

Cystic Fibrosis

A Review

Thida Ong, MD; Bonnie W. Ramsey, MD

 CME at jamacmelookup.com

IMPORTANCE Cystic fibrosis, a genetic disorder defined by variants in the cystic fibrosis transmembrane conductance regulator (*CFTR*) gene, affects more than 30 000 individuals in the US and approximately 89 000 worldwide. Absent or decreased function of the CFTR protein is associated with multiorgan dysfunction and shortened life expectancy.

OBSERVATIONS CFTR is an anion channel in the apical membrane of epithelial cells. Loss of function leads to obstructed exocrine glands. Of people with cystic fibrosis in the US, approximately 85.5% have the gene variant F508del. Manifestations of cystic fibrosis in patients with the F508del gene variant begin in infancy with steatorrhea, poor weight gain, and respiratory symptoms (coughing, wheezing). As people with cystic fibrosis age, chronic respiratory bacterial infections cause loss of lung function and bronchiectasis. With the availability of universal newborn screening in multiple countries including the US, many people with cystic fibrosis are asymptomatic at diagnosis. With multidisciplinary care teams that included dietitians, respiratory therapists, and social workers, treatment of cystic fibrosis can slow disease progression. Median survival has improved from 36.3 years (95% CI, 35.1-37.9) in 2006 to 53.1 years (95% CI, 51.6-54.7) in 2021. Pulmonary therapies for patients with cystic fibrosis consist of mucolytics (eg, dornase alfa), anti-inflammatories (eg, azithromycin), and antibiotics (such as tobramycin delivered by a nebulizer). Four small molecular therapies, termed *CFTR modulators*, that facilitate CFTR production and/or function have received regulatory approval. Examples are ivacaftor and elexacaftor-tezacaftor-ivacaftor. For example, in patients with 1 F508del variant, the combination of ivacaftor, tezacaftor, and elexacaftor improved lung function from -0.2% in the placebo group to 13.6% (difference, 13.8%; 95% CI, 12.1%-15.4%) and decreased the annualized estimated rate of pulmonary exacerbations from 0.98 to 0.37 (rate ratio, 0.37; 95% CI, 0.25-0.55). Improved respiratory function and symptoms have lasted up to 144 weeks in postapproval observational studies. An additional 177 variants are eligible for treatment with the elexacaftor-tezacaftor-ivacaftor combination.

CONCLUSION Cystic fibrosis affects approximately 89 000 people worldwide and is associated with a spectrum of disease related to exocrine dysfunction, including chronic respiratory bacterial infections and reduced life expectancy. First-line pulmonary therapies consist of mucolytics, anti-inflammatories, and antibiotics, and approximately 90% of people with cystic fibrosis who are 2 years or older may benefit from a combination of ivacaftor, tezacaftor, and elexacaftor.

JAMA. 2023;329(21):1859-1871. doi:10.1001/jama.2023.8120

Author Affiliations: Division of Pulmonary and Sleep Medicine, Department of Pediatrics, University of Washington, Seattle Children's Hospital, Seattle.

Corresponding Author: Bonnie W. Ramsey, MD, Division of Pulmonary and Sleep Medicine, Department of Pediatrics, University of Washington, Seattle Children's Hospital, M/S OC7.720, PO Box 5371, Seattle, WA 98145 (bonnie.ramsey@seattlechildrens.org).

Section Editor: Mary McGrae McDermott, MD, Deputy Editor.

Cystic fibrosis is an autosomal recessive, genetic disease characterized by reduced or absent function of the cystic fibrosis transmembrane conductance regulator (CFTR) protein that affects more than 30 000 individuals in the US and approximately 89 000 in registries worldwide.^{1,2} More than 2000 sequence variants of the *CFTR* gene (OMIM 602421) have been identified. Of these, approximately 700 have been shown to cause disease.³

The clinical features of cystic fibrosis result from reduced or absent function of the CFTR protein, a regulated anion channel

located in the apical membrane of epithelia in multiple organs, including the lungs, liver, gastrointestinal tract, and pancreas.^{4,5} The clinical manifestations of CFTR dysfunction and resultant organ damage include pancreatic insufficiency with malnutrition, biliary cirrhosis, absence of the vas deferens resulting in azoospermia, chronic sinusitis, and chronic endobronchial bacterial infections associated with obstructive airway disease.⁶ Identifying the structure and function of CFTR has facilitated development of small molecule CFTR modulator drugs that improve the health of more than 90% of people with cystic fibrosis.⁷ This review summarizes

Box. Commonly Asked Questions on the Management of Cystic Fibrosis

How have the new medicines, termed *CFTR* modulators, affected the lives of people with cystic fibrosis?

Modulator combination therapies such as elexacaftor-tezacaftor-ivacaftor, which received initial regulatory approval in 2019, have been highly effective leading to a marked improvement in lung function as measured by forced expiratory volume in the first second, reductions in respiratory symptoms such as cough and sputum production, and hospitalizations. People with cystic fibrosis also experience weight gain and improvements in several measures of quality of life including reduction in school or work absenteeism.

Do all people with cystic fibrosis have access to highly effective modulators?

People with cystic fibrosis must carry at least 1 copy of a cystic fibrosis gene variant responsive to a modulator therapy such as elexacaftor-tezacaftor-ivacaftor to be eligible to receive the therapy. Approximately 90% of people with cystic fibrosis have the gene variant F508del, which is responsive to the elexacaftor-tezacaftor-ivacaftor combination, but there remain many people with cystic fibrosis who do not yet have access to elexacaftor-tezacaftor-ivacaftor, including children younger than 2 years, people with rare gene variants unresponsive to therapy, and people living in geographic regions without access to the approved drugs.

Are there research efforts to find therapies for all people with cystic fibrosis?

There are multiple research programs across the world trying to develop gene-agnostic therapies for which all people with cystic fibrosis will be eligible including gene therapy, gene editing, messenger RNA therapy, and alternative ion channels to bypass the CFTR protein. Most of these approaches are in preclinical stages.

current evidence regarding clinical manifestations and pulmonary treatments of cystic fibrosis (**Box**).

Methods

We searched PubMed for English-language studies of practical guidelines, meta-analyses, or clinical or randomized clinical trials of cystic fibrosis published from January 1, 2012, to December 31, 2022, and updated the search through March 31, 2023.

A total of 1002 articles were retrieved. A second literature search from January 1, 2002, to December 31, 2022, focused on CFTR modulators and resulted in 26 additional articles. Current practice guidelines were reviewed. We manually inspected reference lists of selected articles for other relevant sources. Articles of highest priority for inclusion were meta-analyses, randomized clinical trials, longitudinal studies with longer follow-up, and studies relevant to general medical practice. The 115 articles included 24 were randomized clinical trials; 6 were pediatric single-arm intervention trials; 7 were meta-analyses; 24 were reviews; 14 were longitudinal observational studies; 9 were cross-sectional studies; 15 were guidelines or consensus documents; 6 were basic science studies; 3 were registry studies; and 7 were other (1 autopsy case series, 1 economic analysis, 1 protocol, and 4 prescribing information).

Discussion and Observations**Epidemiology**

Worldwide, approximately 89 000 individuals are living with cystic fibrosis, including approximately 31 450 people in the US.^{1,2} The prevalence of cystic fibrosis is similar between the US (7.97 per 100 000) and European Union (7.37 per 100 000).⁸ Among people with cystic fibrosis in the US, approximately 3.5% identified as Black or African American, 91.4% as White, and 5.1% as other, which included people who identified as American Indian or Alaska Native, Asian, Native Hawaiian or other Pacific Islander, some other race, or 2 or more races. Among people with cystic fibrosis in the US, approximately 9.8% identified as Hispanic ethnicity and approximately 91.2% as non-Hispanic ethnicity.¹

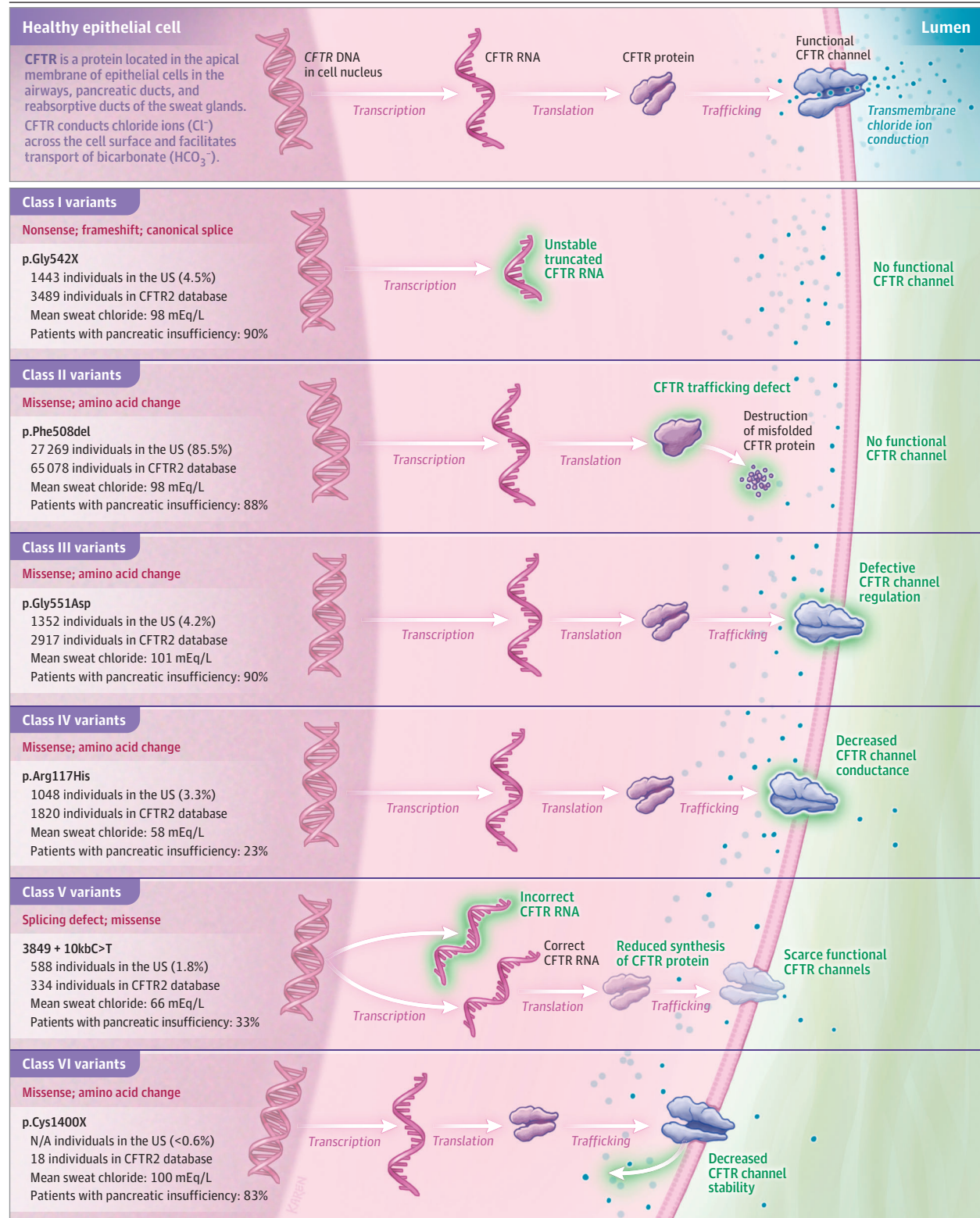
Approximately 85.5% of people in the US have the phenylalanine deleted at position 508 (p.Phe508del) gene variant, also known as F508del.¹ A meta-analysis described 24 to 54 *CFTR* gene variants in regions from South Asia, the Middle East, and East Asia, but populations of non-European ancestry are likely underestimated due to ascertainment bias.^{2,9} In populations from 10 countries in Latin America, F508del was the most frequent *CFTR* variant, ranging from 23% to 59%.¹⁰ Rare variants (<1% of individuals) in Latin American populations reflect diverse Native, African, and European heritages.¹⁰

