (E) Post-treatment CT after three cycles of platinum-based therapy shows right lymph node now to 2.7 cm and left to 3.2 cm. Treatment with chemotherapy was stopped, and the patient proceeded to treatment with a programmed death ligand 1 (PDL-1) inhibitor. (F) Post-treatment CT after 24 infusions of PDL-1 inhibitor shows disappearance of right lymph node and reduction of left lymph node to 0.6 cm. After 26 months on immune therapy, the patient proceeded to radical cystectomy with lymph node resection. Pathology review showed a complete pathologic response without residual carcinoma.

 TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With
 TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With

 Germline Mutations
 Germline Mutations (continued)

Gene	Syndrome
ABL1	
ACVR1	
AKT1	
AKT2	
AKT3	
ALK	Familial neuroblastoma
ALOX12B	
AMER1	
ANKRD11	
APC	Familial adenomatous polyposis
AR	
ARAF	
ARID1A	
ARID1B	
ARID2	
ARID5B	
ASXL1	
ASXL2	
ATM	Ataxia-telangiectasia; ATM-related cancer risk
ATR	
ATRX	
AURKA	
AURKB	
AXIN1	
AXIN2	
AXL	
B2M	
BABAM1	
BAP1	Mesothelioma, uveal melanoma, RCC
BARD1	Hereditary breast and ovarian cancer syndrome
BBC3	
BCL10	
BCL2	
BCL2L11	
BCL2L1	
BCL6	
BCOR	
BIRC3	
BLM	Bloom syndrome
BMPR1A	Juvenile polyposis syndrome
BRAF	
BRCA1	Hereditary breast and ovarian cancer syndrome
	(continued in next column)

Gene	Syndrome
BRCA2	Hereditary breast and ovarian cancer syndrome; Fanconi anemia
BRD4	
BRIP1	BRIP1-related cancer; Fanconi anemia
BTK	
KNSTRN	
CALR	
CARD11	
CARM1	
CASP8	
CBFB	
CBL	
CCND1	
CCND2	
CCND3	
CCNE1	
CD274	
CD276	
CD79A	
CD79B	
CDC42	
CDC73	
CDH1	Hereditary diffuse gastric cancer
CDK12	
CDK4	Familial cutaneous melanoma
CDK6	
CDK8	
CDKN1A	
CDKN1B	
CDKN2A	Familial cutaneous melanoma
CDKN2B	
CDKN2C	
CEBPA	
CENPA	
CHEK1	
CHEK2	CHEK2-related cancer
CIC	
CREBBP	
CRKL	
CRLF2	
CSDE1	
CSF1R	
CSF3R	
	(continued on following page)

TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With Germline Mutations (continued)

Germline Mutations (contin . ما

Gene	Syndrome
CTCF	
CTLA4	
CTNNB1	
CUL3	
CXCR4	
CYLD	
CYSLTR2	
DAXX	
DCUN1D1	
DDR2	
DICER1	DICER1-related disorders
DIS3	
DNAJB1	
DNMT1	
DNMT3A	
DNMT3B	
DOT1L	
DROSHA	
DUSP4	
E2F3	
EED	
EGFL7	
EGFR	Familial lung cancer
EIF1AX	
AGO2	
EIF4A2	
EIF4E	
ELF3	
EP300	
EPAS1	
EPCAM	Lynch syndrome
EPHA3	
EPHA5	
EPHA7	
EPHB1	
ERBB2	
ERBB3	
ERBB4	
ERCC2	Xeroderma pigmentosum
ERCC3	Xeroderma pigmentosum/ Hereditary breast cancer syndrome
ERCC4	
ERCC5	
	(continued in next column)

Gene	Syndrome
ERF	
ERG	
ERRFI1	
ESR1	
ETV1	
ETV6	
EZH1	
EZH2	
FAM175A	Hereditary breast cancer syndrome
FAM46C	
FAM58A	
FANCA	
FANCC	
FAT1	
FBXW7	
FGF19	
FGF3	
FGF4	
FGFR1	
FGFR2	
FGFR3	
FGFR4	
FH	Hereditary leiomyomatosis and renal cell cancer
FLCN	Birt-Hogg-Dubé syndrome
FLT1	
FLT3	
FLT3 FLT4	
FLT3 FLT4 FOXA1	
FLT3 FLT4 FOXA1 FOXL2	
FLT3 FLT4 FOXA1 FOXL2 FOXO1	
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1	
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1	
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN	
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN GATA1	
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN GATA1 GATA2	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN GATA1 GATA3	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN GATA1 GATA2 GATA3 GLI1	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOX01 FOXP1 FUBP1 FYN GATA1 GATA2 GATA3 GLI1 GNA11	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FVN GATA1 GATA2 GATA3 GLI1 GNA11 GNAQ	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOX01 FOXP1 FUBP1 FYN GATA1 GATA2 GATA3 GLI1 GNA11 GNAQ GNAS	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN GATA1 GATA3 GLI1 GNAQ GNAS GPS2	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN GATA1 GATA2 GATA3 GLI1 GNAQ GPS2 GRIN2A	Familial MDS-AML
FLT3 FLT4 FOXA1 FOXL2 FOXO1 FOXP1 FUBP1 FYN GATA1 GATA2 GATA3 GLI1 GNAQ GNAS GPS2 GRIN2A GSK3B	Familial MDS-AML

Journal of Clinical Oncology

TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With Germline Mutations (continued)

Gene	Syndrome
H3F3A	
H3F3B	
H3F3C	
HGF	
HIST1H1C	
HIST1H2BD	
HIST1H3A	
HIST1H3B	
HIST1H3C	
HIST1H3D	
HIST1H3E	
HIST1H3F	
HIST1H3G	
HIST1H3H	
HIST1H3I	
HIST1H3J	
HIST2H3C	
HIST2H3D	
HIST3H3	
HLA-A	
HLA-B	
HNF1A	
HOXB13	
HRAS	Costello syndrome
ICOSLG	
ID3	
IDH1	
IDH2	
IFNGR1	
IGF1	
IGF1R	
IGF2	
IKBKE	
IKZF1	
IL10	
IL7R	
INHA	
INHBA	
INPP4A	
INPP4B	
INPPL1	
INSR	
IRF4	
	(continued in next column)

Germline Mutations (continued)

Gene	Syndrome
IRS1	
IRS2	
JAK1	
JAK2	Familial thrombocytosis
JAK3	
JUN	
KDM5A	
KDM5C	
KDM6A	
KDR	
KEAP1	
KIT	Hereditary gastrointestinal stromal tumors
KLF4	
KRAS	Noonan syndrome
LATS1	
LATS2	
LMO1	
LYN	
MALT1	
MAP2K1	
MAP2K2	
MAP2K4	
MAP3K13	
MAP3K14	
MAP3K1	
MAPK1	
МАРКЗ	
MAPKAP1	
MAX	Hereditary paraganglioma-pheochromocytoma syndromes
MCL1	
MDC1	
MDM2	
MDM4	
MED12	
MEF2B	
MEN1	Multiple endocrine neoplasia, type 1
MET	Hereditary papillary renal carcinoma
MGA	
MITF	Familial melanoma and renal cell carcinoma
MLH1	Lynch syndrome
MLL2	
MLL3	
	(continued on following page)

TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With Germline Mutations (continued)

Germline Mutations (continued)