Pathophysiology

Pathophysiological changes in cystic fibrosis are primarily due to loss of CFTR protein function and its essential role as an anion channel in apical epithelia. Loss of function of the CFTR protein alters hydration and pH concentration in exocrine ducts, leading to obstructed and dilated exocrine glands in multiple organs.¹¹ Reduced CFTR function in the sweat gland leads to increased salt losses and higher chloride concentrations in sweat.¹² The mucinous obstruction of pancreatic acini and ducts biliary ducts and glandular obstruction of the vas deferens and submucosal glands in the airways leads to organ destruction and fibrosis.¹³⁻¹⁵ The endobronchial space of airways in people with cystic fibrosis typically becomes infected initially with bacterial pathogens such as *Staphylococcus aureus* and *Haemophilus influenzae* and later with *Pseudomonas aeruginosa*.¹⁶ These infections are associated with a neutrophilic inflammatory response and persistent mucopurulent plugging that leads to bronchiectasis.¹⁷ With the availability of CFTR modulator therapies, the pathogenesis of clinical disease is changing, and early intervention may partially prevent development of multiorgan pathology. In utero administration of the CFTR modulator ivacaftor to ferret fetuses with the glycine at residue 551 replaced by the aspartic acid (p.Gly551Asp; legacy G551D) variant reduced meconium ileus and improved pancreatic exocrine function, growth, and survival.¹⁸

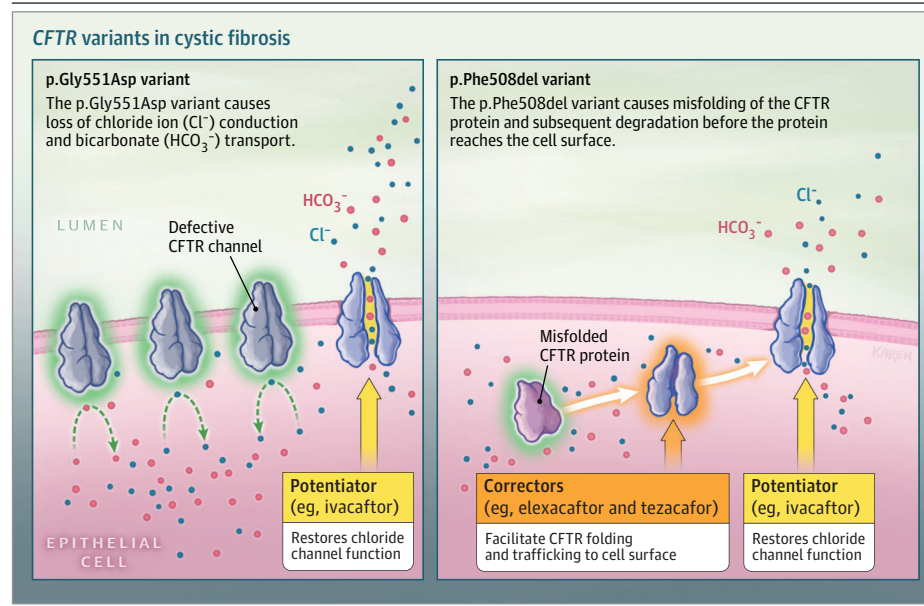
More than 700 disease-causing gene variants of *CFTR* have been identified.^{3,6} The most common are grouped into 6 classes by the processes through which they can cause CFTR dysfunction (**Figure 1**).^{19,20} Three classes (I, II, III) typically result in minimal or no CFTR and are often associated with the highest sweat-chloride values, severe lung disease, and pancreatic insufficiency whereas classes IV, V, and VI are associated with some residual protein function, may have lower sweat chloride, and milder disease. Although there are examples in which single variants affect multiple mechanisms, matching of cystic fibrosis variants with biological pathways

Figure 1. Cystic Fibrosis Transmembrane Conductance Regulator Variant Classes^{1,5,6,19}



Cystic fibrosis transmembrane conductance regulator (CFTR) variants can be generally classified in 6 mechanistic classes based on how they alter CFTR RNA transcription, protein trafficking, channel function, and stability.^{5,19} Reported prevalence, and clinical features (sweat chloride, pancreatic insufficiency) are

summarized for exemplar variants per class.^{1,6} The CFTR2 database provides information on all the CFTR variants and updates it as information becomes available.⁶ The figure is adapted from Boyle and De Boeck.⁵ N/A indicates number not available.

Figure 2. Cystic Fibrosis Transmembrane Conductance Regulator Modulator Therapy Functions²¹⁻²⁴

have aided in the development of CFTR modulator therapies.²¹ Modulator therapies that increase the quantity of CFTR protein at the cell surface are termed *correctors*, and those that augment channel function are termed *potentiators* (Figure 2).

Clinical Presentation

Because disease-causing variants of the *CFTR* gene result in a range of protein dysfunction, the clinical presentation and rate of disease progression are variable (Figure 1).^{5,21} More than 80% of people with cystic fibrosis and 2 severe gene variants have consequences of exocrine pancreatic insufficiency including protein and fat maldigestion, steatorrhea, and poor growth.²⁵ Both upper and lower airway disease begin in infancy with cough, increased respiratory rate, or wheezing or crackles on chest auscultation.²⁶ As patients become infected with pathogens such as *S aureus* and subsequently *P aeruginosa*, they frequently experience acute pulmonary exacerbations, characterized by cough, sputum production, and dyspnea, which require more frequent airway clearance treatments (Table 1) and often hospitalization.⁴⁴ Chronic endobronchial infections and inflammation lead to a decline in lung function, characterized by a decrease in the forced expiratory volume in the first second (FEV_1) of expiration and forced vital capacity (FVC) on spirometry. Most patients with cystic fibrosis develop an obstructive pattern on spirometry. Recurrent pulmonary infections cause bronchiectasis, a major cause of morbidity and mortality.⁷ In addition, patients with advanced cystic fibrosis may develop pulmonary hypertension, which is associated with decreased survival.⁴⁹ Adults in the US have been reported to have increased risk of comorbidities¹ including cystic fibrosis-related diabetes (29.2%),⁵⁰ liver disease with cirrhosis (4.1%),⁵¹ and osteoporosis (7.5%).⁵² People with cystic fibrosis who have at least 1 copy of a *CFTR* variant with residual function often have later onset of lung disease yet have comparable disease progression with those with minimally functional variants.⁵³

Of 563 infants diagnosed by newborn screening in the US in 2021, 88.3% were asymptomatic at the time of diagnosis.¹ Among

the 216 individuals diagnosed at ages older than 6 months, the most common presenting symptoms were acute or persistent respiratory abnormalities (50.2%) such as cough or wheeze, nasal polyps or sinus disease (15.5%), congenital bilateral absence of the vas deferens or infertility (9%), steatorrhea or abnormal stools (7.7%), failure to thrive (6.9%), and digital clubbing (2.6%).¹

Assessment and Diagnosis

Diagnostic criteria for cystic fibrosis consist of 1 or more organ-specific manifestations and elevated sweat chloride levels or genetic confirmation of 2 disease-causing variants in the *CFTR* gene. Most newborn screening methods include measurement of immunoreactive trypsinogen (IRT) from a bloodspot, followed by DNA testing for *CFTR* variants, but the thresholds that define IRT elevation and selection of *CFTR* variants can vary across the US, affecting the prevalence of positive screening results.^{54,55}

Sweat chloride testing is the main diagnostic test for cystic fibrosis with high sensitivity (99%) and specificity (93%) and has established guidelines for technical quality and accuracy at specialized cystic fibrosis centers.^{12,56} Elevated chloride concentration of collected sweat (≥ 60 mEq/L) is consistent with the diagnosis. Intermediate sweat chloride levels (30-59 mEq/L) require further biochemical, genetic testing, or nasal potential difference measurement and long-term follow-up at specialized centers because some patients may later be diagnosed with definite cystic fibrosis, ranging from 6% to 48% based on prospective and retrospective case series and registry studies.^{12,57-59}

Treatment

Long-term Therapies

For patients with cystic fibrosis, at least quarterly visits with a specialized, multidisciplinary team, including physicians, nurses, social workers, and dietitians, are recommended to monitor for disease progression and treat multiorgan manifestations.^{29,30,49,60,61} Annual screening for psychosocial health concerns is recommended in

Table 1. Long-term Pulmonary Therapies for Cystic Fibrosis

| Therapy | Mechanism of action | Indication | Mode of administration and frequency | Evidence summary |
|---|--|--|--|---|
| Mucociliary clearance | | | | |
| Airway clearance techniques such as chest physiotherapy and oscillating devices | Augmented mucociliary clearance of the lung and facilitate cough to remove mucus obstruction from airways; mechanical loosening of airway secretions | Mechanical exercise and devices to relieve retained airway secretions in conjunction with standard maintenance nebulized therapies | Maintenance typically 2/d and increases with pulmonary exacerbations ²⁷ | In a meta-analysis of 38 studies (n = 1114) of multiple oscillating devices for airway clearance techniques, there was no clear evidence of one mode superior to another ²⁸ |
| Dornase alfa | Reduced viscosity of airway secretions through cleaving of extracellular DNA in sputum | Recombinant DNase enzyme used in conjunction with maintenance airway clearance techniques | Recommended as part of maintenance therapy: 2.5-mg nebulized 1/d ^{29,30} | In a meta-analysis of 15 studies (n = 2447), dornase alfa improved lung function and reduced pulmonary exacerbations vs placebo ³¹ |
| Inhaled hypertonic saline | Not established. Proposed mechanisms include airway surface hydration through improved sputum rheological properties and antimicrobial properties | Concentrated saline solution inhaled used in conjunction with standard maintenance airway clearance therapies | Recommended as part of maintenance therapy ²⁹ 7%: 4 mL-nebulized 2/d; Bronchodilator pretreatment recommended to reduce symptoms of cough and wheeze associated with administration | Meta-analysis of 17 randomized controlled trials (n = 966) found evidence of reduced frequency of pulmonary exacerbations, but low evidence for improvement in FEV ₁ ³² In recent trials among young children, 7% hypertonic saline use improved lung clearance index, a measure of lung function ^{33,34} |
| Mannitol | Not directly established; proposed mechanisms are to act as a hyperosmolar agent to rehydrate the airway surface and improve sputum viscosity | Nebulized sugar alcohol as add-on maintenance therapy to manage patients who have passed a tolerance test Mannitol was approved in 2019 for age ≥18 y in the US | 400-mg inhaled 2/d | A meta-analysis of 6 studies (n = 784) reported that mannitol improved measures of lung function with moderate quality evidence ³⁵ Inhaled mannitol was associated with mean absolute improvement of lung function relative to placebo in adults (ppFEV ₁ difference, 1.21%; 95% CI, 0.07%-2.36%; P = .04) ³⁶ |
| Anti-inflammatory | | | | |
| Azithromycin | Proposed anti-inflammatory mechanisms include reducing IL-4 and IL-8, suppressing neutrophil activity, and decreasing production of tumor necrosis factor ³⁷ | Macrolide antibiotic as add-on maintenance therapy for patients who are chronically infected with <i>Pseudomonas aeruginosa</i> and consideration of use for those without <i>P aeruginosa</i> ^{29,30} | Limited data, but largest trials have used <40 kg: 250 mg 3/wk and ≥40 kg: 500 g 3/wk ³⁸ | A 2012 Cochrane review of 10 studies (n = 959) found azithromycin improved mean FEV ₁ by approximately 4% vs placebo for patients infected with <i>P aeruginosa</i> and reduced exacerbations ³⁸ Hospital days for exacerbations were reduced with azithromycin vs placebo among infants, 3-6 mo (mean difference, 6.3; 95% CI; -10.5 to -2.1) ³⁹ |
| High-dose ibuprofen | Not directly established; proposed mechanisms include reduced airway inflammation as an inhibitor of cyclooxygenase-1 and cyclooxygenase-2; blocks release of leukotriene B ₄ , a proinflammatory molecule that promotes neutrophil activity; inhibition of neutrophil chemotaxis and NF-κB-mediated inflammation ²⁹ | Patients aged 6-18 y to reduce annual decline in lung function; US national guidelines recommend use in children with attention to concentration levels, but insufficient evidence for use in adults ²⁹ | Dose adjusted for peak plasma concentrations of 50 to 100 μg/mL; approximately 20 to 30 mg/kg 2/d Maximum dose: 1600 mg 2/d Requirement for annual ibuprofen levels has reduced acceptance of therapy by patients and caregivers | A 2019 Cochrane review of 4 trials (n = 287) found ibuprofen treatment vs placebo reduced annual rates of lung function decline, particularly in children ⁴⁰ |
| Inhaled antibiotics for chronic <i>P aeruginosa</i> | | | | |
| Inhaled tobramycin | Active against most gram-negative bacilli through binding of bacterial ribosome and inhibiting protein synthesis ⁴¹ | Guideline-recommended inhaled maintenance therapy for patients with chronic <i>Pseudomonas aeruginosa</i> infection ²⁹ | 300-mg nebulized or 112-mg dry powder inhaler 2/d for 28 d alternating with 28 d off | A 2022 meta-analysis of inhaled antibiotics to treat <i>P aeruginosa</i> in included 18 trials (n = 3024); among the 12 included, tobramycin had the best available evidence of the antipseudomonal antibiotics reviewed and tobramycin use was associated with improved lung function and reduced exacerbations ⁴² |