Gene	Syndrome	Gene
MLL4		PAK1
MLL		PAK7
MPL		PALB2
MRE11A	Ataxia-telangiectasia-like disorder	PARK
	(recessive); breast cancer	PARP
MSH2	Lynch syndrome	PAX5
MSH3		PBRM
MSH6	Lynch syndrome	PDCD
MSI1		PDCD
MSI2		PDGFR
MST1		PDGF
MST1R		PDPK.
MTOR		PGR
МИТҮН	MUTYH-associated polyposis	РНОХ2
МҮС		
MYCL1		<i>РІКЗС</i>
MYCN		РІКЗС
MYD88		<i>РІКЗС</i>
MYOD1		РІКЗС
NBN	Nijmegen breakage syndrome; NBN-related cancer	<i>РІКЗС</i>
	risk	РІКЗС
NCOA3		PIK3R
NCOR1		PIK3R
NEGR1		PIK3R
NF1	Neurofibromatosis, type 1	PIM1
NF2	Neurofibromatosis, type 2	PLCG2
NFE2L2		PLK2
NFKBIA		PMAI
NKX2-1		PMS1
NKX3-1		PMS2
NOTCH1		PNRC
NOTCH2		POLD
<i>NOTCH3</i>		POLE
NOTCH4		PPAR
NPM1		PPM1
NRAS	Autoimmune lymphoproliferative syndrome	PPP2
NSD1		PPP4
NTHL1		PPP6
NTRK1		PRDM
NTRK2		PRDM
NTRK3		PREX
NUF2		PRKA
NUP93		PRKC
	(continued in next column)	TANCI

Gene	Syndrome
PAK1	
PAK7	
PALB2	PALB2-related cancer; Fanconi anemia
PARK2	
PARP1	
PAX5	B cell precursor acute lymphoblastic leukemia
PBRM1	
PDCD1	
PDCD1LG2	
PDGFRA	Hereditary gastrointestinal stromal tumors
PDGFRB	
PDPK1	
PGR	
РНОХ2В	Familial neuroblastoma; congenital central hypoventilation syndrome
PIK3C2G	
PIK3C3	
РІКЗСА	
РІКЗСВ	
PIK3CD	
PIK3CG	
PIK3R1	
PIK3R2	
PIK3R3	
PIM1	
PLCG2	
PLK2	
PMAIP1	
PMS1	
PMS2	Lynch syndrome
PNRC1	
POLD1	
POLE	Colorectal cancer and endometrial cancer
PPARG	
PPM1D	
PPP2R1A	
PPP4R2	
PPP6C	
PRDM14	
PRDM1	
PREX2	
PRKAR1A	
PRKCI	
	(continued on following page)

TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With

 Germline Mutations (continued)

Gene	Syndrome
PRKD1	
PTCH1	Nevoid basal cell carcinoma syndrome
PTEN	PTEN hamartoma tumor syndrome
PTP4A1	
PTPN11	
PTPRD	
PTPRS	
PTPRT	
RAB35	
RAC1	
RAC2	
RAD21	
RAD50	Nijmegen breakage syndrome-like disorder
RAD51	Hereditary breast cancer
RAD51B	Hereditary breast cancer
RAD51C	RAD51C-related cancer; Fanconi anemia
RAD51D	Hereditary ovarian cancer
RAD52	
RAD54L	
RAF1	
RARA	
RASA1	
RB1	Retinoblastoma
RBM10	
RECQL4	Rothmund-Thomson syndrome
RECQL	
REL	
RET	Multiple endocrine neoplasia, type 2
RFWD2	
RHEB	
RHOA	
RICTOR	
RIT1	
RNF43	
ROS1	
RPS6KA4	
RPS6KB2	
RPTOR	
RRAGC	
RRAS2	
RRAS	
RTEL1	
	(continued in next column)

TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With Germline Mutations (continued) Gene Syndrome RUNX1 Familial platelet disorder with predisposition to acute myelogenous leukemia RXRA RYBP SDHA Hereditary paraganglioma-pheochromocytoma syndromes SDHAF2 Hereditary paraganglioma-pheochromocytoma syndromes SDHB Hereditary paraganglioma-pheochromocytoma syndromes SDHC Hereditary paraganglioma-pheochromocytoma syndromes SDHD Hereditary paraganglioma-pheochromocytoma (PGL/PCC) syndromes SESN1 SESN2 SESN3 SETD2 SETD8 SF3B1 SH2B3 SH2D1A SHOC2 SHQ1 SLX4 SMAD2 SMAD3 Thoracic aortic aneurysms and aortic dissections SMAD4 Juvenile polyposis syndrome SMARCA4 Rhabdoid tumor predisposition syndrome type 2 SMARCB1 Rhabdoid tumor predisposition syndrome type 1 SMARCD1 SMO SMYD3 SOCS1 SOS1 SOX17 SOX2 SOX9 SPEN SPOP SPRED1 SRC SRSF2 STAG2 (continued on following page)

Germline Mutations (continued)

Gene	Syndrome
STAT3	
STAT5A	
STAT5B	
STK11	Peutz-Jeghers syndrome
STK19	
STK40	
SUFU	Medulloblastoma
SUZ12	
SYK	
TAP1	
TAP2	
ТВХЗ	
TCEB1	
TCF3	
TCF7L2	
TEK	
TERT	Familial pulmonary fibrosis; dyskeratosis congenita
TET1	
TET2	
TGFBR1	Thoracic aortic aneurysms and aortic dissections
TGFBR2	Thoracic aortic aneurysms and aortic dissections
TMEM127	Familial pheochromocytoma syndrome
TMPRSS2	
TNFAIP3	
TNFRSF14	
TOP1	
TP53	Li-Fraumeni syndrome
TP53BP1	
TP63	
TRAF2	
TRAF7	
TSC1	Tuberous sclerosis complex
TSC2	Tuberous sclerosis complex
TSHR	
U2AF1	
UPF1	
VEGFA	
VHL	Von Hippel-Lindau syndrome; familial erythrocytosis, type 2
VTCN1	
WHSC1	
WHSC1L1	
	(continued in next column)

TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With TABLE A1. Genes on MSK-IMPACT and Syndromes Associated With Germline Mutations (continued) ~ .

Gene	Syndrome
WT1	Wilms tumor-aniridia-genital anomalies-retardation syndrome, Denys-Drash syndrome, Frasier syndrome, and isolated Wilms tumor
WWTR1	
XIAP	
XPO1	
XRCC2	
YAP1	
YES1	
ZFHX3	
ZRSR2	

NOTE. Genes in bold were included for germline analysis.

Abbreviations: MDS-AML, myelodysplastic syndrome/acute myeloid leukemia; RCC, renal cell carcinoma.

Journal of Clinical Oncology

MMR	NER	HR	FA	Checkpoint	Other
MLH1	ERCC2	BRCA1	BRCA2	ATM	POLE
MSH2	ERCC3	MRE11A	BRIP1	ATR	MUTYH
MSH6	ERCC4	NBN	FANCA	CHEK1	PARP1
PMS1	ERCC5	RAD50	FANCC	CHEK2	RECQL4
PMS2	—	RAD51	PALB2	MDC1	—
_	_	RAD51B	RAD51C	—	—
—		RAD51D	BLM	_	
_	_	RAD52	_	_	_
	_	RAD54L	_	_	_

TABLE A2. DDR Gene Panel DDR Pathway

Abbreviations: DDR, DNA-damage repair; HR, homologous recombination; FA, Fanconi anemia; MMR, mismatch repair; NER, nucleotide excision repair.

TABLE A3. Ashkenazi Jewish and European Founder Mutations

BRCA1 c.68_69delAG (p.Glu23Valfs*17)
BRCA1 c.5266dupC (p.Gln1756Profs*74)
BRCA2 c.5946delT (p.Ser1982Argfs*22)
CHEK2 c.1100delC (p.Thr367Metfs*15)
CHEK2 c.1283C>T (p.Ser428Phe)
APC c.3920T>A (p.lle1307Lys)
MUTYH c.1187G>A (p.Gly396Asp)
MUTYH c. 536A>G (p.Tyr179Cys)
<i>ERCC3</i> c.325C>T (p.Arg109X)