(continued)

Table 1. Long-term Pulmonary Therapies for Cystic Fibrosis (continued)

| Therapy | Mechanism of action | Indication | Mode of administration and frequency | Evidence summary |
|--|---|--|---|---|
| Inhaled aztreonam lysine | Inhibits synthesis of bacterial cell walls; active against gram-negative bacteria and stable against β -lactamases ⁴³ | Inhaled maintenance therapy for patients with chronic <i>P aeruginosa</i> infection ²⁹ | 75-mg nebulized 3/d for 28 d alternating with 28 d off | A 2022 meta-analysis of inhaled antibiotics to treat <i>P aeruginosa</i> included 18 trials (n = 3024); Of 18 trials, 1 trial found moderate-quality evidence for aztreonam use for improved lung function (mean difference, FEV ₁ , -3.4%; 95% CI, -6.63% to -0.17%) with fewer hospitalizations vs inhaled tobramycin ⁴² |
| Pulmonary exacerbations | | | | |
| Management of pulmonary exacerbations, clinically presenting as recurrent episodes of clinical symptoms including increased cough, sputum production, dyspnea, decreased energy level and appetite, weight loss, and/or decreases in measures of spirometry ^{44,45} | Antibiotic therapy for 10- to 14-d courses associated with decreased bacterial density in sputum and improved lung function ⁴⁶ | Guideline-recommended increased frequency for airway clearance therapy sessions and antibiotic courses ⁴⁴ | Antibiotics based on respiratory microbiological cultures; increased frequency of airway clearance; hospitalization may be needed | A 2021 trial of adults (n = 982) treated for pulmonary exacerbation found lung function change from treatment initiation was noninferior at 10 d vs 14 d of therapy among early responders within 7 to 10 d of therapy (mean ppFEV ₁ change, 12.8% vs 13.4%; difference, -0.65%; 95% CI, -3.3% to 2.0%) and noninferior at 14 vs 21 d of therapy for later responders (difference, -0.10%; 95% CI, -1.3% to 1.1%) ⁴⁷ Lung function improvement was higher for those treated in the hospital vs at home (mean ppFEV ₁ change, 8.0%; 95% CI, 6.7% to 9.4% vs 5.0%; 95% CI, 3.5% to 6.5%) ⁴⁸ |

Abbreviations: NF- κ B, nuclear factor kappa-B; ppFEV₁, predicted percent of forced expiratory volume in the first second of expiration.

children aged 12 years or older.⁶² Monitoring for comorbidities includes annual oral glucose tolerance testing (≥ 10 years) for cystic fibrosis-related diabetes,⁵⁰ dual-energy x-ray absorptiometry scanning every 2 to 5 years (>8 years) for bone density, and colonoscopy every 5 years (≥ 40 years) for colorectal cancer.⁶³

Disease progression is measured by monitoring for trends in nutritional status (height, weight, body mass index [BMI], calculated as weight in kilograms divided by height in meters squared), lung health (spirometry, respiratory microbiology, chest imaging), and assessments for pulmonary exacerbations. Exacerbations manifest as an acute worsening of respiratory symptoms and lung function (percent predicted FEV₁ [ppFEV₁]) and usually require oral or intravenous antibiotic treatments specific for respiratory microbiology, increased airway clearance therapies (eg, high-frequency oscillatory percussive devices), and high-calorie, high-protein diets to limit permanent loss of lung function.^{44,47} In a randomized clinical trial of 982 participants with cystic fibrosis and pulmonary exacerbation defined by providers as necessitating intravenous antibiotic treatment, antibiotic therapy duration of 10 days was noninferior to 14 days, based on the outcome of lung function (change in ppFEV₁) among those who had improved lung function and symptoms within 7 to 10 days of treatment (mean ppFEV₁ change, 12.8% vs 13.4%; difference, -0.65%; 95% CI, -3.3% to 2.0%). In addition, those without improved lung function or symptoms within days 7 to 10 days, 21 days of intravenous antibiotics was not superior to 14 days⁴⁷ (mean ppFEV₁ change, 3.3% vs 3.4%; difference, -0.10%; 95% CI, -1.3% to 1.1%).⁴⁷ In subsequent analysis, lung function improvement was higher for those treated in the hospital (mean ppFEV₁ change, 8.0%;

95% CI, 6.7% to 9.4%) vs at home (mean ppFEV₁ change, 5.0%; 95% CI, 3.5% to 6.5%).⁴⁸

Long-term pharmacological pulmonary therapies such as mucolytics to thin secretions to facilitate clearance from the upper and lower airways (such as dornase alfa), airway surface liquid hydration (inhaled hypertonic saline, mannitol), and anti-inflammatory drugs (azithromycin, ibuprofen) have been based on phase 3 randomized, placebo-controlled clinical trials (Table 1). In a clinical trial of 968 patients with cystic fibrosis, dornase alfa compared with placebo increased the mean percent change in FEV₁ by 5.8% (SE, 0.7%) vs 0% (SE, 0.6%) and reduced the proportion of patients with 1 or more pulmonary exacerbations from 89 (27%) to 61 (19%).⁶⁴ In a clinical trial of 164 participants, 7% hypertonic saline compared with 0.9% saline reduced pulmonary exacerbations (mean exacerbations per participant, 0.39 [7% saline] vs 0.89 [0.9% saline]); difference, 0.5; 95% CI, 0.14-0.86; $P = .02$).⁶⁵ In a randomized clinical trial of 185 patients chronically infected with *P aeruginosa*, azithromycin, compared with placebo, significantly improved lung function from baseline (ppFEV₁, 4.4% vs -1.8%; mean difference, 6.2%; 95% CI, 2.6%-9.8%) at end of 168 days of treatment.⁶⁶

CFTR Modulator Therapies

CFTR modulator therapies act by 2 mechanisms to enhance CFTR function. Potentiators, like ivacaftor, increase the probability that the protein channel is open, so chloride or bicarbonate can flow more easily through the cell membrane (Figure 2). Correctors, like lumacaftor, tezacaftor, and elexacaftor, improve channel quantity at the cell surface by helping the protein fold properly, enabling

transport to the cell surface (Figure 2). Severe variants such as F508del need both potentiators and correctors to improve channel quantity and function (Figure 2). Four modulators are currently approved by US and European drug regulatory agencies, and eligibility for each treatment depends on the specific *CFTR* genetic variants present (Table 2). Ivacaftor is available as a monotherapy, and lumacaftor-ivacaftor, tezacaftor-ivacaftor, and elexacaftor-tezacaftor-ivacaftor are available as combination therapies (Table 2).

Ivacaftor (formerly VX-770) was the first *CFTR* modulator tested in randomized clinical trials of patients with cystic fibrosis in 2006. Ivacaftor was tested first for patients with cystic fibrosis who have a *G551D-CFTR* variant where the *CFTR* protein is transported to the cell membrane, but the *CFTR* channel does not open properly. In a randomized clinical trial of 161 patients with at least 1 copy of *G551D*, compared with placebo, patients at 24 weeks' follow-up had improved ppFEV₁ (10.1% vs -0.4%; mean difference, 10.5%; 95% CI, 8.5%-12.5%), a 55% reduction in pulmonary exacerbations (28 vs 44; rate ratio, 0.43; 95% CI, 0.27-0.68), and increased weight (3.1 kg vs 0.4 kg; mean difference, 2.7 kg; 95% CI, 1.3-4.1 kg).⁶⁸ Respiratory symptoms were scored on a 100-point scale on the respiratory domain of the Cystic Fibrosis Questionnaire revised (CFQ-R), for which higher numbers indicate a lower effect of these symptoms on quality of life (minimal clinically important difference, 4 points).⁸¹ Ivacaftor improved respiratory symptom scores by 8.6 points relative to placebo (5.9 vs -2.7; mean difference, 8.6; *P* < .001). Ivacaftor is approved for patients 4 months or older.