Study ID	Gene	Variant	Protein	Penetrance	Zygosity	Site	DDR Gene	Cohort
A1	APC	c.3920T>A	p.lle1307Lys	Low	No LOH	Bladder	No	Anonymized
A2	APC	c.3920T>A	p.lle1307Lys	Low	LOH	Bladder	No	Anonymized
A3	APC	c.3920T>A	p.lle1307Lys	Low	LOH	Bladder	No	Anonymized
A4	APC	c.3920T>A	p.lle1307Lys	Low	LOH	Bladder	No	Anonymized
A5	APC	c.3920T>A	p.lle1307Lys	Low	No LOH	Bladder	No	Anonymized
B1	APC	c.3920T>A	p.lle1307Lys	Low	LOH	Bladder	No	Identified
B164	APC	c.3920T>A	p.lle1307Lys	Low	LOH	Bladder	No	Identified
B23	APC	c.3920T>A	p.lle1307Lys	Low	LOH	Bladder	No	Identified
A7	ATM	c.5932G>T	p.Glu1978Ter	Moderate	No LOH	Bladder	Yes	Anonymized
A8	ATM	c.7638_7646delTAGAATTTC	p.Arg2547_Ser2549del	Moderate	LOH	Bladder	Yes	Anonymized
A9	BAP1	c.1203T>G	p.Tyr401Ter	High	No LOH	Bladder	No	Anonymized
A10	BARD1	c.1652C>G	p.Ser551Ter	Moderate	No LOH	Bladder	Yes	Anonymized
A6	APC	c.3920T>A	p.lle1307Lys	Low	No LOH	Upper tract	No	Anonymized
A11	BRCA1	c.5319dupC	p.Asn1774GInfsTer56	High	No LOH	Bladder	Yes	Anonymized
A21	CHEK2	c.444+1G>A		Moderate	No LOH	Upper tract	Yes	Anonymized
A12	BRCA1	c.116G>A	p.Cys39Tyr	High	No LOH	Bladder	Yes	Anonymized
A13	BRCA1	c.68_69delAG	p.Glu23ValfsTer17	High	No LOH	Bladder	Yes	Anonymized
B13	BRCA1	c.1687C>T	p.Gln563*	High	No LOH	Bladder	Yes	Identified
B148	BRCA1	c.68_69delAG	p.Glu23Valfs*17	High	LOH	Bladder	Yes	Identified
B21	BRCA1	c.68_69delAG	p.Glu23Valfs*17	High	No LOH	Bladder	Yes	Identified
B22	BRCA1	c.5074G>C	p.ASp1692His	High	No LOH	Bladder	Yes	Identified
A14	BRCA2	c.5799_5802delCCAA	p.Asn1933LysfsTer29	High	No LOH	Bladder	Yes	Anonymized
A15	BRCA2	c.8537_8538delAG	p.Glu2846GlyfsTer22	High	LOH	Bladder	Yes	Anonymized
A16	BRCA2	c.1238delT	p.Leu413HisfsTer17	High	LOH	Bladder	Yes	Anonymized
A17	BRCA2	c.8869C>T	p.Gln2957Ter	High	No LOH	Bladder	Yes	Anonymized
A18	BRCA2	c.7878G>C	p.Trp2626Cys	High	No LOH	Bladder	Yes	Anonymized
A32	MSH2	c.1906G>C	p.Ala636Pro	High	Somatic	Upper tract	Yes	Anonymized
B23	BRCA2	c.5946delT	p.Ser1982Argfs*22	High	LOH	Bladder	Yes	Identified
B24	BRCA2	c.1796_1800delCTTAT	p.Ser599*	High	LOH	Bladder	Yes	Identified
B28	BRCA2	c.5946delT	p.Ser1982Argfs*22	High	LOH	Bladder	Yes	Identified
A19	CHEK2	c.1100delC	p.Thr367MetfsTer15	Moderate	No LOH	Bladder	Yes	Anonymized
A20	CHEK2	c.1283C>T	p.Ser428Phe	Moderate	No LOH	Bladder	Yes	Anonymized
B21	CHEK2	c.1283C>T	p.Ser428Phe	Moderate	No LOH	Bladder	Yes	Identified
B51	CHEK2	c.444+1G>A		Moderate	No LOH	Bladder	Yes	Identified
B57	CHEK2	c.1283C>T	p.Ser428Phe	Moderate	No LOH	Bladder	Yes	Identified
A40	PALB2	c.940C>T	p.Gln314Ter	High	No LOH	Upper tract	Yes	Anonymized
B109	APC	c.3920T>A	p.lle1307Lys	Low	No LOH	Upper tract	No	Identified
B180	BLM	c.2207_2212delinsTAGATTC	p.Tyr736LeufsTer5	Recessive	No LOH	Bladder	Yes	Identified
B47	CHEK2	c.470T>C	p.lle157Thr	Uncertain	LOH	Upper tract	Yes	Identified
A11	ERCC2	c.1847G>C	p.Arg616Pro	Recessive	LOH	Bladder	Yes	Anonymized
A25	ERCC3	c.325C>T	p.Arg109Ter	Moderate	No LOH	Bladder	Yes	Anonymized
A26	ERCC3	c.325C>T	p.Arg109Ter	Moderate	No LOH	Bladder	Yes	Anonymized
A22	ERCC2	c.2150C>G	p.Ala717Gly	Recessive	Somatic	Bladder	Yes	Anonymized
			(continued on followin	ig page)				