For people with 2 copies of the F508del variant, ivacaftor alone did not improve *CFTR* activity or demonstrate clinical efficacy.⁸² These patients required the combination of corrector and potentiator medications. Randomized, placebo-controlled clinical trials of first-generation correctors, lumacaftor or tezacaftor, in combination with ivacaftor demonstrated modest improvements in ppFEV₁ and reduction in pulmonary exacerbations in patients homozygous for the F508del variant.^{73,74,77,83,84} For example, in a clinical trial of 509 patients homozygous for the F508del variant, the tezacaftor-ivacaftor combination drug compared with placebo improved ppFEV₁ (3.4% vs -0.6%; mean difference, 4%; 95% CI, 3.1% to 4.8%) and reduced the pulmonary exacerbations annualized rate (0.64 vs 0.99 events per year; rate ratio, 0.65; 95% CI, 0.48 to 0.88) at 24 weeks' follow-up.⁷⁰ The tezacaftor-ivacaftor combination reduced sweat chloride (-9.9 vs 0.2 mEq/L; mean difference, -10.1 mEq/L; 95% CI, -11.4 to -8.8 mEq/L) for patients homozygous for the F508del variant, but had no effect in patients with 1 copy.^{53,77,85,86}

The combination of a second-generation corrector, elexacaftor (formerly VX-445) with the first-generation corrector tezacaftor (formerly VX-661) had an additive effect in stabilizing the nascent *CFTR* protein and facilitated increased expression of the mature *CFTR* protein channel at the cell surface.^{87,88} When the 2 correctors were combined with the potentiator, ivacaftor, in phase 3 randomized trials, this triple combination was effective and had similar clinical responses for people with cystic fibrosis who were either homozygous for the F508del variant or who had 1 copy of the F508del variant and 1 copy of a minimal function variant on the second allele (Table 2).^{22,23} In a randomized clinical trial of 107 patients who were homozygous for the F508del variant, the elexacaftor-tezacaftor-ivacaftor combination compared with tezacaftor-ivacaftor alone increased ppFEV₁ (10.4% vs 0.4%; difference, 10.0%; 95% CI, 7.4% to 12.6%), decreased sweat chloride concentration (-43.4 vs 1.7 mEq/L;

difference, -45.1; 95% CI, -50.1 to -40.1 mEq/L), and improved respiratory symptom scores above the 4-point minimally important clinical difference for CFQ-R (16 vs -1.4; difference, 17.4; 95% CI, 11.8 to 23) at 4 weeks' follow-up.²²

In a randomized clinical trial²³ of 403 patients heterozygous for the F508del variant and a minimal function variant, elexacaftor-tezacaftor-ivacaftor compared with placebo improved ppFEV₁ (13.6% vs -0.2%; mean difference, 13.8%; 95% CI, 12.1% to 15.4%) at 4 weeks and through 24 weeks (13.9% vs -0.4%; mean difference, 14.3%; 95% CI, 12.7% to 15.8%).²³ The elexacaftor-tezacaftor-ivacaftor combination decreased the pulmonary exacerbations annualized rate (0.37 vs 0.98; rate ratio, 0.37; 95% CI, 0.25 to 0.55), increased absolute change in body mass index from baseline (1.13 vs 0.09; difference, 1.04; 95% CI, 0.85 to 1.23), improved respiratory symptom scores by CFQ-R (17.5 vs -2.7; difference, 20.2 points; 95% CI, 17.5 to 23.0) and decreased sweat chloride concentration (-42.2 vs -0.4 mEq/L; difference, -41.8 mEq/L; 95% CI, -44.4 to -39.3 mEq/L) at 24 weeks' follow-up.^{22,23} Preliminary open-label observational studies involving people taking elexacaftor-tezacaftor-ivacaftor therapy have reported similar results at up to 144 weeks of follow-up.⁸⁹

Elexacaftor-tezacaftor-ivacaftor is approved for patients aged 2 years or older; approximately 90% of people with cystic fibrosis, including for those with variants that have demonstrated in vitro culture response to treatment.^{80,90-92} This technique known as *thera-typing* has increased access to therapy with modulator drugs among people with rare (<1%) *CFTR* variants.⁹²

Ivacaftor and its combination *CFTR* modulator were generally well tolerated and had similar safety profiles in phase 3 studies involving younger age groups (Table 2).^{69-71,74,75,80,90,91,93,94} Compared with placebo, elexacaftor-tezacaftor-ivacaftor had a similar incidence of adverse events (93.1% vs 96.1%) including headache (17%), upper respiratory tract infection (16%), abdominal pain (14%), diarrhea (13%), exanthem (10%), increased alanine transaminase (10%), or aspartate transaminase (9%).²³ Serious adverse events were less common in the treatment group (13.9% vs 20.9%).²³ For all *CFTR* modulator therapies, liver function monitoring is recommended quarterly for the first year of treatment and then annually.^{67,72,76,78} Ophthalmologic examinations for children are recommended annually based on toxicology studies of ivacaftor that identified cataracts in juvenile rats, although this adverse effect was not observed in human trials.⁶⁷ Drug interactions are important considerations because ivacaftor and combination therapies are both substrates and inducers in the cytochrome P450 (*CYP3A*) pathway (Table 2).^{67,72,76,78}

Prognosis

In 2021, the median age of survival in the US was approximately 53.1 years (95% CI, 51.6-54.7 years) for people born from 2017 through 2021. In comparison, for people with cystic fibrosis born from 2001 through 2006, life expectancy was approximately 36.3 years (95% CI, 35.1-37.9 years). The US annual mortality rate was 1.5 deaths per 100 in 2006 and 0.7 deaths per 100 in 2021.¹ Registry data across multiple other countries such as the UK, Germany, and Canada have reported similar improvements in life expectancy. Current life expectancy in the UK, Germany, and Canada is approximately 47 to 53 years.⁹⁵

Newborn screening for early diagnosis of cystic fibrosis has been associated with improved health.¹² A prospective, observational

Table 2. CFTR Modulator Therapies for Cystic Fibrosis

| Modulator name | Common eligible genotypes (prevalence in US) ¹ | Ages approved and adult dosing | Efficacy | Adverse effects | Monitoring and drug interactions |
|----------------------|---|---|---|--|---|
| Ivacaftor | Approved use for use with ≥1 copy of G551D (4.2%) R117H (3.3%) 3849 + 10kbC→T (1.8%) ^a 2789 + 5G→A (1.5%) ^a D1152H (1.1%) 3272-26A→G (0.8%) ^a L206W (0.7%) A455E (0.6%) For all eligible 97 variants, refer to prescribing information ⁶⁷ | Approved for ages ≥1 mo; Dose for ages ≥6 y: one 150-mg tablet every 12 h; for ages 4 mo to 6 y, see prescribing information ⁶⁷ | For patients with a G551D-CFTR variant, ivacaftor vs placebo increased mean absolute change from baseline lung function (ppFEV ₁) through wk 24 (10.4% vs -0.2%; 95% CI, 8.6% to 12.6%; <i>P</i> < .001) and wk 48 (10.1% vs -0.4%; 95% CI, 8.5% to 12.5%; <i>P</i> < .001); reduced pulmonary exacerbations at wk 28 (rate ratio, 0.38; 95% CI, 0.22 to 0.64; <i>P</i> < .001); and improved weight gain from baseline to wk 28 (difference, 2.8 kg; 95% CI, 1.8 to 3.7; <i>P</i> < .001); reduced sweat chloride concentration (-48.7 vs -0.06 mEq/L; mean difference, -48 mEq/L; <i>P</i> < .001) ⁶⁸ In an open-label, single-arm study of children (aged 2-5 y) who received ivacaftor and had 1 gating gene variant, mean-weight-for-age z score increased 0.2 from baseline at wk 24 (<i>P</i> < .001) ⁶⁹ In single-arm study of infants (12-24 mo) receiving ivacaftor, mean weight-for-age z score was maintained 0.15 (95% CI, -0.05 to 0.36) from baseline ⁷⁰ Ivacaftor for 4 mo-<12 mo, mean weight-for-age z score increased 0.52 (95% CI, 0.23 to 0.82) from baseline ⁷¹ | AEs were similar for ivacaftor vs placebo to 48 wk (82 vs 78 participants), but lower incidence of cough (33% vs 42%), pulmonary exacerbation (13% vs 33%) in the treated group; AEs more commonly reported for ivacaftor vs placebo: headache (22.9%), respiratory tract infection (22.9%), nasal congestion (17%), rash (14.5%), dizziness (12%), increased hepatic enzyme levels led to study drug discontinuation for 1 in the ivacaftor group vs 4 in the placebo group ⁶⁸ | Elevated transaminases: ALT or AST assessed prior to initiation and every 3 mo for first y of treatment, then annually; increase monitoring frequency for history of elevations; interrupt dose if ALT or AST >5 × ULN Cataracts: reported in pediatric patients; baseline and follow-up eye examinations recommended for patients ≤18 y Drug interactions: reduce ivacaftor dose or avoid CYP3A inhibitors (eg, ketoconazole, voriconazole, clarithromycin, erythromycin, food containing grapefruit); avoid coadministration with strong CYP3A inducers (eg, rifampin, phenobarbital, St John's wort) that decrease ivacaftor exposure; caution and monitoring for medications categorized as CYP2C9 substrates (eg, warfarin, glipizide) and CYP3A and/or P-gp substrates (eg, digoxin, cyclosporine, tacrolimus) Ivacaftor may increase exposure of such medications |
| Lumacaftor-ivacaftor | F508del homozygous (44.1%) | Approved for ages ≥1 y Dose for ages ≥12 y: 2 tablets combined of 200-mg lumacaftor and 125-mg ivacaftor every 12 h Doses for ages 1 y to 11 y, refer to prescribing information ⁷² | In pooled analysis of 2 studies, lumacaftor-ivacaftor vs placebo improved: mean absolute difference of ppFEV ₁ (range, 2.8 to 3.3, <i>P</i> < .001); reduced pulmonary exacerbations rate (range, 0.61 to 0.70; <i>P</i> = .001); increased absolute change in BMI (range, 0.24 to 0.28; <i>P</i> < .001). ^{73,74} | Incidence of AEs were similar in lumacaftor-ivacaftor-treated and placebo groups up to 24 wk, but higher proportion of patients who discontinued study drug because of an AE in lumacaftor-ivacaftor group (4.2% vs 1.6%) ⁷³ Common AEs: dyspnea (13%), nasopharyngitis (13%), nausea (13%), rash (7%), and elevated blood creatine phosphokinase (7%); elevation of liver transaminases, rash; and drug interactions have also been reported ^{74,75} | Patients: caution use and consider reduced dose in patients with advanced liver disease; increased monitoring for respiratory symptoms at initiation in patients with ppFEV ₁ < 40% Elevated transaminases (ALT, AST, bilirubin): same monitoring as per ivacaftor and interrupt dose if ALT or AST >3 × ULN with bilirubin >2 × ULN Blood pressure: periodically measure blood pressure in all patients for elevations Drug interactions: same as ivacaftor above; and interacts with CYP3A substrates or CYP3A substrates with narrow therapeutic index, including reducing effects of hormonal contraceptives Cataracts: same monitoring as ivacaftor |
| | F508del homozygous (44.1%); or 1 copy of all variants for ivacaftor; or 1 copy of additional 57 variants; for all eligible variants, refer to prescribing information ⁷⁶ | Approved for ages ≥6 y Dose for ages ≥12 y: 1 tablet tezacaftor 100 mg/ivacaftor 150 mg in AM; 1 tablet ivacaftor 150 mg in PM Doses for ages 6 y to 11 y, refer to prescribing information ⁷⁶ | Tezacaftor-ivacaftor vs placebo improved mean absolute difference of ppFEV ₁ (3.4% vs -0.6%, <i>P</i> < .001) through wk 24; reduced pulmonary exacerbation rate (0.65; 95% CI, 0.48, 0.88; <i>P</i> = .005) ⁷⁷ Tezacaftor-ivacaftor improved ppFEV ₁ (6.8%, <i>P</i> < .001) vs placebo ⁵³ | Most AEs deemed unrelated to study drug; common adverse drug reactions (occurring in ≥3% of patients) were headache, nausea, sinus congestion, and dizziness; study drug interruption most commonly from elevated liver transaminases ⁵³ | Elevated transaminases (ALT, AST, bilirubin): same monitoring as per lumacaftor-ivacaftor Cataracts: same monitoring as ivacaftor Drug Interactions: same as ivacaftor; reduce dose with strong or moderate CYP3A inhibitors; avoid coadministration with strong CYP3A inducers and food containing grapefruit |

(continued)