TABLE A4. Detail on Pathogenic/Likely Pathogenic Germline Variants

TABLE A4.	Detail on	Pathogenic/Likelv	Pathogenic	Germline	Variants	(continued)
	Bottani oni	i autobotho, Entoly		G 011111110	• 411141110	(0011011000)

Study ID	Gene	Variant	Protein	Penetrance	Zygosity	Site	DDR Gene	Cohort
A6	MSH2	c.1906G>C	p.Ala636Pro	High	Somatic	Upper tract	Yes	Anonymized
A27	ERCC3	c.325C>T	p.Arg109Ter	Moderate	No LOH	Bladder	Yes	Anonymized
B20	ERCC3	c.325C>T	p.Arg109Ter	Moderate	No LOH	Bladder	Yes	Identified
A23	ERCC2	c.1847G>C	p.Arg616Pro	Recessive	LOH	Bladder	Yes	Anonymized
A24	ERCC3	c.576_583delCGTGATCC	p.Val193ArgfsTer8	Recessive	LOH	Bladder	Yes	Anonymized
A28	FH	c.1431_1433dupAAA	p.Lys477dup	Recessive	No LOH	Bladder	No	Anonymized
B101	MSH2	c.1906G>C	p.Ala636Pro	High	Somatic	Upper tract	Yes	Identified
A29	FH	c.1431_1433dupAAA	p.Lys477dup	Recessive	LOH	Bladder	No	Anonymized
B109	MSH6	c.3261dupC	p.Phe1088Leufs*5	High	No LOH	Upper tract	Yes	Identified
B111	MSH6	c.3463C>T	p.Gln1155*	High	Somatic	Upper tract	Yes	Identified
B161	MITF	c.952G>A	p.Glu318Lys	Moderate	No LOH	Bladder	No	Identified
A30	MRE11A	c.1222dupA	p.Thr408AsnfsTer49	Moderate	No LOH	Bladder	Yes	Anonymized
A31	MSH2	c.1255C>T	p.Gln419Ter	High	Somatic	Bladder	Yes	Anonymized
B100	MSH2	c.1216C>T	p.R406*	High	LOH	Bladder	Yes	Identified
B70	FH	c.1431_1433dupAAA	p.Lys477dup	Recessive	No LOH	Bladder	No	Identified
B14	BRCA1	c.68_69delAG	p.Glu23Valfs*17	High	CNLOH	Upper tract	Yes	Identified
B61	FH	c.1431_1433dupAAA	p.Lys477dup	Recessive	No LOH	Upper tract	No	Identified
A33	MUTYH	c. 536A>G	p.Tyr179Cys	Low	No LOH	Bladder	Yes	Anonymized
A34	MUTYH	c.1187G>A	p.Gly396Asp	Low	No LOH	Bladder	Yes	Anonymized
A35	MUTYH	c. 536A>G	p.Tyr179Cys	Low	No LOH	Bladder	Yes	Anonymized
B174	MLH1	c.1591delG	p.Val531Trpfs*4	High	LOH	Upper tract	Yes	Identified
A36	MUTYH	c.1187G>A	p.Gly396Asp	Low	No LOH	Bladder	Yes	Anonymized
A38	NBN	c.2140C>T	p.Arg714Ter	Moderate	No LOH	Bladder	Yes	Anonymized
A39	NBN	c.657_661delACAAA	p.Lys219AsnfsTer16	Moderate	No LOH	Bladder	Yes	Anonymized
B119	NBN	c.2T>C	p.Met1Thr	Moderate	No LOH	Bladder	Yes	Identified
A41	RAD50	c.326_329delCAGA	p.Thr109AsnfsTer20	Moderate	No LOH	Bladder	Yes	Anonymized
A42	RAD50	c.1270_1271delCT	p.Leu424GlufsTer7	Moderate	No LOH	Bladder	Yes	Anonymized
A43	RAD50	c.2467C>T	p.Arg823Ter	Moderate	No LOH	Bladder	Yes	Anonymized
B139	RAD51C	Exons 1-3 deletion		Moderate	No LOH	Bladder	Yes	Identified
A37	MUTYH	c.1187G>A	p.Gly396Asp	Low	No LOH	Bladder	Yes	Anonymized
B43	BRIP1	c.918+1G>A		Moderate	LOH	Upper tract	Yes	Identified
B118	MUTYH	c.536A>G	p.Tyr179Cys	Low	No LOH	Bladder	Yes	Identified
B5	ATM	c.748C>T	p.Arg250*	Moderate	CNLOH	Upper tract	Yes	Identified
A45	SDHA	c.245_252delAGGCAGGG	p.Glu82ValfsTer2	High	LOH	Bladder	No	Anonymized
B159	SDHA	c.1A>G	p.Met1Val	High	No LOH	Bladder	No	Identified
A44	RECQL4	c.2464-1G>C		Recessive	LOH	Bladder	Yes	Anonymized
B127	TP53	c.374C>T	p.Thr125Met	High	Somatic	Bladder	No	Identified
B71	MLH1	c.790+2T>C		High	LOH	Upper tract	Yes	Identified
B73	MSH2	c.1046C>G	p.Pro349Arg	High	Somatic	Upper tract	Yes	Identified
B79	MSH2	c.942+3A>T		High	Hom del	Upper tract	Yes	Identified
B85	MSH2	c.1784T>G	p.L595R	High	No LOH	Upper tract	Yes	Identified
A46	APC	c.3920T>A	p.lle1307Lys	Low	LOH	Upper tract	No	Anonymized
A46	BRCA2	c.5722_5723delCT	p.Leu1908ArgfsTer2	High	LOH	Upper tract	Yes	

Abbreviations: CNLOH, copy neutral loss of heterozygosity; DDR, DNA-damage repair; Hom del, homologous deletion; LOH, loss of heterozygosity; mt, mutation.

TABLE A5. Germline Mutations Identified by Sequencing and Clinical Criteria in

 Identified Cohort

Genetic Mutation	No. Identified by Sequencing	No. (%) Identified by Clinical Criteria
All high- or moderate-penetrance mutations	26	14 (53)
MSH2/MLH1/MSH6	9	6 (75)
BRCA1/BRCA2	8	7 (88)
CHEK2	2	1 (33)
RAD51C	1	0
BRIP1	1	0
SDHA	1	0
TP53	1	0
MITF	1	0
ATM	1	0
NBN	1	0
ERCC3	1	1*

*Patient met criteria for BRCA testing.