Table 2. *CFTR* Modulator Therapies for Cystic Fibrosis (continued)

| Modulator name | Common eligible genotypes (prevalence in US) ¹ | Ages approved and adult dosing | Efficacy | Adverse effects | Monitoring and drug interactions |
|----------------------------------|--|---|--|---|---|
| Elexacaftor-tezacaftor-ivacaftor | <p>≥1 copy of <i>F508del</i> (85.5%); <i>G85E</i> (0.7%)</p> <p>or all variants for tezacaftor-ivacaftor (except as indicated)^a</p> <p>or of additional 30 <i>CFTR</i> variants; all eligible variants, refer to prescribing information⁷⁸</p> | <p>Approved for ages ≥2 y</p> <p>Dose for ages ≥12 y: 2 tablets, each containing 100-mg elexacaftor, 50-mg tezacaftor, and 75-mg ivacaftor in AM; 1 tablet of 150-mg ivacaftor in PM</p> <p>Doses for ages 2 y to 11 y, refer to prescribing information⁷⁸</p> | <p>In patients who were <i>F508del</i> homozygous, elexacaftor-tezacaftor-ivacaftor vs placebo increased: absolute change in lung function (ppFEV₁) from baseline at 29 d (13.8 vs 0.4; difference, 11.0; 95% CI, 7.9 to 14.0; <i>P</i> < .001)²²</p> <p>In patients with <i>F508del</i>-minimal function genotypes, elexacaftor-tezacaftor-ivacaftor vs placebo increased absolute change in lung function (ppFEV₁) from baseline at wk 4 (13.6% vs -0.2%; mean difference, 13.8%; 95% CI, 12.1% to 15.4%; <i>P</i> < .001) and through wk 24 (13.9% vs -0.4%; mean difference, 14.3%; 95% CI, 12.7% to 15.8%; <i>P</i> < .001)²³</p> <p>In participants with <i>F508del</i>-gating or <i>F508del</i>-residual function variants, elexacaftor-tezacaftor-ivacaftor vs active control increased lung function (ppFEV₁) from active control by 3.5 percentage points (95% CI, 2.2 to 4.7)⁷⁹</p> <p>In children aged 6-11 y, elexacaftor-tezacaftor-ivacaftor improved the lung clearance index (-2.29 vs -0.02 units; <i>P</i> < .001)⁸⁰</p> | <p>Similar incidence of AEs seen in elexacaftor-tezacaftor-ivacaftor vs placebo; common AEs (≥5% of patients and higher than placebo by ≥1%): headache (17%), upper respiratory tract infection (16%), abdominal pain (14%), diarrhea (13%), rash (10%), increased ALT levels (10%), increased nasal congestion, blood creatine phosphokinase levels (9%), and AST increased (9%); rash led to 1% study drug discontinuation vs <1% placebo²³</p> | <p>Elevated transaminases (ALT, AST, bilirubin): same monitoring as per lumacaftor-ivacaftor and tezacaftor-ivacaftor; more frequent monitoring for people with advanced liver disease or history of elevations</p> <p>Cataracts: same monitoring as ivacaftor</p> <p>Drug Interactions: same as tezacaftor-ivacaftor; reduce dose with strong or moderate CYP3A inhibitors; avoid coadministration with strong CYP3A inducers and food containing grapefruit</p> |

Abbreviations: AE, adverse effect; ALT, alanine transaminase; AST, aspartate transaminase; BMI, body mass index; *CFRT*, cystic fibrosis transmembrane conductance regulator; ppFEV₁, predicted percent of forced expiratory volume in the first second of expiration; ULN, upper limit of normal.

^a Variants eligible for ivacaftor and tezacaftor-ivacaftor only and not eligible for elexacaftor-tezacaftor-ivacaftor therapy.

study of 231 infants with cystic fibrosis diagnosed by universal newborn screening in the US demonstrated that infants diagnosed during newborn screening had minimal respiratory symptoms and were able to achieve normal weight for age by 1 year, although their mean lengths were lower than the World Health Organization standard growth curves for healthy infants.⁹⁶ A retrospective cohort study comparing the longitudinal outcomes of 9571 infants in the same birth cohort before and after implementation of the cystic fibrosis newborn screening program by state found that a newborn screening program was associated with higher median weight (6.0, 95% CI, 3.1-8.4) and median height (6.6; 95% CI, 3.8-9.3) percentiles in the first year of life and was associated with older age at onset of chronic *Pseudomonas* infection (hazard ratio [HR], 0.69; 95% CI, 0.54-0.89), but no association with lung function (ppFEV₁) at age 6 years.⁹⁷

Observational data showed that modulator therapy was associated with improved health.^{98,99} Among 2509 patients with cystic fibrosis from the US and UK who were followed up from 2011 (pretreatment baseline) to 2016, the 635 patients treated with ivacaftor maintained higher lung function and higher BMI compared with the 1874 patients not treated with ivacaftor.⁹⁸ Patients who received ivacaftor had an FEV₁ decline of -0.7% at the 5-year follow-up compared with 8.3% in the group that did not receive ivacaftor. In the PROMISE⁹⁹ observational study of 487 people with cystic fibrosis who were treated with elexacaftor-tezacaftor-ivacaftor, sustained improvement in lung function, measured by ppFEV₁, was observed at the 6-month follow-up compared with pretreatment baseline (90.9% vs 80.5%; difference, 9.8%; 95% CI, 8.8% to 10.8%). In this cohort, elexacaftor-tezacaftor-ivacaftor use was associated with improved *CFTR* function as measured by sweat chloride concentration (after treatment, 45.7 mEq/L vs baseline, 88.0 mEq/L; difference, -41.7 mEq/L; 95% CI, -43.8 to -39.6 mEq/L).⁹⁹ In a prospective

observational study of people with cystic fibrosis with advanced lung disease (defined as ppFEV₁ < 40%), elexacaftor-tezacaftor-ivacaftor use was associated with an absolute change from baseline ppFEV₁ of 15.1%.¹⁰⁰ The proportion of people with cystic fibrosis who required supplemental oxygen declined from 43.4% at initiation to 23.4% after 3 months of elexacaftor-tezacaftor-ivacaftor treatment and requirements for noninvasive ventilation were reduced from 28.1% to 19.8%.¹⁰⁰ The 2021 Cystic Fibrosis Foundation Patient Registry¹ reported a decline in number of lung transplants from 197 in 2006 to 54 in 2021.

Improvements in longevity observed in cystic fibrosis are not equal by race or ethnicity in the US. Hispanic patients compared with non-Hispanic patients had significantly higher mortality after adjustment for clinical and socioeconomic factors (9.1% vs 3.3%; HR, 2.8; 95% CI, 1.7 to 4.6).¹⁰¹ These differences in outcomes may be due in part to delayed diagnosis and care initiation. Among 6354 infants born between 2010 through 2018, initiating cystic fibrosis care occurred later among infants described as American Indian and Native Alaskan, Asian, Black or African American, and/or other race, and/or Hispanic ethnicity (group 1), compared with infants described as White and non-Hispanic (group 2). The median age at the first clinical evaluation for cystic fibrosis among infants in group 1 was 31 days (IQR, 19-49 days) vs 22 days (IQR, 14-36) for those in group 2. Delayed cystic fibrosis care was associated with worse nutrition. Measured by weight-for-age z scores at 1 year of age, the median z score for group 1 was -0.11 (IQR, -0.75 to 0.59) vs 0.062 (IQR, -0.57 to 0.65) for group 2.¹⁰² *CFTR* genetic panels and selection of variants tested differ by state and often represent the most common variants. In a cross-sectional study of 7 *CFTR* genetic panels used in newborn screening, detection of at least 1 *CFTR* variant was lowest in infants identified as Black, Asian, and Hispanic compared with infants categorized as non-Hispanic White

for all panels (41.9%-93.1% vs 87.5%-97.0%).¹⁰³ Panel choice by state may contribute to inequities in delays in diagnosis of cystic fibrosis. The *CFTR* genetic variants present in Black patients (69.7%) and Hispanic patients (75.6%) have fewer matching *CFTR* modulators available for treatment compared with the variants present in White patients (92.4%).¹⁰⁴

Practical Considerations and Application of Evidence

As health improves, particularly after initiation of elexacaftor-tezacaftor-ivacaftor, people with cystic fibrosis have expressed interest in reducing the number of treatments.¹⁰⁵ Randomized clinical trials and observational studies are underway to identify therapies that can be reduced or eliminated in patients receiving *CFTR* modulator therapies. The SIMPLIFY study¹⁰⁵ included 2 independently conducted randomized, clinical trials assessing the noninferiority of discontinuing vs continuing 6 weeks of either hypertonic saline (184 discontinued vs 186 continued) or dornase alfa (240 discontinued vs 234 continued). In the hypertonic saline trial, discontinuing hypertonic saline was non-inferior to continuing with respect to the 6-week change in ppFEV₁ (-0.19% vs 0.14%; difference, -0.32%; 95% CI, -1.25 to 0.60). In the dornase alfa trial, discontinuing dornase alfa was noninferior to continuing with respect to the 6-week change in ppFEV₁ (0.18% vs 0.16%; difference, 0.35%; 95% CI, -0.45% to 1.14%).¹⁰⁵ The SIMPLIFY study is the first step toward understanding the need for standard therapies for people receiving modulators.

Wholesale acquisition costs of *CFTR* modulators can range from \$272 623 to \$311 741 per year and raise concerns about affordability and access.¹⁰⁶ However, the final reimbursement cost is based on agreements with health authorities or private payers and is variable. In the US, *CFTR* modulator therapies were prescribed to 91% (range, 75.9%-100%) of people with eligible *CFTR* variants in 2021, funded by both private and public payers.¹ More than 40 countries, including Australia, Europe, Israel, New Zealand, and North America have regulatory and reimbursement approvals permitting access to elexacaftor-tezacaftor-ivacaftor combination therapy. However, other global areas such as India, and in regions of Central and South America, the Middle East, and Southern Africa are awaiting approval for elexacaftor-tezacaftor-ivacaftor, which could widen disparities in cystic fibrosis outcomes between high-income and low- and middle-income countries.¹⁰⁷

Several knowledge gaps about modulator therapies must be addressed. Long-term pharmacovigilance is needed to understand the safety of these drugs including drug-drug interactions and effects such as weight gain, elevated blood pressure, mental health effects, and liver function abnormalities.⁹⁴ The long-term safety and effectiveness of *CFTR* modulators prenatally and in infants younger than 4 months is unknown.¹⁰⁸

Adults comprise 58.3% of the total US cystic fibrosis population. As this population ages, expanded services will be needed

across multiple subspecialties to manage comorbidities and age-related complications including gastrointestinal cancers, diabetes, obesity, and hypertension.^{1,50,63,94,109} Rates of pregnancy in women with cystic fibrosis have increased from 210 in 2011 to 675 in 2021, underscoring the importance of attending to reproductive health.^{1,110} The association of cystic fibrosis with mental health has been characterized in observational studies, but interventional studies are needed.^{62,111,112} In an observational study of 1005 patients with cystic fibrosis, patients who tested positive (mean hazard rate, 29.4 deaths per 1000 patients per year) for any depression screening tool had a higher 5-year mortality rate than those who tested negative for depression (mean hazard rate, 15.7 deaths per 1000 patients per year; unadjusted HR, 2.0; 95% CI, 1.3-3.0), but the association was attenuated and no longer statistically significant after adjustment for potential confounders (adjusted HR, 1.4; 95% CI, 0.9-2.2).¹¹¹

Future studies should identify effective therapies for all people with cystic fibrosis, regardless of the genetic variant. For example, therapies are needed for people with genetic variants that are not modulator responsive such as premature termination codons, large deletions or frameshifts that produce little or no stable protein.¹¹³ Although multiple gene replacement programs, both virally and nonvirally based, have been attempted,¹¹⁴ none have demonstrated efficacy. The availability of gene editing technologies such as clustered regularly interspaced short palindromic repeats (CRISPR) CRISPR-associated protein 9 may lead to functional *CFTR* repair in intestinal or pulmonary epithelia.¹¹⁵ All genetic based therapies face the challenge of developing efficient vectors that can deliver stable product to the target stem cells in the airway or intestinal tract.¹¹³

Limitations

This review has several limitations. First, some relevant publications may have been missed. Second, this was not a systematic review. Third, the review focused on *CFTR* modulators for which long-term safety and efficacy data are not available. Fourth, topics such as lung transplant, the effect of COVID-19, digital health, or management of cystic fibrosis-related comorbidities such as diabetes or liver disease were not covered.

Conclusions

Cystic fibrosis affects approximately 89 000 identified people worldwide and is associated with a spectrum of disease related to exocrine dysfunction, including chronic respiratory bacterial infections and reduced life expectancy. First-line pulmonary therapies consist of mucolytics, anti-inflammatories, and antibiotics, and approximately 90% of people with cystic fibrosis 2 years and older benefit from a combination of ivacaftor, tezacaftor, and elexacaftor.

ARTICLE INFORMATION

Accepted for Publication: May 2, 2023.

Conflict of Interest Disclosures: Dr Ong reported receiving grants from the Agency for Healthcare Research and Quality, the National Institutes of Health, and the Cystic Fibrosis Foundation; nonfinancial support from the Cystic Fibrosis

Foundation; and honorarium from the Cincinnati Children's Hospital for participation in the Cystic Fibrosis Learning Network Leadership outside the submitted work. Dr Ramsey reported receiving grants from the National Institutes of Health and the Cystic Fibrosis Foundation and having served on the advisory boards of Vertex Pharmaceuticals

Scientific, Sionna Therapeutics, and Cystic Medicines outside the submitted work.

Additional Contributions: We thank Elisabeth Nylander, MSc, medical librarian, for reference management and Melissa Harkleroad for aid in manuscript preparation and formatting, both at

Seattle Children's Hospital. Neither received financial compensation.

Submissions: We encourage authors to submit papers for consideration as a Review. Please contact Mary McGrae McDermott, MD, at mdm608@northwestern.edu.

REFERENCES

- Cystic Fibrosis Foundation. *National Patient Registry 2021: Annual Data Report*. 2023. Accessed April 23, 2023. <https://www.cff.org/sites/default/files/2021-11/Patient-Registry-Annual-Data-Report.pdf>
- Bell SC, Mall MA, Gutierrez H, et al. The future of cystic fibrosis care: a global perspective. *Lancet Respir Med*. 2020;8(1):65-124. doi:10.1016/S2213-2600(19)30337-6
- Cystic fibrosis mutation database (CFTR1). Updated April 7, 2023. Accessed April 23, 2023. <http://www.genet.sickkids.on.ca/>
- Gentzsch M, Mall MA. Ion channel modulators in cystic fibrosis. *Chest*. 2018;154(2):383-393. doi:10.1016/j.chest.2018.04.036
- Boyle MP, De Boeck K. A new era in the treatment of cystic fibrosis: correction of the underlying CFTR defect. *Lancet Respir Med*. 2013;1(2):158-163. doi:10.1016/S2213-2600(12)70057-7
- The clinical and functional translation of CFTR (CFTR2). Cystic Fibrosis Foundation, Johns Hopkins Medicine, and Sequenom Laboratories. Accessed April 23, 2023. <http://cftr2.org>
- Shteinberg M, Haq IJ, Polineni D, Davies JC. Cystic fibrosis. *Lancet*. 2021;397(10290):2195-2211. doi:10.1016/S0140-6736(20)32542-3
- Farrell PM. The prevalence of cystic fibrosis in the European Union. *J Cyst Fibros*. 2008;7(5):450-453. doi:10.1016/j.jcf.2008.03.007
- Singh M, Rebordosa C, Bernholz J, Sharma N. Epidemiology and genetics of cystic fibrosis in Asia: in preparation for the next-generation treatments. *Respirology*. 2015;20(8):1172-1181. doi:10.1111/resp.12656
- Pérez MM, Luna MC, Pivetta OH, Keyeux G. CFTR gene analysis in Latin American CF patients: heterogeneous origin and distribution of mutations across the continent. *J Cyst Fibros*. 2007;6(3):194-208. doi:10.1016/j.jcf.2006.07.004
- Stoltz DA, Meyerholz DK, Welsh MJ. Origins of cystic fibrosis lung disease. *N Engl J Med*. 2015;372(4):351-362. doi:10.1056/NEJMra1300109
- Farrell PM, White TB, Ren CL, et al. Diagnosis of cystic fibrosis: consensus guidelines from the Cystic Fibrosis Foundation. *J Pediatr*. 2017;181S(181)(suppl):S4-S15, 15.e1. doi:10.1016/j.jpeds.2016.09.064
- Birket SE, Davis JM, Fernandez CM, et al. Development of an airway mucus defect in the cystic fibrosis rat. *JCI Insight*. 2018;3(1):e97199. doi:10.1172/jci.insight.97199
- Oppenheimer EH, Esterly JR. Pathology of cystic fibrosis review of the literature and comparison with 146 autopsied cases. *Perspect Pediatr Pathol*. 1975;2:241-278.
- Sturgess J, Imrie J. Quantitative evaluation of the development of tracheal submucosal glands in infants with cystic fibrosis and control infants. *Am J Pathol*. 1982;106(3):303-311.
- Sly PD, Gangell CL, Chen L, et al; AREST CF Investigators. Risk factors for bronchiectasis in children with cystic fibrosis. *N Engl J Med*. 2013;368(21):1963-1970. doi:10.1056/NEJMoa1301725
- Stick SM, Brennan S, Murray C, et al; Australian Respiratory Early Surveillance Team for Cystic Fibrosis (AREST CF). Bronchiectasis in infants and preschool children diagnosed with cystic fibrosis after newborn screening. *J Pediatr*. 2009;155(5):623-8.e1. doi:10.1016/j.jpeds.2009.05.005
- Sun X, Yi Y, Yan Z, et al. In utero and postnatal VX-770 administration rescues multiorgan disease in a ferret model of cystic fibrosis. *Sci Transl Med*. 2019;11(485):eaau7531. doi:10.1126/scitranslmed.aau7531
- Veit G, Avramescu RG, Chiang AN, et al. From CFTR biology toward combinatorial pharmacotherapy: expanded classification of cystic fibrosis mutations. *Mol Biol Cell*. 2016;27(3):424-433. doi:10.1091/mbc.e14-04-0935
- Welsh MJ, Smith AE. Molecular mechanisms of CFTR chloride channel dysfunction in cystic fibrosis. *Cell*. 1993;73(7):1251-1254. doi:10.1016/0092-8674(93)90353-R
- Mall MA, Mayer-Hamblett N, Rowe SM. Cystic fibrosis: emergence of highly effective targeted therapeutics and potential clinical implications. *Am J Respir Crit Care Med*. 2020;201(10):1193-1208. doi:10.1164/rccm.201910-1943S0
- Heijerman HGM, McKone EF, Downey DG, et al; VX17-445-103 Trial Group. Efficacy and safety of the elxacaftor plus tezacaftor plus ivacaftor combination regimen in people with cystic fibrosis homozygous for the F508del mutation: a double-blind, randomised, phase 3 trial. *Lancet*. 2019;394(10121):1940-1948. doi:10.1016/S0140-6736(19)32597-8
- Middleton PG, Mall MA, Dřevíněk P, et al; VX17-445-102 Study Group. Elxacaftor-tezacaftor-ivacaftor for cystic fibrosis with a single Phe508del allele. *N Engl J Med*. 2019;381(19):1809-1819. doi:10.1056/NEJMoa1908639
- Cutting GR. Treating specific variants causing cystic fibrosis. *JAMA*. 2017;318(21):2130-2131. doi:10.1001/jama.2017.16823
- Singh VK, Schwarzenberg SJ. Pancreatic insufficiency in cystic fibrosis. *J Cyst Fibros*. 2017;16(suppl 2):S70-S78. doi:10.1016/j.jcf.2017.06.011
- Ranganathan SC, Hall GL, Sly PD, Stick SM, Douglas TA; Australian Respiratory Early Surveillance Team for Cystic Fibrosis (AREST-CF). Early lung disease in infants and preschool children with cystic fibrosis: what have we learned and what should we do about it? *Am J Respir Crit Care Med*. 2017;195(12):1567-1575. doi:10.1164/rccm.201606-1107CI
- Flume PA, Robinson KA, O'Sullivan BP, et al; Clinical Practice Guidelines for Pulmonary Therapies Committee. Cystic fibrosis pulmonary guidelines: airway clearance therapies. *Respir Care*. 2009;54(4):522-537.
- Wilson LM, Morrison L, Robinson KA. Airway clearance techniques for cystic fibrosis: an overview of Cochrane systematic reviews. *Cochrane Database Syst Rev*. 2019;1(1):CD011231. doi:10.1002/14651858.CD011231.pub2
- Mogayzel PJ Jr, Naureckas ET, Robinson KA, et al; Pulmonary Clinical Practice Guidelines Committee. Cystic fibrosis pulmonary guidelines: chronic medications for maintenance of lung health. *Am J Respir Crit Care Med*. 2013;187(7):680-689. doi:10.1164/rccm.201207-11600E
- Castellani C, Duff AJA, Bell SC, et al. ECFS best practice guidelines: the 2018 revision. *J Cyst Fibros*. 2018;17(2):153-178. doi:10.1016/j.jcf.2018.02.006
- Yang C, Montgomery M. Dornase alfa for cystic fibrosis. *Cochrane Database Syst Rev*. 2021;3(3):CD001127. doi:10.1002/14651858.cd001127.pub5
- Wark P, McDonald VM. Nebulised hypertonic saline for cystic fibrosis. *Cochrane Database Syst Rev*. 2018;9(9):CD001506. doi:10.1002/14651858.cd001127.pub5
- Ratjen F, Davis SD, Stanojevic S, et al; SHIP Study Group. Inhaled hypertonic saline in preschool children with cystic fibrosis (SHIP): a multicentre, randomised, double-blind, placebo-controlled trial. *Lancet Respir Med*. 2019;7(9):802-809. doi:10.1016/S2213-2600(19)30187-0
- Stahl M, Wielpütz MO, Ricklefs I, et al. Preventive Inhalation of Hypertonic Saline in Infants with Cystic Fibrosis (PRESES): a randomized, double-blind, controlled study. *Am J Respir Crit Care Med*. 2019;199(10):1238-1248. doi:10.1164/rccm.201807-1203OC
- Nevitt SJ, Thornton J, Murray CS, Dwyer T. Inhaled mannitol for cystic fibrosis. *Cochrane Database Syst Rev*. 2020;5(5):CD008649. doi:10.1002/14651858.cd008649.pub4
- Flume PA, Amelina E, Daines CL, et al. Efficacy and safety of inhaled dry-powder mannitol in adults with cystic fibrosis: an international, randomized controlled study. *J Cyst Fibros*. 2021;20(6):1003-1009. doi:10.1016/j.jcf.2021.02.011
- Zimmermann P, Ziesenitz VC, Curtis N, Ritz N. The immunomodulatory effects of macrolides—a systematic review of the underlying mechanisms. *Front Immunol*. 2018;9:302. doi:10.3389/fimmu.2018.00302
- Southern KW, Barker PM, Solis-Moya A, Patel L. Macrolide antibiotics for cystic fibrosis. *Cochrane Database Syst Rev*. 2012;11(11):CD002203. doi:10.1002/14651858.cd002203.pub4
- Stick SM, Foti A, Ware RS, et al; COMBAT CF Study Group. The effect of azithromycin on structural lung disease in infants with cystic fibrosis (COMBAT CF): a phase 3, randomised, double-blind, placebo-controlled clinical trial. *Lancet Respir Med*. 2022;10(8):776-784. doi:10.1016/S2213-2600(22)00165-5
- Lands LC, Stanojevic S. Oral non-steroidal anti-inflammatory drug therapy for lung disease in cystic fibrosis. *Cochrane Database Syst Rev*. 2019;9(9):CD001505. doi:10.1002/14651858.CD001505.pub5
- Mendelman PM, Smith AL, Levy J, Weber A, Ramsey B, Davis RL. Aminoglycoside penetration, inactivation, and efficacy in cystic fibrosis sputum. *Am Rev Respir Dis*. 1985;132(4):761-765. doi:10.1164/arrd.1985.132.4.761
- Smith S, Rowbotham NJ. Inhaled anti-pseudomonal antibiotics for long-term therapy in cystic fibrosis. *Cochrane Database Syst Rev*. 2018;3(3):CD001021. doi:10.1002/14651858.cd001021.pub4
- Heijerman H, Westerman E, Conway S, Touw D, Döring G; Consensus Working Group. Inhaled medication and inhalation devices for lung disease in patients with cystic fibrosis: a European

- consensus. *J Cyst Fibros*. 2009;8(5):295-315. doi:10.1016/j.jcf.2009.04.005
44. Flume PA, Mogayzel PJ Jr, Robinson KA, et al; Clinical Practice Guidelines for Pulmonary Therapies Committee. Cystic fibrosis pulmonary guidelines: treatment of pulmonary exacerbations. *Am J Respir Crit Care Med*. 2009;180(9):802-808. doi:10.1164/rccm.200812-1845PP
45. Goss CH. Acute pulmonary exacerbations in cystic fibrosis. *Semin Respir Crit Care Med*. 2019;40(6):792-803. doi:10.1055/s-0039-1697975
46. Regelman WE, Elliott GR, Warwick WJ, Clawson CC. Reduction of sputum *Pseudomonas aeruginosa* density by antibiotics improves lung function in cystic fibrosis more than do bronchodilators and chest physiotherapy alone. *Am Rev Respir Dis*. 1990;141(4 pt 1):914-921. doi:10.1164/ajrccm/141.4.Pt.1.914
47. Goss CH, Heltshe SL, West NE, et al; STOP2 Investigators. A randomized clinical trial of antimicrobial duration for cystic fibrosis pulmonary exacerbation treatment. *Am J Respir Crit Care Med*. 2021;204(11):1295-1305. doi:10.1164/rccm.202102-0461OC
48. Sanders DB, Khan U, Heltshe SL, et al; STOP2 Investigators. Association of site of treatment with clinical outcomes following intravenous antimicrobial treatment of a pulmonary exacerbation. *J Cyst Fibros*. 2022;21(4):574-580. doi:10.1016/j.jcf.2021.11.009
49. Kapnadak SG, Dimango E, Hadjilias D, et al. Cystic Fibrosis Foundation consensus guidelines for the care of individuals with advanced cystic fibrosis lung disease. *J Cyst Fibros*. 2020;19(3):344-354. doi:10.1016/j.jcf.2020.02.015
50. Ode KL, Ballman M, Battezzati A, et al. ISPAD Clinical Practice Consensus Guidelines 2022: management of cystic fibrosis-related diabetes in children and adolescents. *Pediatr Diabetes*. 2022;23(8):1212-1228. doi:10.1111/ledi.13453
51. Sasame A, Stokes D, Bourke B, Connolly L, Fitzpatrick E, Rowland M. The impact of liver disease on mortality in cystic fibrosis—a systematic review. *J Cyst Fibros*. 2022;21(2):202-211. doi:10.1016/j.jcf.2021.07.014
52. Putman MS, Anabawi A, Le T, Tangpricha V, Sermet-Gaudelus I. Cystic fibrosis bone disease treatment: current knowledge and future directions. *J Cyst Fibros*. 2019;18(suppl 2):S56-S65. doi:10.1016/j.jcf.2019.08.017
53. Rowe SM, Daines C, Ringshausen FC, et al. Tezacaftor-ivacaftor in residual-function heterozygotes with cystic fibrosis. *N Engl J Med*. 2017;377(21):2024-2035. doi:10.1056/NEJMoa1709847
54. Martiniano SL, Croak K, Bonn G, Sontag MK, Sagel SD. Improving outcomes for Colorado's IRT-IRT-DNA cystic fibrosis newborn screening algorithm by implementing floating cutoffs. *Mol Genet Metab*. 2021;134(1-2):65-67. doi:10.1016/j.ymgme.2021.08.005
55. Rehani MR, Marcus MS, Harris AB, Farrell PM, Ren CL. Variation in cystic fibrosis newborn screening algorithms in the United States. *Pediatr Pulmonol*. 2023;58(3):927-933. doi:10.1002/ppul.26279
56. Rueegg CS, Kuehni CE, Gallati S, et al; Swiss Cystic Fibrosis Screening Group. Comparison of two sweat test systems for the diagnosis of cystic fibrosis in newborns. *Pediatr Pulmonol*. 2019;54(3):264-272. doi:10.1002/ppul.24227
57. Ren CL, Borowitz DS, Gonska T, et al. Cystic fibrosis transmembrane conductance regulator-related metabolic syndrome and cystic fibrosis screen positive, inconclusive diagnosis. *J Pediatr*. 2017;181S(181)(suppl):S45-S51. doi:10.1016/j.jpeds.2016.09.066
58. Barben J, Castellani C, Munck A, et al; European CF Society Neonatal Screening Working Group (ECFS NSWG). Updated guidance on the management of children with cystic fibrosis transmembrane conductance regulator-related metabolic syndrome/cystic fibrosis screen positive, inconclusive diagnosis (CRMS/CFSPID). *J Cyst Fibros*. 2021;20(5):810-819. doi:10.1016/j.jcf.2020.11.006
59. Fajac I, Hubert D, Guillemot D, et al. Nasal airway ion transport is linked to the cystic fibrosis phenotype in adult patients. *Thorax*. 2004;59(11):971-976. doi:10.1136/thx.2003.020933
60. Conway S, Balfour-Lynn IM, De Rijcke K, et al. European Cystic Fibrosis Society Standards of Care: framework for the Cystic Fibrosis Centre. *J Cyst Fibros*. 2014;13(suppl 1):S3-S22. doi:10.1016/j.jcf.2014.03.009
61. Borowitz D, Robinson KA, Rosenfeld M, et al; Cystic Fibrosis Foundation. Cystic Fibrosis Foundation evidence-based guidelines for management of infants with cystic fibrosis. *J Pediatr*. 2009;155(6)(suppl):S73-S93. doi:10.1016/j.jpeds.2009.09.001
62. Quittner AL, Abbott J, Georgiopoulos AM, et al; International Committee on Mental Health; EPOS Trial Study Group. International Committee on Mental Health in Cystic Fibrosis: Cystic Fibrosis Foundation and European Cystic Fibrosis Society consensus statements for screening and treating depression and anxiety. *Thorax*. 2016;71(1):26-34. doi:10.1136/thoraxjnl-2015-207488
63. Hadjilias D, Khoruts A, Zuber AG, Hempstead SE, Maisonneuve P, Lowenfels AB; Cystic Fibrosis Colorectal Cancer Screening Task Force. Cystic fibrosis colorectal cancer screening consensus recommendations. *Gastroenterology*. 2018;154(3):736-745.e14. doi:10.1053/j.gastro.2017.12.012
64. Fuchs HJ, Borowitz DS, Christiansen DH, et al; The Pulmozyme Study Group. Effect of aerosolized recombinant human DNase on exacerbations of respiratory symptoms and on pulmonary function in patients with cystic fibrosis. *N Engl J Med*. 1994;331(10):637-642. doi:10.1056/NEJM199409083311003
65. Elkins MR, Robinson M, Rose BR, et al; National Hypertonic Saline in Cystic Fibrosis (NHSCF) Study Group. A controlled trial of long-term inhaled hypertonic saline in patients with cystic fibrosis. *N Engl J Med*. 2006;354(3):229-240. doi:10.1056/NEJMoa043900
66. Saiman L, Marshall BC, Mayer-Hamblett N, et al; Macrolide Study Group. Azithromycin in patients with cystic fibrosis chronically infected with *Pseudomonas aeruginosa*: a randomized controlled trial. *JAMA*. 2003;290(13):1749-1756. doi:10.1001/jama.290.13.1749
67. Ivacaftor prescribing information. Vertex Pharmaceuticals Inc. Accessed April 23, 2023. https://pi.vrtx.com/files/uspi_ivacaftor.pdf
68. Ramsey BW, Davies J, McElvaney NG, et al; VX08-770-102 Study Group. A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. *N Engl J Med*. 2011;365(18):1663-1672. doi:10.1056/NEJMoa1105185
69. Davies JC, Cunningham S, Harris WT, et al; KIWI Study Group. Safety, pharmacokinetics, and pharmacodynamics of ivacaftor in patients aged 2-5 years with cystic fibrosis and a CFTR gating mutation (KIWI): an open-label, single-arm study. *Lancet Respir Med*. 2016;4(2):107-115. doi:10.1016/S2213-2600(15)00545-7
70. Rosenfeld M, Wainwright CE, Higgins M, et al; ARRIVAL study group. Ivacaftor treatment of cystic fibrosis in children aged 12 to <24 months and with a CFTR gating mutation (ARRIVAL): a phase 3 single-arm study. *Lancet Respir Med*. 2018;6(7):545-553. doi:10.1016/S2213-2600(18)30202-9
71. Davies JC, Wainwright CE, Sawicki GS, et al. Ivacaftor in infants aged 4 to <12 months with cystic fibrosis and a gating mutation. results of a two-part phase 3 clinical trial. *Am J Respir Crit Care Med*. 2021;203(5):585-593. doi:10.1164/rccm.202008-3177OC
72. Lumacaftor-ivacaftor prescribing information. Vertex Pharmaceuticals Inc. Accessed April 23, 2023. https://pi.vrtx.com/files/uspi_lumacaftor_ivacaftor.pdf
73. Wainwright CE, Elborn JS, Ramsey BW, et al; TRAFFIC and TRANSPORT Study Groups. Lumacaftor-ivacaftor in patients with cystic fibrosis homozygous for Phe508del CFTR. *N Engl J Med*. 2015;373(3):220-231. doi:10.1056/NEJMoa1409547
74. Ratjen F, Hug C, Marigowda G, et al; VX14-809-109 investigator group. Efficacy and safety of lumacaftor and ivacaftor in patients aged 6-11 years with cystic fibrosis homozygous for F508del-CFTR: a randomised, placebo-controlled phase 3 trial. *Lancet Respir Med*. 2017;5(7):557-567. doi:10.1016/S2213-2600(17)30215-1
75. Konstan MW, McKone EF, Moss RB, et al. Assessment of safety and efficacy of long-term treatment with combination lumacaftor and ivacaftor therapy in patients with cystic fibrosis homozygous for the F508del-CFTR mutation (PROGRESS): a phase 3, extension study. *Lancet Respir Med*. 2017;5(2):107-118. doi:10.1016/S2213-2600(16)30427-1
76. Tezacaftor-ivacaftor prescribing information. Vertex Pharmaceuticals Inc. Accessed April 23, 2023. https://pi.vrtx.com/files/uspi_tezacaftor_ivacaftor.pdf
77. Taylor-Cousar JL, Munck A, McKone EF, et al. Tezacaftor-ivacaftor in patients with cystic fibrosis homozygous for Phe508del. *N Engl J Med*. 2017;377(21):2013-2023. doi:10.1056/NEJMoa1709846
78. Elexacaftor-tezacaftor-ivacaftor prescribing information. Vertex Pharmaceuticals Inc. Accessed April 23, 2023. https://pi.vrtx.com/files/uspi_elexacaftor_tezacaftor_ivacaftor.pdf
79. Barry PJ, Mall MA, Álvarez A, et al; VX18-445-104 Study Group. Triple therapy for cystic fibrosis *Phe508del*-gating and -residual function genotypes. *N Engl J Med*. 2021;385(9):815-825. doi:10.1056/NEJMoa2100665
80. Mall MA, Brugha R, Gartner S, et al. Efficacy and safety of elexacaftor/tezacaftor/ivacaftor in children 6 through 11 years of age with cystic fibrosis heterozygous for F508del and a minimal

function mutation: a phase 3b, randomized, placebo-controlled study. *Am J Respir Crit Care Med*. 2022;206(11):1361-1369. doi:10.1164/rccm.202202-0392OC

81. Quittner AL, Buu A, Messer MA, Modi AC, Watrous M. Development and validation of the Cystic Fibrosis Questionnaire in the United States: a health-related quality-of-life measure for cystic fibrosis. *Chest*. 2005;128(4):2347-2354. doi:10.1378/chest.128.4.2347

82. Flume PA, Liou TG, Borowitz DS, et al; VX 08-770-104 Study Group. Ivacaftor in subjects with cystic fibrosis who are homozygous for the *F508del-CFTR* mutation: a multicentre, phase 3, open-label, extension study. *Lancet Respir Med*. 2021;9(9):977-988. doi:10.1016/S2213-2600(21)00069-2

83. Hoppe JE, Chilvers M, Ratjen F, et al. Long-term safety of lumacaftor-ivacaftor in children aged 2-5 years with cystic fibrosis homozygous for the *F508del-CFTR* mutation: an open-label, extension study. *Lancet Respir Med*. 2021;9(9):977-988. doi:10.1016/S2213-2600(21)00069-2

84. Chilvers MA, Davies JC, Milla C, et al. Long-term safety and efficacy of lumacaftor-ivacaftor therapy in children aged 6-11 years with cystic fibrosis homozygous for the *F508del-CFTR* mutation: a phase 3, open-label, extension study. *Lancet Respir Med*. 2021;9(7):721-732. doi:10.1016/S2213-2600(20)30517-8

85. Donaldson SH, Pilewski JM, Griese M, et al; VX11-661-101 Study Group. Tezacaftor/ivacaftor in subjects with cystic fibrosis and *F508del/F508del-CFTR* or *F508del/G551D-CFTR*. *Am J Respir Crit Care Med*. 2018;197(2):214-224. doi:10.1164/rccm.201704-0717OC

86. Boyle MP, Bell SC, Konstan MW, et al; VX09-809-102 study group. A CFTR corrector (lumacaftor) and a CFTR potentiator (ivacaftor) for treatment of patients with cystic fibrosis who have a *phe508del CFTR* mutation: a phase 2 randomised controlled trial. *Lancet Respir Med*. 2014;2(7):527-538. doi:10.1016/S2213-2600(14)70132-8

87. Veit G, Roldan A, Hancock MA, et al. Allosteric folding correction of *F508del* and rare *CFTR* mutants by elxacaftor-tezacaftor-ivacaftor (Trikafta) combination. *JCI Insight*. 2020;5(18):e139983. doi:10.1172/jci.insight.139983

88. Keating D, Marigowda G, Burr L, et al; VX16-445-001 Study Group. VX-445-tezacaftor-ivacaftor in patients with cystic fibrosis and one or two *Phe508del* alleles. *N Engl J Med*. 2018;379(17):1612-1620. doi:10.1056/NEJMoa1807120

89. Griese M, Tullis E, Chilvers M, et al. Long-term safety and efficacy of elxacaftor/tezacaftor/ivacaftor in people with cystic fibrosis and at least one *F508del* allele: 144-week interim results from an open-label extension study. *J Cyst Fibros*. 2022;21:S99-S100. doi:10.1016/S1569-1993(22)00861-X

90. Zemanick ET, Taylor-Cousar JL, Davies J, et al. A phase 3 open-label study of elxacaftor/tezacaftor/ivacaftor in children 6 through 11 years of age with cystic fibrosis and at least one *F508del* allele. *Am J Respir Crit Care Med*. 2021;203(12):1522-1532. doi:10.1164/rccm.202102-0509OC

91. Goralski JL, Hoppe JE, Mall MA, et al; VX20-445-111 Study Group. Phase 3 open-label

clinical trial of elxacaftor/tezacaftor/ivacaftor in children aged 2 through 5 years with cystic fibrosis and at least one *F508del* allele. *Am J Respir Crit Care Med*. 2023. doi:10.1164/rccm.202301-0084OC

92. Clancy JP, Cotton CU, Donaldson SH, et al. CFTR modulator therotyping: current status, gaps and future directions. *J Cyst Fibros*. 2019;18(1):22-34. doi:10.1016/j.jcf.2018.05.004

93. McNamara JJ, McColley SA, Marigowda G, et al. Safety, pharmacokinetics, and pharmacodynamics of lumacaftor and ivacaftor combination therapy in children aged 2-5 years with cystic fibrosis homozygous for *F508del-CFTR*: an open-label phase 3 study. *Lancet Respir Med*. 2019;7(4):325-335. doi:10.1016/S2213-2600(18)30460-0

94. Dagenais RVE, Su VCH, Quon BS. Real-world safety of CFTR modulators in the treatment of cystic fibrosis: a systematic review. *J Clin Med*. 2020;10(1):23. doi:10.3390/jcm10010023

95. Scotet V, L'Hostis C, Férec C. The changing epidemiology of cystic fibrosis: incidence, survival and impact of the *CFTR* gene discovery. *Genes (Basel)*. 2020;11(6):589. doi:10.3390/genes11060589

96. Leung DH, Heltshe SL, Borowitz D, et al; Baby Observational and Nutrition Study (BONUS) Investigators of the Cystic Fibrosis Foundation Therapeutics Development Network. Effects of diagnosis by newborn screening for cystic fibrosis on weight and length in the first year of life. *JAMA Pediatr*. 2017;171(6):546-554. doi:10.1001/jamapediatrics.2017.0206

97. Rosenfeld M, Ostrenga J, Cromwell EA, et al. Real-world associations of US cystic fibrosis newborn screening programs with nutritional and pulmonary outcomes. *JAMA Pediatr*. 2022;176(10):990-999. doi:10.1001/jamapediatrics.2022.2674

98. Volkova N, Moy K, Evans J, et al. Disease progression in patients with cystic fibrosis treated with ivacaftor: data from national US and UK registries. *J Cyst Fibros*. 2020;19(1):68-79. doi:10.1016/j.jcf.2019.05.015

99. Nichols DP, Paynter AC, Heltshe SL, et al; PROMISE Study group. Clinical effectiveness of elxacaftor/tezacaftor/ivacaftor in people with cystic fibrosis: a clinical trial. *Am J Respir Crit Care Med*. 2022;205(5):529-539. doi:10.1164/rccm.202108-1986OC

100. Burgel PR, Durieu I, Chiron R, et al; French Cystic Fibrosis Reference Network Study Group. Rapid improvement after starting elxacaftor-tezacaftor-ivacaftor in patients with cystic fibrosis and advanced pulmonary disease. *Am J Respir Crit Care Med*. 2021;204(1):64-73. doi:10.1164/rccm.202011-4153OC

101. Buu MC, Sanders LM, Mayo JA, Milla CE, Wise PH. Assessing differences in mortality rates and risk factors between Hispanic and non-Hispanic patients with cystic fibrosis in California. *Chest*. 2016;149(2):380-389. doi:10.1378/chest.14-2189

102. McColley SA, Martiniano SL, Ren CL, et al. Disparities in first evaluation of infants with cystic fibrosis since implementation of newborn screening. *J Cyst Fibros*. 2023;22(1):89-97. doi:10.1016/j.jcf.2022.07.010

103. McGarry ME, Ren CL, Wu R, Farrell PM, McColley SA. Detection of disease-causing *CFTR*

variants in state newborn screening programs. *Pediatr Pulmonol*. 2023;58(2):465-474. doi:10.1002/ppul.26209

104. McGarry ME, McColley SA. Cystic fibrosis patients of minority race and ethnicity less likely eligible for CFTR modulators based on *CFTR* genotype. *Pediatr Pulmonol*. 2021;56(6):1496-1503. doi:10.1002/ppul.25285

105. Mayer-Hamblett N, Ratjen F, Russell R, et al. Discontinuation versus continuation of hypertonic saline or dornase alfa in modulator treated people with cystic fibrosis (SIMPLIFY): results from two parallel, multicentre, open-label, randomised, controlled, non-inferiority trials. *Lancet Respir Med*. 2022. doi:10.1016/S2213-2600(22)00434-9

106. Tice JA, Kuntz KM, Wherry K, et al. Modulator treatments for cystic fibrosis: effectiveness and value: final evidence report and meeting summary. Institute of Economic Review. September 23, 2020. Accessed April 23, 2023. https://icer.org/wp-content/uploads/2020/08/ICER_CF_Final_Report_092320.pdf

107. Zampoli M, Morrow BM, Paul G. Real-world disparities and ethical considerations with access to CFTR modulator drugs: mind the gap! *Front Pharmacol*. 2023;14:1163391. doi:10.3389/fphar.2023.1163391

108. Jain R, Magaret A, Vu PT, et al. Prospectively evaluating maternal and fetal outcomes in the era of CFTR modulators: the MAYFLOWERS observational clinical trial study design. *BMJ Open Respir Res*. 2022;9(1):e001289. doi:10.1136/bmjresp-2022-001289

109. Bailey J, Krick S, Fontaine KR. The changing landscape of nutrition in cystic fibrosis: the emergence of overweight and obesity. *Nutrients*. 2022;14(6):1216. doi:10.3390/nu14061216

110. Jain R, Kazmerski TM, Zuckerwise LC, et al. Pregnancy in cystic fibrosis: review of the literature and expert recommendations. *J Cyst Fibros*. 2022;21(3):387-395. doi:10.1016/j.jcf.2021.07.019

111. Schechter MS, Ostrenga JS, Fink AK, Barker DH, Sawicki GS, Quittner AL. Decreased survival in cystic fibrosis patients with a positive screen for depression. *J Cyst Fibros*. 2021;20(1):120-126. doi:10.1016/j.jcf.2020.07.020

112. Quittner AL, Goldbeck L, Abbott J, et al. Prevalence of depression and anxiety in patients with cystic fibrosis and parent caregivers: results of The International Depression Epidemiological Study across nine countries. *Thorax*. 2014;69(12):1090-1097. doi:10.1136/thoraxjnl-2014-205983

113. Egan ME. Non-modulator therapies: developing a therapy for every cystic fibrosis patient. *Clin Chest Med*. 2022;43(4):717-725. doi:10.1016/j.ccm.2022.06.011

114. Yan Z, McCray PB Jr, Engelhardt JF. Advances in gene therapy for cystic fibrosis lung disease. *Hum Mol Genet*. 2019;28(R1):R88-R94. doi:10.1093/hmg/ddz139

115. Schwank G, Koo BK, Sasselli V, et al. Functional repair of *CFTR* by CRISPR/Cas9 in intestinal stem cell organoids of cystic fibrosis patients. *Cell Stem Cell*. 2013;13(6):653-658. doi:10.1016/j.stem.2013.11.002